SUPPLEMENTARY INFORMATION

Highly Fluorescent Distyrylnaphthalene Derivatives as a Tool for Visualization of Cellular Membranes

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1. General remarks

Unless stated otherwise, all air and water sensitive reactions were carried out under an argon atmosphere using freshly distilled dry solvents. All glassware was dried prior to use by heating under vacuum. Dimethyl 2,6-naphthalenedicarboxylate, diethanolamine, morpholine and trimethyl phosphite were purchased from Sigma-Aldrich. Commercial grade reagents and solvents were used without further purification except as indicated below. THF was distilled over Na/benzophenone prior to use. NMR spectra were recorded on Bruker DRX 500 spectrometer. ¹H, ¹³C chemical shifts are reported relative to the residual proton resonance in the deuterated solvents. All chemical shifts (δ) are given in ppm and the coupling constants (*J*) in Hz. Positive ion MALDI mass spectra were recorded on a Voyager-Elite (PerSeptive Biosystems Inc., Framingham, MA, USA) instrument equipped with nitrogen laser (337 nm) in a linear mode at an acceleration voltage of 20 kV and delayed extraction. Melting points are uncorrected.

2,6-Bis[4-(N,N-bis{6-[bis(2-hydroxyethyl)amino]hexyl}amino)styryl]naphthalene (DSNN

-DEA): A solution of DSNN-I (68mg, 0.056mmol) and diethanolamine (0.71g, 6.78mmol) in THF (10mL) was stirred at 70°C for 6h. The solvent and an excess of diethanolamine were evaporated under the reduced pressure. To the residue, saturated aqueous NaHCO₃ solution (5mL) was added and the mixture was extracted with CH₂Cl₂ (3 x 20mL). The combined organic extracts were washed with water and dried over Na₂SO₄. Evaporation of the solvent afforded DSNN-DEA (62mg, 98%) as a yellow-green solid. Mp 86-87 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.74 (d, *J* = 8.3 Hz, 2 H, C_{Ar}H), 7.73 (s, 2 H, C_{Ar}H), 7.68 (d, *J* = 8.7 Hz, 2 H,

 $C_{Ar}H$), 7.41 (d, J = 8.5 Hz, 4 H, $C_{Ar}H$), 7.14 (d, J = 16.2 Hz, 2 H, CH=CH), 7.02 (d, J = 16.1 Hz, 2 H, CH=CH), 6.63 (d, J = 8.6 Hz, 4 H, $C_{Ar}H$), 3.62 (t, J = 5.3 Hz, 16 H, OCH_2), 3.29 (t, J = 7.2 Hz, 8 H, $C_{Ar}NCH_2$), 2.65 (t, J = 5.2 Hz, 24 H, NCH_2CH_2O and OH), 2.53 (t, J = 7.3 Hz, 8 H, (HOCH₂CH₂)₂NCH₂), 1.61 (br s, 8 H, CH_2), 1.49 (br s, 8 H, CH_2), 1.35 (br s, 16 H, CH_2); ¹³C NMR (126 MHz, CDCl₃) δ 147.76 (2 C, $C_{Ar}N$), 135.32 (2 C, C_{Ar}), 132.94 (2 C, C_{Ar}), 128.82 (2 C, CH=CH), 127.98 (2 C, $C_{Ar}H$), 127.77 (4 C, $C_{Ar}H$), 125.21 (2 C, $C_{Ar}H$), 124.76 (2 C, C_{Ar}), 123.88 (2 C, $C_{Ar}H$), 123.83 (2 C, CH=CH), 111.79 (4 C, $C_{Ar}H$), 59.53 (8 C, OCH_2), 56.09 (8 C, NCH_2CH_2OH), 54.71 (4 C, (HOCH₂CH₂)₂NCH₂), 51.12 (4 C, $C_{Ar}NCH_2$), 27.39 (4 C, CH_2), 27.13 (8 C, CH_2), 26.99 (4 C, CH_2); MALDI MS: [M+H]⁺ 1111.7; Anal. calcd. for $C_{66}H_{106}N_6O_8$: C, 71.31; H, 9.61. Found: C, 71.09; H, 9.73.

2. NMR spectra



Figure S1. The ¹H-NMR and ¹³C-NMR spectra of DSNN-DEA derivative.

