Figure S1. **Gating strategy for the 10-color 13-antibody panel OMIP-023** using a sample of one subject. EDTA-anticoagulated whole blood was pre-lysed to remove CD45- erythrocytes and to enrich CD45+ leukocytes. The stained peripheral blood sample was prepared for analysis by excluding air bubbles (time *versus* sideward scatter, plot 1) followed by gating on single-cell events (exclusion of non-single events outside the gate) in plot 2 (time of flight forward scatter *versus* forward scatter) before gating in plot 3 (CD45+ leukocytes) on the three major subsets, granulocytes (A, SSC_{high}), monocytes (B, SSC_{med}) and lymphocytes (C, SSC_{low}) for further analyses (see Figure S2; plot 3 shown here corresponds to plot 1 in Figure S2). Further details can be found in Methods* and in [15] and [16].


According to SSC height, gates of granulocytes (SSC$^{\text{high}}$; plot 2), monocytes (SSC$^{\text{med}}$; plots 3 to 5), and lymphocytes (SSC$^{\text{low}}$; plots 6 to 11) were discriminated and further subdivided. Neutrophils (plot 2: CD16$^+$) and eosinophils (plot 2: CD16) in granulocytes were discriminated. After excluding CD14 HLA-DR$^+$ events (plot 3) and CD4$^+$ events (plot 4) from monocyte analysis, classical (typical) monocytes (plot 5: CD14$^+$CD16$^+$) and nonclassical monocytes (plot 5: atypical [CD14$^{\text{dim}}$CD16$^+$] and intermediate [CD14$^+$CD16$^+$]) were discriminated as well. Lymphocytes (plot 6) were gated into CD3$^-$ (left; after exclusion of CD4$^+$ events [plot 7]) further analyzed in plot 8A: CD16/56$^+$ NK cells; plot 8B: CD16/56 B-lymphocytes and CD3$^+$ events (right; further analyzed in 9A: CD3$^+$CD16/56$^+$ NKT cells and 9B: CD3$^+$CD16/56$^+$ T-lymphocytes). Three T-lymphocyte subsets were differentiated (plot 10): CD8$^+$ cytotoxic T cells (Tc), CD4$^+$ T-helper cells (Th) and CD4$^+$CD8$^+$ double positive T-cells (DPT). The gated T-helper cells were also used to identify CD25$^+$ regulatory T-cells (Treg: anti-CD127 versus anti-CD25, plot 11). Further details can be found in Methods* and in [15] and [16].

* According to SSC height, gates of granulocytes (plot 1 A: CD45$^+$SSC$^{\text{high}}$), monocytes (B: CD45$^+$SSC$^{\text{med}}$) and lymphocytes (C: CD45$^+$SSC$^{\text{low}}$) were discriminated and further subdivided. Neutrophils (plot 2: CD16$^+$) and eosinophils (plot 2: CD16) in granulocytes were discriminated. After excluding CD14 HLA-DR$^+$ events (plot 3) and CD4$^+$ events (plot 4) from monocyte analysis, classical (typical) monocytes (plot 5: CD14$^+$CD16$^+$) and nonclassical monocytes (plot 5: atypical [CD14$^{\text{dim}}$CD16$^+$] and intermediate [CD14$^+$CD16$^+$]) were discriminated as well. Lymphocytes (plot 6) were gated into CD3$^-$ (left; after exclusion of CD4$^+$ events [plot 7]) further analyzed in plot 8A: CD16/56$^+$ NK cells; plot 8B: CD16/56 B-lymphocytes and CD3$^+$ events (right; further analyzed in 9A: CD3$^+$CD16/56$^+$ NKT cells and 9B: CD3$^+$CD16/56$^+$ T-lymphocytes). Three T-lymphocyte subsets were differentiated (plot 10): CD8$^+$ cytotoxic T cells (Tc), CD4$^+$ T-helper cells (Th) and CD4$^+$CD8$^+$ double positive T-cells (DPT). The gated T-helper cells were also used to identify CD25$^+$ regulatory T-cells (Treg: anti-CD127 versus anti-CD25, plot 11). Further details can be found in [15] and [16].