

Article

First Detection of Tetrodotoxins in the Cotylean Flatworm *Prosthiosomum trilineatum*

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Abstract: Several polyclad flatworm species are known to contain high levels of tetrodotoxin (TTX), but currently TTX-bearing flatworms seem to be restricted to specific *Planocera* lineages belonging to the suborder Acotylea. During our ongoing study of flatworm toxins, high concentrations of TTXs were detected for the first time in the flatworm *Prosthiosomum trilineatum*, suborder Cotylea, from the coastal area of Hayama, Kanagawa, Japan. Toxin levels were investigated by high performance liquid chromatography-tandem mass spectrometry (LC-MS/MS), revealing that this species contains comparable concentrations of toxins as seen in planoceric flatworms such as *Planocera multitentaculata*. This finding indicated that there may be other species with significant levels of TTXs. The distribution of TTXs among other flatworm species is thus of great interest.



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1. Introduction

Tetrodotoxin (TTX), a well-known potent neurotoxin first isolated from pufferfish [1–3], has been discovered in a wide range of marine and terrestrial organisms, such as amphibians, flatworms, crustaceans, mollusks, and several species of marine bacteria [4–6]. TTX and its analogs are grouped with sodium channel blockers, preventing sodium ion influx through sodium channels in an excitable membrane, resulting in serious toxicity [7–9]. The toxicity of TTX-bearing organisms depends upon geography and season. In addition, the wide distribution of TTX across phylogenetically diverse organisms has led to the hypothesis of a bacterial origin of TTX, either from accumulation through the food chain or from symbiotic relationships with TTX-bearing organisms [5,10–12]. However, the origin of TTX remains controversial, and the biosynthetic pathway remains unclear [13,14].

Some marine polyclad flatworms are known to contain high concentrations of TTX and are thought to be one of the animals that might be responsible for the presence of TTX in some marine organisms. The sea slug *Pleurobranchaea maculata*, the organisms ascribable to the neurotoxicosis of dogs in some beaches of Auckland, New Zealand, are reportedly toxicated by feeding on the TTX-bearing flatworm *Stylochoplana* sp. [15–17]. Additionally, we have investigated the ecological function of *Planocera multitentaculata*, inhabiting the coastal area of Hayama, Kanagawa, Japan and have confirmed the following: the DNA sequence of the flatworm *Pl. multitentaculata* was detected in the intestinal contents of juveniles and young of the pufferfish *Takifugu niphobles* (at present *Takifugu alboplumbeus*); the pufferfish at various growth stages effectively toxicated itself by feeding on the flatworm *Pl. multitentaculata* and its eggs; and, non-toxic pufferfish became toxicated after feeding on the flatworm *Pl. multitentaculata* [18,19]. Moreover, Next Generation Sequencing

(NGS) analysis of the cytochrome *c* oxidase subunit I (COI) gene of the intestinal contents from juveniles of TTX-bearing organisms demonstrated that the majority of the sequence is identical to that of *Pl. multitentaculata* [20]. These results indicate that the flatworm contributes to the toxification of pufferfish at various life stages and functions as one of the major TTX sources for other TTX-bearing fish species. This leads to great interest in finding out the distribution of TTXs among other flatworm species.

The polyclad flatworms have been traditionally divided into two suborders, Cotylea and Acotylea, which are distinguished by the presence or absence of a ventral sucker behind the genital openings [21–23]. They are globally distributed in the marine environment, found under coastal rocks and in interstitial spaces. Currently, the toxins have been detected within the suborder Acotylea, such as *Pl. multitentaculata*, *Planocera reticulata* and *Stylochoplana* sp., but toxification by TTX is still undetected within the suborder Cotylea [16,17,24–27]. However, our ongoing research on toxins from marine polyclad flatworms has now detected a high concentration of TTX and its analog from the cotylean flatworm, *Prosthiosomum trilineatum*, found in a coastal area of Hayama, Kanagawa, Japan. Herein, we report the first detection of TTXs within this suborder and discuss its habitat, relationship with other marine organisms and potential adverse human health risk.

2. Results and Discussion

2.1. External Morphology and Molecular Phylogenetic Inference of the Flatworm *Pr. trilineatum*

Two flatworm specimens were collected from the coastal area of Hayama, Kanagawa, Japan under stones in the intertidal zone, and a rocky bottom at a depth of approximately 10 m. These were identified as *Pr. trilineatum* based on the description of external morphology [28,29]; broadly elongated body, rounded anteriorly and tapered posteriorly; longitudinal orange-yellow stripe bordered by black band running medially (Figure 1).

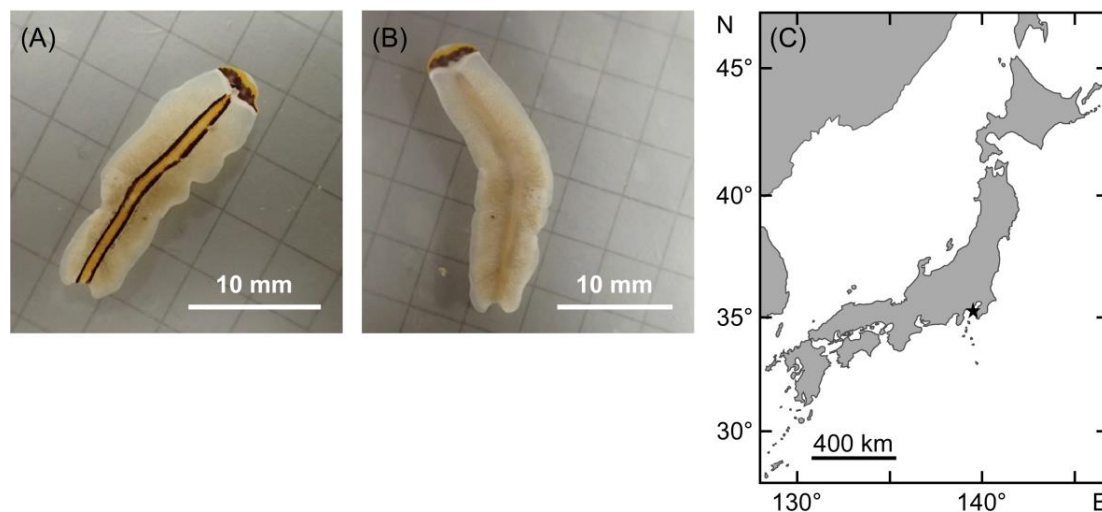


Figure 1. External morphology of *Prosthiosomum trilineatum* collected in this study. (A,B) represent dorsal and ventral views of specimen 1, respectively. (C) represents map of sampling location (marked by a star).

