**GLUL Ablation Can Confer Drug Resistance to Cancer Cells via a Malate-Aspartate Shuttle-Mediated Mechanism**

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Supplementary information:

**Figure 1.** GLUL KO/KD induces drug resistance. (A) Western blot shows screening of CRISPR/Cas9 mediated GLUL KO clones. (B,C) A549 cells either transfected with non-target control or with doxycycline inducible shRNA (1, 2, 3) targeting GLUL. Subsequently cell lysates were analysed by western blot (WB) as in methods for GLUL expression levels and individual clones were screened and selected as shown above. The western blot membranes were subsequently probed with anti-tubulin antibody to assess equal loading. The presence of GLUL protein is indicated in the right side of each blot. The signals for the GLUL and α-tubulin proteins were quantified by densitometry, and the numbers below GLUL blot indicate levels of GLUL protein in each lane following normalization of the signals with actin levels. For the sake of comparison, the signal intensity for GLUL in control lane was assigned an arbitrary value of 1. Approximate location of various molecular weight is indicated on left side of each blot. kDa, kilodalton. (D) Upon doxycycline induction, A549 GLUL shRNA3 cl3...
cells were treated with respective drugs of various concentrations as indicated and cell viability were
assessed by MTS assay after 72 h. Further, IC₅₀ were determined as indicated. The standard error (SE)
bars in cell viability curve represent means of three independent experiments.

![Figure 2](image)

**Figure 2.** KO cells display drug resistance in A549 but not in H1299 cells. (A,D) A549 & H1299 cells
were either transfected with negative control or with LentiviralCRISPR/Cas9 coupled GLUL guide RNA.
Subsequently cells were treated with indicated chemotherapeutic drugs and cell viability was
analysed by MTS assay after 72–96 h. IC₅₀ for each drug was determined as indicated. (B,E) A549 &
H1299 control and GLUL KO cells were treated with the indicated drugs and subjected to clonogenic
assay. Histograms represent total number of colonies counted and representative images of crystal
violet stained cells are shown. The standard error (SE) bars in cell viability assay and clonogenic assay
represent means of three independent experiments. (C) H1299 cells were either transfected with
negative control or with LentiviralCRISPR/Cas9 coupled GLUL guide RNA. Subsequently cell lysates
were analysed by western blot (WB) as in methods for GLUL expression levels and individual clones
were screened and selected as shown above. The western blot membranes were subsequently probed
with anti-tubulin antibody to assess equal loading. The presence of GLUL and tubulin protein is
indicated in the right side of each blot. The presence of GLUL protein is indicated in the right side of each blot. The signals for the GLUL and α-tubulin proteins were quantified by densitometry, and the numbers below GLUL blot indicate levels of GLUL protein in each lane following normalization of the signals with actin levels. For the sake of comparison, the signal intensity for GLUL in control lane was assigned an arbitrary value of 1. Approximate location of various molecular weight is indicated on left side of each blot. kDa, kilodalton. The data shown as mean ± SEM; p-values were determined using a two-tailed unpaired t-test; ** \( p \leq 0.004 \), * \( p \leq 0.01 \), ns – not significant.

**Figure 3.** A549 GLUL KD exhibit less or no PARP cleavage. A549 Control (Non-target) or GLUL KD (GLUL shRNA3-cl3) cells were treated with doxycycline at a concentration of 5 µg/mL for 72 h and then were either treated with DMSO (control) or treated with chemotherapeutic drugs as mentioned for 12 h. Protein levels of cleaved PARP were assessed by western blot analysis. Subsequently, the membranes were probed with anti-tubulin antibody to assess equal loading. The presence of GLUL and tubulin protein is indicated in the right side of each blot. Approximate location of various molecular weight is indicated on left side of each blot. Quantification of protein levels is presented on right side of each western blot. kDa, kilodalton.
Figure 4. Reduced glutamine do not correlates with drug resistance in GLUL KO cells. A549 & H1299 (Control and GLUL KO) cells were cultured in various concentrations glutamine as indicated for 72 h and cell viability was analysed by MTS assay. Data for each pair of cell lines (Control/GLUL KO) was normalized to the data for the highest concentration of glutamine (4 mM) as shown above. Data shown as ± standard deviation. The standard deviation (SD) bars in cell viability curve represent means of three independent experiments.

Figure 5. Targeted metabolomics reveals glutamine accumulation in resistant cells without increased glutamine uptake or flux through the glutaminolytic pathway. (A) Fold change for metabolites that displayed significant difference between GLUL KO/Control A549 drug resistant cells. (B) Fold change for metabolites that displayed significant difference between GLUL KO/Control H1299 drug sensitive cells. (C) Left, relative quantitation (area units) for glutamine (GLN), glutamate (GLU) and α-ketoglutarate (α-KG). Right, %-fractional labeling obtained through feeding 13C5-glutamine to the cells. Data shown as ± standard deviation; p-values were determined using a two-tailed unpaired t-test; *** p ≤ 0.001; ** p ≤ 0.01.
Figure 6. Reduced GLUL expression induces glutaminolysis. A549 Control (Non-target) or GLUL KD (GLUL shRNA3 c3) cells were treated with doxycycline at a concentration of 5 μg/mL for 72 h and various Protein expressions were assessed by western blot analysis as shown above. Subsequently, the membranes were probed with anti-tubulin antibody to assess equal loading. The presence of various proteins is indicated in the right side of each blot and approximate location of various molecular weight is indicated on left side of each blot. kDa, kilodalton.

Figure 7. %-Fractional labelling of glutaminolysis intermediates. Glutamine (GLN), Glutamate (GLU) and alpha-ketoglutarate (α-KG). Glutamate dehydrogenase (GLUD), Glutaminase (GLS), Glutamate-ammonia ligase (GLUL).
Figure 8. H1299 KO cells do not display sensitivity to ATT inhibition. H1299 control and GLUL KO cl3 (Clone 2) cells were either treated with DMSO or treated with inhibitor Aminoxyacetic acid of various concentration and cell viability were analysed by MTS assay after 96 h. Further, IC₅₀ were determined as indicated in method section (260 µM). The standard error (SE) bars in cell viability curve represent means of three independent experiments.

Figure 9. Reduced GLUL mRNA expression correlates with poor prognosis in breast and lung cancer patients. The kaplan-Meier Plotter database (http://kmplot.com) was quereid using all default settings in breast cancer and lung cancer patient cohorts containing expression of mRNA and survival data. Black line shows survival for patients with low expression of GLUL. Low expression is strongly correlating with poorer survival (p values top right corners of both panels).

Raw blots:
Figure 2A

[Image of a scientific diagram with labels and annotations, possibly showing data or experimental results.]
Figure 3
Figure 3
Figure 6C
Suppl. Figures:
Figure S1A

Figure S1B
Figure S3
Figure S3
Figure S6