Additionally, partial sequences of the 28S rRNA (approximately 1100 bp) were obtained from both specimens, and the resulting sequences showed high similarity with those of related cotylean flatworm species, *Prosthiosomum vulgare* for 94.4% and *Prosthiosomum siphuncululus* for 92.4%. Maximum likelihood analysis based on 28S rRNA sequences from various polyclad flatworms revealed that the *Pr. trilineatum* specimens in this study formed a cluster containing sequences from other *Prosthiosomum* species in the suborder Cotylea (Figure 2). The DNA sequences of the 28S rRNA gene fragment from this study have been submitted to the DDBJ/EMBL/GenBank databases.

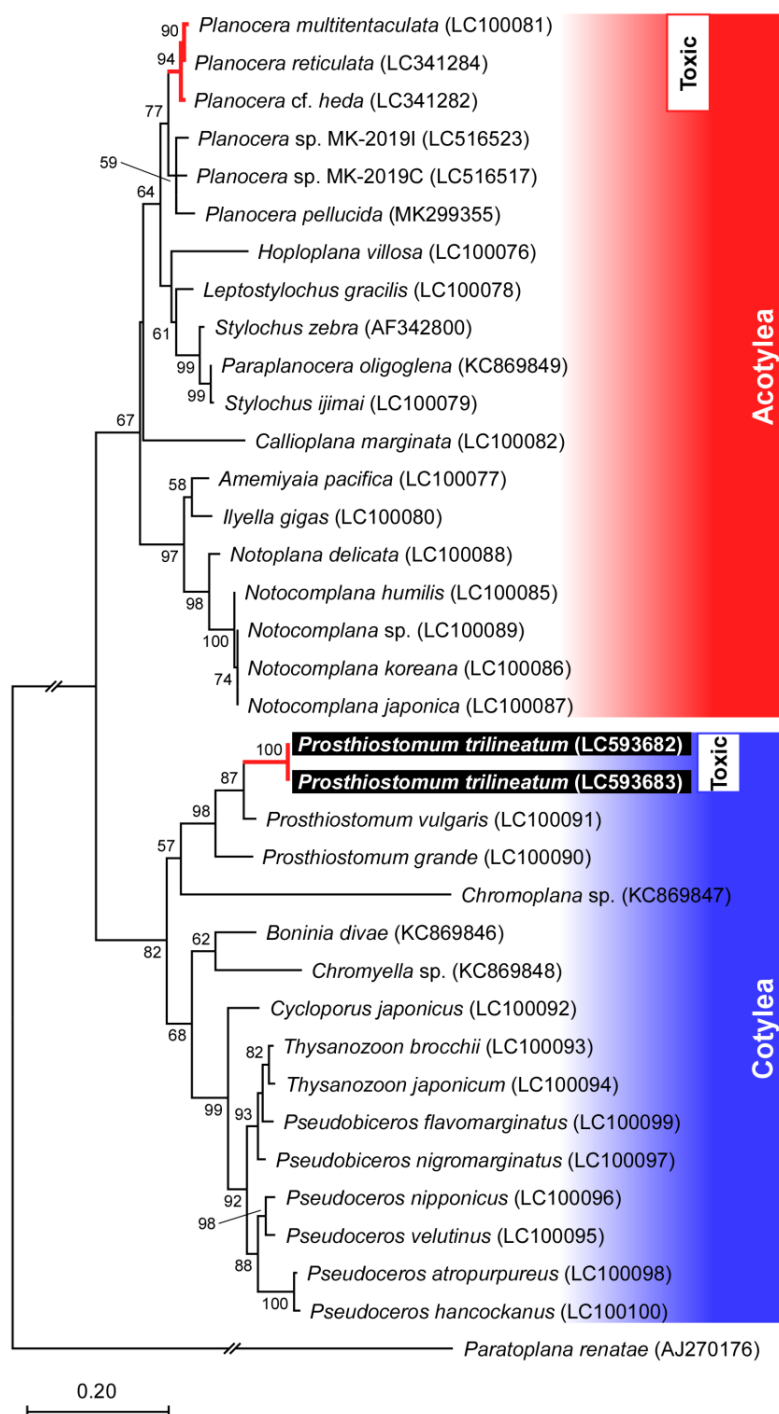


Figure 2. Phylogenetic position of *Prosthiostomum trilineatum* among polyclad species inferred from the 28S rRNA gene sequence. The phylogenetic tree was constructed by maximum likelihood analysis. Numbers at branches denote the bootstrap percentages from 1000 replicates. The accession numbers for the sequences are shown in parentheses. The accession numbers LC593682 and LC593683 refer to those deposited in the DDBJ/EMBL/GenBank databases in this study. The sequence from *Paratoplana renatae* was used as the outgroup. Only bootstrap values exceeding 50% are presented. The scale refers to nucleotide substitutions per site.

2.2. TTXs Analysis

It is well-known that the TTX-bearing organisms, such as pufferfish and newt, contain 4-*epi*TTX and 4,9-anhydroTTX, which are the chemically equilibrium analogs of TTX [30].

Additionally, several non-equilibrium TTX analogs with different oxidation states and stereochemistries have been isolated [31–38]. In our study, we focused on the detection of the major components, TTX and its analog 5,6,11-trideoxyTTX. The compound mixture containing TTX and 5,6,11-trideoxyTTX was used as the standard, and detected at m/z 320 > 302 for qualification and m/z 320 > 162 for quantification of TTX, and at m/z 272 > 254 for qualification and m/z 272 > 162 for quantification of 5,6,11-trideoxyTTX. The amount of TTXs in the two individual flatworms was calculated by measuring extracts with appropriate dilution, respectively. High concentrations of TTX and 5,6,11-trideoxyTTX were detected from both extracts of the specimens. The total amount of TTX in the body of specimen 1 and 2 was calculated at 223 $\mu\text{g}/\text{individual}$ and 8.4 $\mu\text{g}/\text{individual}$, respectively, whereas 5,6,11-trideoxyTTX was 170 $\mu\text{g}/\text{individual}$ and 6.2 $\mu\text{g}/\text{individual}$, respectively (Figure 3, Table 1).

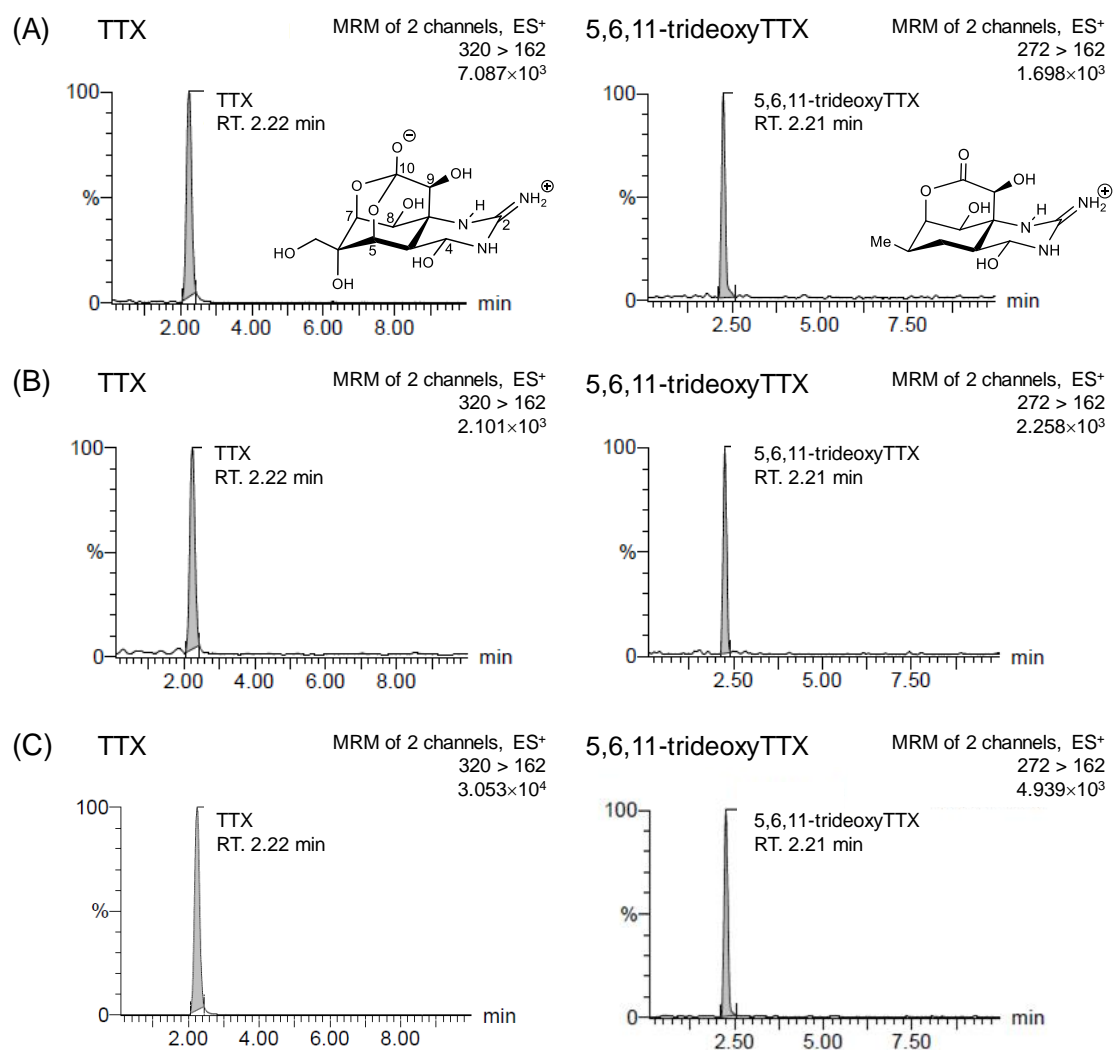


Figure 3. Liquid chromatography-tandem mass spectrometry (LC-MS/MS) chromatograms with multiple reaction monitoring mode (TTX: m/z 320 > 162, 5,6,11-trideoxyTTX: m/z 272 > 162). (A) Standards of TTXs, (B) the extracts from *Prosthiostomum trilineatum* specimen 1, (C) the extracts from *Pr. trilineatum* specimen 2. The concentration of the standard is shown as below: TTX 50 ng/mL, 5,6,11-trideoxyTTX 50 ng/mL.

Table 1. The contents of TTXs in *Pr. trilineatum*.

Compounds	Content ($\mu\text{g}/\text{individual}$)	
	Specimen 1	Specimen 2
TTX	223	8.4
5,6,11-trideoxyTTX	170	6.2

So far, TTXs have been detected from the acotylean flatworms *Pl. multitentaculata*, *Pl. reticulata*, *Stylochoplana* sp., and *Planocera* sp. in significant levels [16,17,24–27]. Previously, we have investigated the toxicity of the acotylean flatworms *Pl. multitentaculata* ($n = 122$) and *Pl. reticulata* ($n = 21$), and found that all individuals contained TTXs [39–41]. Based on the phylogenetic tree of polyclad flatworms and toxin detection data as reported so far, TTX-bearing flatworm species seem to be restricted to specific *Planocera* lineages in the suborder Acotylea [41] (Figure 2). However, the data present in this study raise the intriguing possibility that the distribution of TTXs among marine polyclad flatworms might be more diverse than expected. Although TTX contents vary between the individuals, and the number of samples ($n = 2$) in this study is limited, both *Pr. trilineatum* specimens contained levels of TTXs comparable to those of other planocerid flatworms [26,39,40]. As far as we know, in the last 10 years, the cotylean flatworm *Pr. trilineatum* has rarely been observed in the intertidal zone of Hayama. Thus, in this study, only two individuals were subjected to the TTXs analysis due to the difficulty of sample collection. Further study will be required to confirm whether the toxins are constantly or variably present in *Pr. trilineatum*. Besides, a similar restricted taxonomic distribution of TTX might also be seen in the *Prosthiostomum* and related lineages; therefore, a wider and more intensive search of toxins distribution among cotylean flatworms, especially among the genus *Prosthiostomum*, is necessary.

2.3. *Prosthiostomum trilineatum* and Possible Relationship with Other Marine Organisms

The distribution of toxins among individual *Pr. trilineatum* or its presence at specific geographical areas have not been investigated. The distribution of *Pr. trilineatum* is quite wide, extending from the North Equatorial Current to the Kuroshio Current including Japan, Micronesia, Guam, and Papua New Guinea, and even to Singapore and India [28,29,42,43], suggesting that the sampling site for this study may be the northern limit in the Kuroshio basin. If so, *Pr. trilineatum* may be more abundant at the headwaters of the Kuroshio Current. Based on our study and the other reports regarding the collection of *Pr. trilineatum*, we deduce that *Pr. trilineatum* probably inhabits the deeper water (≥ 10 m), which is deeper than our collection sites. However, the biology and ecology of *Pr. trilineatum* are unstudied, and neither the spawning season nor the amount of biomass around Hayama are clear. Further studies to identify *Pr. trilineatum* habitat and toxin distribution are needed.

In the current study, flatworms were collected from the coastal area of Hayama, close to the geographical distribution of *Pl. multitentaculata* and *Pl. reticulata*. Our previous report showed that the pufferfish *T. alboplumbeus* inhabiting an intertidal zone of Hayama effectively acquired the toxins by feeding on *Pl. multitentaculata*, and its eggs and planktonic larvae [19,44]. Although the biomass of *Pr. trilineatum* seems to be limited compared with that of *Pl. multitentaculata* in the waters around Hayama area, there is a possibility that *Pr. trilineatum* also contributes to the toxification of pufferfish.

Generally, flatworms are considered carnivores that feed on immobile animals such as ascidians, mollusks, and bryozoans [45]. The origin of TTX in pufferfish, as well as in flatworms, is still unresolved, although TTX is hypothesized to be biosynthesized by bacteria and bio-accumulated through the food chain [5,12]. Therefore, it is important to investigate how the flatworms acquired the toxins, and why they possess highly concentrated toxins. In addition, it is necessary to investigate the effect of flatworms on the toxification of edible marine organisms, which are traditionally thought of as non-toxic.

Recently, there has been a problematic world-wide increase in TTXs detection in edible fish, bivalves, and gastropods. TTXs have spread rapidly in the Mediterranean with levels high enough to cause food poisoning [46–48]. Additionally, TTXs have been detected in high concentrations in over ten marine bivalve and gastropod species in the Pacific and Mediterranean [49–54]. However, many aspects regarding the origin of TTX in shellfish as well as pufferfish are unclear, accelerating the interest surrounding TTX in these organisms. In our previous research, we detected high percentages of the toxic planoceric DNA sequence in the gut content of TTX-bearing fishes, and we suggested that planoceric flatworm larvae are involved in toxification process of TTX-bearing marine fishes around the Sakishima Islands, and Hayama, Kanagawa, Japan, [19,20]. Although there is no direct evidence to show that flatworms are the origin of toxified edible marine organisms, more studies, such as information about the relationship between feeding habits of TTX-bearing organisms and flatworms or other planktonic organisms, will provide evidence on the origin of TTX in marine habitats. This will ultimately lead to better protection of human health.

3. Materials and Methods

3.1. Flatworm Specimens

Flatworms were collected under stones in the intertidal zone (specimen 1), and a rocky bottom at a depth of approximately 10 m (specimen 2) from the coastal area of Hayama, Kanagawa, Japan (35°15' N, 139°34' E). The mass of the specimens could not be recorded due to their very small size. Each individual was photographed and then stored at −20 °C until DNA analysis and TTX extraction.

3.2. Preparation of Sample Solutions of the Flatworm for LC-MS Analysis

The flatworms after a small portion was excised were subjected to TTXs extraction in 1 mL of 0.1% acetic acid by heating for 10 min in boiling water. The extracts were centrifuged for 20 min at 14,000× *g*; then the supernatants were filtered through a membrane of pore size 0.45 μm (SupraPure Syringe Filter, PTFE-Hydrophilic, Recentec, Taipei, Taiwan) and subjected to liquid chromatography-tandem mass spectrometry (LC-MS/MS) analysis.

3.3. LC-MS/MS Analysis

LC-MS/MS with multiple reaction monitoring (MRM) mode was performed using a Quattro Premier XE (Waters, Milford, MA, USA) with an electrospray ionization (ESI) source coupled to an Acquity UPLC system (Waters). The sample solutions were analyzed by LC-MS/MS at a flow rate of 0.3 mL/min on an Atlantis HILIC Silica column (2.1 mm × 150 mm, 5 μm, Waters) at 40 °C. Gradient elution was performed using solvent A (H₂O containing 0.1% formic acid) and solvent B (MeCN) with the following linear gradient combination: 95% (B), decreasing to 40% (B) after 0.1 min, then kept for 7.9 min, and back to 95% (B), and finally kept at 95% (B) for 2.0 min before the next injection [15]. In advance of sample analysis, the standard calibration curve was created using 1 to 100 ng/mL of TTXs standards (TTX and 5,6,11-trideoxyTTX [55]) which showed good linearity and precision (TTX: $y = 83.6085x + 708.342$, $R^2 = 0.96$; 5,6,11-trideoxyTTX: $y = 18.8439x - 2.37614$, $R^2 = 0.99$). The optimized transitions for TTX were m/z 320 > 302 for confirmation and m/z 320 > 162 for quantification, and those for 5,6,11-trideoxyTTX were m/z 272 > 254 for confirmation and m/z 272 > 162 for quantification, as shown below (Table 2).

Table 2. MRM transitions used for detection and quantification of TTXs.

Compounds	Transition	Cone (V)	CE (V)
TTX	320 > 302	42	24
	320 > 162	42	38
5,6,11-trideoxyTTX	272 > 254	46	19
	272 > 162	46	28

3.4. Molecular Analysis

Total genomic DNA was extracted from the excised tissue using standard phenol–chloroform extraction with some modification [56]. The 28S rRNA genes were PCR-amplified from the extracted DNA using the primers, HRNT-F2 (5'-AGTTC AAGAG TACGT GAAAC C-3') and HRNT-R2 (5'-AACAC CTTTT GTGGT ATCTG ATGA-3') [56]. The PCR reaction mixture contained 1 µL of DNA template, 2 µL of 10 × buffer for Ex Taq, 1.6 µL of dNTPs, 2.6 µL of 5 mM of each primer, 0.625 unit of Ex Taq (Takara Bio, Shiga, Japan), and H₂O for a total volume of 20 µL. The PCR reaction was performed on a T100 Thermal Cycler (Bio-Rad) as follows: initial denaturation at 95 °C for 1 min, 35 cycles of denaturation 95 °C for 10 s, annealing at 55 °C for 30 s, and a final elongation for 1 min at 72 °C. The PCR products were electrophoresed on 2% agarose gel in 1 × TAE buffer, stained with ethidium bromide, and purified followed by direct sequencing. The resulting strands were sequenced with a 3130xl genetic analyzer (Applied Biosystems, Foster City, CA, USA) using a BigDye Terminator v3.1 Cycle Sequencing Ready Reaction Kit (Applied Biosystems). The sequences of the 28S rRNA gene were submitted to the DDBJ/EMBL/GenBank databases under the accession number LC593682–LC593683. Then, 28S rRNA gene sequences for the following species were obtained from the DDBJ/EMBL/GenBank databases: *Stylochus zebra* (AF342800), *Boninia divae* (KC869846), *Chromoplana* sp. (KC869847), *Chromyella* sp. (KC869848), *Paraplanocera oligoglana* (KC869849), *Planocera pellucida* (MK299355), *Amemiyaia pacifica* (LC100077), *Leptostylochus gracilis* (LC100078), *Stylochus ijimai* (LC100079), *Discoplana gigas* (LC100080), *Pl. multitentaculata* (LC100081), *Callioplana marginata* (LC100082), *Notocomplana humilis* (LC100085), *Notocomplana koreana* (LC100086), *Notocomplana japonica* (LC100087), *Notoplana delicata* (LC100088), *Notocomplana* sp. (LC100089), *Prosthiostomum grande* (LC100090), *Pr. vulgaris* (LC100091), *Cycloporus japonicus* (LC100092), *Thysanozoon brochii* (LC100093), *Thysanozoon japonicum* (LC100094), *Pseudoceros velutinus* (LC100095), *Pseudoceros nipponicus* (LC100096), *Pseudobiceros nigromarginatus* (LC100097), *Pseudoceros atropurpureus* (LC100098), *Pseudobiceros flavomarginatus* (LC100099), *Pseudoceros hancockanus* (LC100100), *Planocera* cf. *heda* (LC341282), *Pl. reticulata* (LC341284), *Planocera* sp. MK-2019C (LC516517), *Planocera* sp. MK-2019I (LC516523), and *Paratoplana renatae* (AJ270176). The 28S rRNA gene sequences of acotylean and cotylean flatworms were aligned using Clustal W [57]. A phylogenetic analysis based on the partial sequence of 28S rRNA gene (approximately 700 bp) was constructed by the maximum likelihood method with Kimura 2-parameter + G with 1000 bootstrap using MEGA X version 10.1 [58].

4. Conclusions

In summary, we report the first detection of TTX and 5,6,11-trideoxyTTX from the cotylean flatworm, *Pr. trilineatum*, collected from the coastal area of Hayama, Kanagawa, Japan. Although investigation of further specimens is suggested, our results suggest that the distribution of TTXs among flatworms might be wider than speculated, and it would thus be interesting to search for TTX or its precursor analogs in other flatworms. These findings will be useful in determining the origin of TTX, and elucidating its biosynthetic pathway.

Author Contributions: M.K. collected samples. M.K., R.S. (Rei Suo), and H.O. performed the LC-MS/MS and molecular analysis. M.A., R.N., R.S. (Ryo Sakakibara), and T.N. synthesized 5,6,11-trideoxyTTX. S.I. designed the study, supervised the data analysis, and interpreted the results. R.S. (Rei Suo) wrote the first draft of the manuscript, and R.S. (Rei Suo), S.I., T.N., and H.S. discussed the results and commented on the manuscript. All authors have read and agreed to the published version of the manuscript.

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References

1. Tsuda, K.; Ikuma, S.; Kawamura, M.; Tachikawa, R.; Sakai, K.; Tamura, C.; Amakasu, O. Tetrodotoxin. VII. On the structures of tetrodotoxin and its derivatives. *Chem. Pharm. Bull.* **1964**, *12*, 1357–1374. [[CrossRef](#)]
2. Woodward, R.B. The structure of tetrodotoxin. *Pure Appl. Chem.* **1964**, *86*, 49–74. [[CrossRef](#)]
3. Goto, T.; Kishi, Y.; Takahashi, S.; Hirata, Y. Tetrodotoxin. *Tetrahedron* **1965**, *21*, 2059–2088. [[CrossRef](#)]
4. Yasumoto, T.; Nagai, H.; Yasumura, D.; Michishita, T.; Endo, A.; Yotsu, M.; Kotaki, Y. Interspecies distribution and possible origin of tetrodotoxin. *Ann. N. Y. Acad. Sci.* **1986**, *479*, 44–51. [[CrossRef](#)] [[PubMed](#)]
5. Noguchi, T.; Arakawa, O.; Takatani, T. TTX accumulation in pufferfish. *Comp. Biochem. Physiol. Part D* **2006**, *1*, 145–152. [[CrossRef](#)]
6. Miyazawa, K.; Noguchi, T. Distribution and origin of tetrodotoxin. *J. Toxicol. Toxin Rev.* **2001**, *20*, 11–33. [[CrossRef](#)]
7. Narahashi, T. Pharmacology of tetrodotoxin. *J. Toxicol. Toxin Rev.* **2001**, *20*, 67–84. [[CrossRef](#)]
8. Colquhoun, D.; Henderson, R.; Ritchie, J.M. The binding of labeled tetrodotoxin to non-myelinated nerve fibres. *J. Physiol.* **1972**, *227*, 95–126. [[CrossRef](#)]
9. Makarova, M.; Rycek, L.; Hajicek, J.; Baidilov, D.; Hudlicky, T. Tetrodotoxin: History, biology, and synthesis. *Angew. Chem. Int. Ed.* **2019**, *58*, 18338–18387. [[CrossRef](#)]
10. Yasumoto, T.; Yasumura, D.; Yotsu, M.; Michishita, T.; Endo, A.; Kotaki, Y. Bacterial production of tetrodotoxin and anhydrotetrodotoxin. *Agric. Biol. Chem.* **1986**, *50*, 793–795.
11. Noguchi, T.; Jeon, J.K.; Arakawa, O.; Sugita, H.; Deguchi, Y.; Shida, Y.; Hashimoto, K. Occurrence of tetrodotoxin and anhydrotetrodotoxin in *Vibrio* sp. isolated from the intestines of a xanthid crab, *Atergatis floridus*. *J. Biochem.* **1986**, *99*, 311–314. [[CrossRef](#)] [[PubMed](#)]
12. Noguchi, T.; Arakawa, O. Tetrodotoxin—Distribution and accumulation in aquatic organisms, and cases of human intoxication. *Mar. Drugs* **2008**, *6*, 220–242. [[CrossRef](#)]
13. Ueyama, N.; Sugimoto, K.; Kudo, Y.; Onodera, K.; Cho, Y.; Konoki, K.; Nishikawa, T.; Yotsu-Yamashita, M. Spiro bicyclic guanidino compounds from pufferfish: Possible biosynthetic intermediates of tetrodotoxin in marine environments. *Chem. Eur. J.* **2018**, *24*, 7250–7258. [[CrossRef](#)] [[PubMed](#)]
14. Kudo, Y.; Yasumoto, T.; Mebs, D.; Cho, Y.; Konoki, K.; Yotsu-Yamashita, M. Cyclic guanidine compounds from newts support the hypothesis that tetrodotoxin is derived from monoterpene. *Angew. Chem. Int. Ed.* **2016**, *55*, 8728–8731. [[CrossRef](#)] [[PubMed](#)]
15. McNabb, P.; Selwood, A.I.; Munday, R.; Wood, S.A.; Taylor, D.I.; MacKenzie, L.A.; van Ginkel, R.; Rhodes, L.L.; Cornelisen, C.; Heasman, K.; et al. Detection of tetrodotoxin from the grey side-gilled sea slug—*Pleurobranchaea maculata*, and associated dog neurotoxicosis on beaches adjacent to the Hauraki Gulf, Auckland, New Zealand. *Toxicon* **2010**, *56*, 466–473. [[CrossRef](#)] [[PubMed](#)]
16. Salvitti, L.; Wood, S.A.; Taylor, D.I.; McNabb, P.; Cary, S.C. First identification of tetrodotoxin (TTX) in the flatworm *Stylochoplana* sp.; a source of TTX for the sea slug *Pleurobranchaea maculata*. *Toxicon* **2015**, *95*, 23–29. [[CrossRef](#)] [[PubMed](#)]
17. Salvitti, L.R.; Wood, S.A.; Winsor, L.; Cary, S.C. Intracellular immunohistochemical detection of tetrodotoxin in *Pleurobranchaea maculata* (Gastropoda) and *Stylochoplana* sp. (Turbellaria). *Mar. Drugs* **2015**, *13*, 756–769. [[CrossRef](#)]
18. Itoi, S.; Kozaki, A.; Komori, K.; Tsunashima, T.; Noguchi, S.; Kawane, M.; Sugita, H. Toxic *Takifugu pardalis* eggs found in *Takifugu niphobles* gut: Implications for TTX accumulation in the pufferfish. *Toxicon* **2015**, *108*, 141–146. [[CrossRef](#)]
19. Itoi, S.; Ueda, H.; Yamada, R.; Takei, M.; Sato, T.; Oshikiri, S.; Wajima, Y.; Ogata, R.; Oyama, H.; Shitto, T.; et al. Including planoceric flatworms in the diet effectively toxifies the pufferfish, *Takifugu niphobles*. *Sci. Rep.* **2018**, *8*, 12302. [[CrossRef](#)]
20. Itoi, S.; Sato, T.; Takei, M.; Yamada, R.; Ogata, R.; Oyama, H.; Teranishi, S.; Kishiki, A.; Wada, T.; Noguchi, K.; et al. The planoceric flatworm is a main supplier of toxin to tetrodotoxin-bearing fish juveniles. *Chemosphere* **2020**, *249*, 126217. [[CrossRef](#)]
21. Faubel, A. The Polycladida, Turbellaria. Proposal and establishment of a new system. Part I. The Acotylea. *Mitt. Hambg. Zool. Mus. Inst.* **1983**, *80*, 17–121.
22. Faubel, A. The Polycladida, Turbellaria. Proposal and establishment of a new system. Part II The Cotylea. *Mitt. Hambg. Zool. Mus. Inst.* **1984**, *81*, 189–259.
23. Rawlinson, K.A.; Litvaitis, M.K. Cotylea (Platyhelminthes, Polycladida): A cladistic analysis of morphology. *Invertebr. Biol.* **2008**, *127*, 121–138. [[CrossRef](#)]
24. Miyazawa, K.; Jeon, J.K.; Maruyama, J.; Noguchi, T.; Ito, K.; Hashimoto, K. Occurrence of tetrodotoxin in the flatworm *Planocera multitentaculata* (Platyhelminthes). *Toxicon* **1986**, *24*, 645–650. [[CrossRef](#)]
25. Tanu, M.B.; Mahmud, Y.; Arakawa, O.; Takatani, T.; Kajihara, H.; Kawatsu, K.; Hamano, Y.; Asakawa, M.; Miyazawa, K.; Noguchi, T. Immunoenzymatic visualization of tetrodotoxin (TTX) in *Cephalothrix* species (Nemertea: Anopla: Palaeonemertea: Cephalotrichidae) and *Planocera reticulata* (Platyhelminthes: Turbellaria: Polycladida: Planocericidae). *Toxicon* **2004**, *44*, 515–520. [[CrossRef](#)] [[PubMed](#)]

26. Ueda, H.; Itoi, S.; Sugita, H. TTX-bearing planoceric flatworm (Platyhelminthes: Acotylea) in the Ryukyu Islands, Japan. *Mar. Drugs* **2018**, *16*, 37. [[CrossRef](#)]
27. Ritson-Williams, R.; Yotsu-Yamashita, M.; Paul, V.J. Ecological functions of tetrodotoxin in a deadly polyclad flatworm. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 3176–3179. [[CrossRef](#)]
28. Yeri, M.; Kaburaki, T. Notes on two new species of Japanese polyclads. *Annot. Zool. Jpn.* **1920**, *9*, 591–598.
29. Pitale, R.; Bhawe, V.; Apte, D. First record of family *Prosthiostomidae* and *Prosthiostomum trilineatum* (Platyhelminthes: Polycladida) from the west coast of India. *Mar. Biodivers. Rec.* **2014**, *7*, 1–6. [[CrossRef](#)]
30. Nakamura, M.; Yasumoto, T. Tetrodotoxin derivatives in puffer fish. *Toxicon* **1985**, *23*, 271–276. [[CrossRef](#)]
31. Yotsu-Yamashita, M.; Schimmele, B.; Yasumoto, T. Isolation and structural assignment of 5-deoxytetrodotoxin from the puffer fish *Fugu poecilonotus*. *Biosci. Biotechnol. Biochem.* **1999**, *63*, 961–963. [[CrossRef](#)] [[PubMed](#)]
32. Yotsu-Yamashita, M.; Abe, Y.; Kudo, Y.; Ritson-Williams, R.; Paul, V.J.; Konoki, K.; Cho, Y.; Adachi, M.; Imazu, T.; Nishikawa, T.; et al. First identification of 5,11-dideoxytetrodotoxin in marine animals, and characterization of major fragment ions of tetrodotoxin and its analogs by high resolution ESI-MS/MS. *Mar. Drugs* **2013**, *11*, 2799–2813. [[CrossRef](#)] [[PubMed](#)]
33. Yasumoto, T.; Yotsu, M.; Murata, M.; Naoki, H. New tetrodotoxin analogue from the newt *Cynops ensicauda*. *J. Am. Chem. Soc.* **1988**, *110*, 2344–2345. [[CrossRef](#)]
34. Jang, J.H.; Yotsu-Yamashita, M. 6,11-DideoxyTTX from the puffer fish, *Fugu pardalis*. *Toxicon* **2007**, *50*, 947–951. [[CrossRef](#)]
35. Yotsu-Yamashita, M.; Yamagishi, Y.; Yasumoto, T. 5,6,11-Trideoxytetrodotoxin from the puffer fish, *Fugu poecilonotus*. *Tetrahedron Lett.* **1995**, *36*, 9329–9332. [[CrossRef](#)]
36. Khora, S.S.; Yasumoto, T. Isolation of 11-oxotetrodotoxin from the puffer *Arothron nigropunctatus*. *Tetrahedron Lett.* **1989**, *30*, 4393–4394. [[CrossRef](#)]
37. Endo, A.; Khora, S.S.; Murata, M.; Naoki, H.; Yasumoto, T. Isolation of 11-nortetrodotoxin-6(R)-ol and other tetrodotoxin derivatives from the puffer *Fugu niphobles*. *Tetrahedron Lett.* **1988**, *29*, 4127–4128. [[CrossRef](#)]
38. Yotsu, M.; Hayashi, Y.; Kohra, S.S.; Sato, S.; Yasumoto, T. Isolation and structural assignment of 11-nortetrodotoxin-6(S)-ol from the puffer *Arothron nigropunctatus*. *Biosci. Biotechnol. Biochem.* **1992**, *56*, 370–371. [[CrossRef](#)]
39. Yamada, R.; Tsunashima, T.; Takei, M.; Sato, T.; Wajima, Y.; Kawase, M.; Oshikiri, S.; Kajitani, Y.; Kosoba, K.; Ueda, H.; et al. Seasonal changes in the tetrodotoxin content of the flatworm *Planocera multitentaculata*. *Mar. Drugs* **2017**, *15*, 56. [[CrossRef](#)]
40. Itoi, S.; Tabuchi, S.; Abe, M.; Ueda, H.; Oyama, H.; Ogata, R.; Okabe, T.; Kishiki, A.; Sugita, H. Difference in tetrodotoxin content between two sympatric planoceric flatworms, *Planocera multitentaculata* and *Planocera reticulata*. *Toxicon* **2020**, *173*, 57–61. [[CrossRef](#)]
41. Kashitani, M.; Okabe, T.; Oyama, H.; Noguchi, K.; Yamazaki, H.; Suo, R.; Mori, T.; Sugita, H.; Itoi, S. Taxonomic distribution of tetrodotoxin in acotylean flatworms (Polycladida: Platyhelminthes). *Mar. Biotechnol.* **2020**, *22*, 805–811. [[CrossRef](#)] [[PubMed](#)]
42. Newman, L.J.; Cannon, L.R.G. *Marine Flatworms: The World of Polyclads*; CSIRO Publishing: Collingwood, Australia, 2003.
43. Newman, L.J.; Cannon, L.R.G. *Fabulous Flatworms: A Guide to Marine Polyclads*; CSIRO Publishing: Collingwood, Australia, 2005.
44. Okabe, T.; Oyama, H.; Kashitani, M.; Ishimaru, Y.; Suo, R.; Sugita, H.; Itoi, S. Toxic flatworm egg plates serve as a possible source of tetrodotoxin for pufferfish. *Toxins* **2019**, *11*, 402. [[CrossRef](#)] [[PubMed](#)]
45. Rawlinson, K.A. The diversity, development and evolution of polyclad flatworm larvae. *EvoDevo* **2014**, *5*, 9. [[CrossRef](#)] [[PubMed](#)]
46. Rodríguez, P.; Alfonso, A.; Otero, P.; Katikou, P.; Georgantelis, D.; Botana, L.M. Liquid chromatography–mass spectrometry method to detect tetrodotoxin and its analogues in the puffer fish *Lagocephalus sceleratus* (Gmelin, 1789) from European waters. *Food Chem.* **2012**, *132*, 1103–1111. [[CrossRef](#)]
47. Kosker, A.R.; Ozogul, F.; Ayas, D.; Durmus, M.; Ucar, Y.; Regenstein, J.M.; Ozogul, Y. Tetrodotoxin levels of three pufferfish species (*Lagocephalus* sp.) caught in the North-Eastern Mediterranean sea. *Chemosphere* **2019**, *219*, 95–99. [[CrossRef](#)]
48. Katikou, P.; Georgantelis, D.; Sinouris, N.; Petsi, A.; Fotaras, T. First report on toxicity assessment of the Lessepsian migrant pufferfish *Lagocephalus sceleratus* (Gmelin, 1789) from European waters (Aegean Sea, Greece). *Toxicon* **2009**, *54*, 50–55. [[CrossRef](#)]
49. McNabb, P.S.; Taylor, D.I.; Ogilvie, S.C.; Wilkinson, L.; Anderson, A.; Hamon, D.; Wood, S.A.; Peake, B.M. First detection of tetrodotoxin in the bivalve *Paphies australis* by liquid chromatography coupled to triple quadrupole mass spectrometry with and without precolumn reaction. *J. AOAC Int.* **2014**, *97*, 325–333. [[CrossRef](#)]
50. Turner, A.D.; Powell, A.; Schofield, A.; Lees, D.N.; Baker-Austin, C. Detection of the pufferfish toxin tetrodotoxin in European bivalves, England, 2013 to 2014. *Eurosurveillance* **2015**, *20*, 2–8. [[CrossRef](#)]
51. Turner, A.D.; Dhanji-Rapkova, M.; Coates, L.; Bickersta, L.; Milligam, S.; O’Neill, A.; Faulkner, D.; McEneny, H.; Baker-Austin, C.; Lees, D.N.; et al. Detection of tetrodotoxin shellfish poisoning (TSP) toxins and causative factors in bivalve molluscs from the UK. *Mar. Drugs* **2017**, *15*, 277. [[CrossRef](#)]
52. Gerssen, A.; Bovee, T.; Klijnstra, M.; Poelman, M.; Portier, L.; Hoogenboom, R. First report on the occurrence of tetrodotoxins in bivalve mollusks in the Netherlands. *Toxins* **2018**, *10*, 450. [[CrossRef](#)]
53. Dell’Aversano, C.; Tartaglione, L.; Polito, G.; Dean, K.; Giacobbe, M.; Casabianca, S.; Capellacci, S.; Penna, A.; Turner, A.D. First detection of tetrodotoxin and high levels of paralytic shellfish poisoning toxins in shellfish from Sicily (Italy) by three different analytical methods. *Chemosphere* **2019**, *215*, 881–892. [[CrossRef](#)] [[PubMed](#)]
54. Biessy, L.; Boundy, M.J.; Smith, K.F.; Harwood, D.T.; Hawes, I.; Wood, S.A. Tetrodotoxin in marine bivalves and edible gastropods: A mini-review. *Chemosphere* **2019**, *236*, 124404. [[CrossRef](#)] [[PubMed](#)]

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55. Adachi, M.; Sakakibara, R.; Satake, Y.; Isobe, M.; Nishikawa, T. Synthesis of 5,6,11-trideoxytetrodotoxin. *Chem. Lett.* **2014**, *43*, 1719–1721. [[CrossRef](#)]
 56. Tsunashima, T.; Hagiya, M.; Yamada, R.; Koito, T.; Tsuyuki, N.; Izawa, S.; Kosoba, K.; Itoi, S.; Sugita, H. A molecular framework for the taxonomy and systematics of Japanese marine turbellarian flatworms (Platyhelminthes, Polycladida). *Aquat. Biol.* **2017**, *26*, 159–167. [[CrossRef](#)]
 57. Thompson, J.D.; Higgins, D.G.; Gibson, T.J. CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **1994**, *22*, 4673–4680. [[CrossRef](#)] [[PubMed](#)]
 58. Kumar, S.; Stecher, G.; Li, M.; Knyaz, C.; Tamura, K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Mol. Biol. Evol.* **2018**, *35*, 1547–1549. [[CrossRef](#)] [[PubMed](#)]