3. Cytotoxicity results

		MDM	NIH/3T3	293T	Fibroblasts	HCT116	HeLa	K562	HUVEC
DSNN-NMe₃⁺	1µM	60.0±3.7	80.1±3.5	90.8±3.9	93.1±7.6	104.2±7.7	*91.5±2.8	107.5±2.9	88.8±5.2
	5µM	19.2±1.0	43.1±1.7	50.7±4.2	46.0±5.2	59.4±1.9	*59.9±2.0	43.5±1.6	79.1±4.9
	10µM	17.0±1.4	41.7±1.3	46.1±3.2	47.7±3.1	43.6±3.0	*47.8±2.5	37.1±0.1	64.7±4.7
DSNN-P	1µM	76.5±4.3	100.5±2.5	99.9±3.5	86.3±4.2	103.4±8.6	*92.5±4.0	91.2±4.2	88.2±2.4
	5µM	74.5±5.8	100.5±1.4	100.9±4.4	97.5±4.0	112.0±6.7	*91.1±1.7	90.5±1.1	73.4±3.2
	10µM	76.0±4.2	100.1±2.4	103.3±2.5	97.5±6.1	104.7±7.7	*95.8±1.9	93.2±2.2	67.6±4.1
DSNN-Mor	1µM	82.5±7.1	107.1±2.9	97.7±2.1	97.3±3.0	108.6±5.8	*96.6±3.7	94.5±0.9	89.5±5.5
	5µM	69.5±6.9	109.7±3.5	107.7±2.6	91.3±2.6	104.2±2.9	*99.3±4.1	86.4±4.0	66.9±0.5
	10µM	82.8±6.5	103.0±4.0	96.0±3.3	104.4±3.7	115.0±5.3	*90.1±3.5	94.9±3.3	70.2±4.7
DSNN-DEA	1µM	70.6±5.4	80.7±5.3	97.0±1.5	90.8±5.7	97.8±1.9	85.3±4.5	85.2±4.5	83.3±3.2
	5µM	22.7±5.8	90.4±1.1	56.7±3.6	51.5±4.4	35.3±3.6	36.0±3.4	38.8±0.8	66.3±0.8
	10µM	20.4±1.2	27.7±2.2	41.9±3.4	8.5±2.0	24.7±3.1	30.4±2.9	37.1±0.3	58.4±3.5
DSNN-POK	1µM	101.5±3.4	100.8±5.1	97.8±1.3	84.6±2.4	97.8±2.7	*83.7±3.1	92.7±4.9	97.8±3.2
	5µM	94.3±4.8	98.9±2.7	92.6±2.7	81.6±3.5	82.2±2.1	*86.0±2.1	95.5±2.3	83.8±3.7
	10µM	86.6±4.3	84.9±4.5	100.9±1.6	103.9±4.2	88.9±3.3	*84.7±3.4	84.0±3.2	67.8±5.3
DSNN-NH ₂	1µM	73.3±4.7	99.2±3.4	98.5±1.3	76.8±2.2	89.1±2.5	*87.8±3.1	96.8±4.8	93.5±4.8
	5µM	31.2±3.4	59.3±3.1	90.7±4.9	62.9±3.5	82.9±2.6	*87.2±1.2	78.8±9.7	99.0±2.9
	10µM	32.7±5.9	47.4±1.9	93.9±2.0	62.8±2.7	80.6±4.4	*83.5±2.3	53.4±3.5	93.7±4.6
DSNN-Py⁺	1µM	40.4±1.9	77.1±1.4	89.9±2.9	59.7±4.9	95.6±6.4	*87.1±3.8	107.3±2.1	89.8±2.3
	5µM	19.6±1.9	58.5±2.0	68.2±9.4	37.9±2.6	42.3±0.9	*75.5±3.4	46.0±1.0	95.9±2.4
	10µM	17.3±4.4	46.4±2.9	72.2±7.6	41.2±9.9	39.9±2.3	*68.9±3.6	38.9±0.8	99.0±3.4

Table S1. In vitro cytotoxicity assay results (% living cells) performed on various cell lines after 72h of treatment with DSNN-compounds in 1 μ M, 5 μ M and 10 μ M. The results represent the mean ± standard error.



4. Absorption and emission spectra of DSNN-DEA derivative

Figure S2. Normalized excitation and emission spectra (PL) of DSNN-DEA derivative in various solvents: water (excitation – dark blue, emission - violet), MeOH (excitation - red, emission - blue), DMSO (excitation - green, emission - orange).



5. Fluorescence and confocal microscopy images

Figure S3. Fluorescence microscopy images of live HeLa cells after 24-hours incubation with DSNN-NMe₃⁺ in various culture media. A) HBSS, B) RPMI1640, C) complete medium (RPMI1640, 10% FBS, antibiotics), collected at 40x magnification. Left panel - phase contrasts, middle panel – B2A filter (400ms), right panel – UV-2A filter (300ms). All images are 400x320 μ m



Figure S4. Fluorescence microscopy images of HeLa cells incubated 24h with 1mM DSSN, next incubated 2,5h with 5mM staurosporine. Left panel – phase contrast, right panel – B2A filter (1s).



Figure S5. Fluorescence microscopy images of fixed HeLa cells after 24-hours incubation with 1 μ M DSNN-DEA at 1 μ M concentration, co-labeled 2-hours with A) 5 μ g/ml DAPI; B) 5 μ M BODIPY® TR; C) 1 μ M ErTracker; D) 0.1 μ M MitoTracker Orange, collected at 60x (A) and 40x (B-D) magnification. Left panel - B2A filter (exposure time - A 3s, B 1s, C 3s, D 1s); middle panel – DAPI filter (A 1s) or Texas red filter (B 3s, C 3s, D 3s); right panel – merged of left and middle panels. All images are 80x65 μ m



Figure S6. Confocal microscopy images of fixed HeLa cells after 24-hours incubation with 1 μ M DSNN-DEA at 1 μ M concentration, co-labeled 2-hours with 0.1 μ M MitoTracker Orange. Left panel - green fluorescence of tested compounds, middle panel - red fluorescence of MitoTracker, right panel - merged of left and middle panels. All images are 35x35 μ m.

5. FACS data



Figure S7 - A. K562 cells incubated1h with 1, 5 i $10\mu M$ DSNN-NMe3⁺



Figure S7 - B. K562 cells incubated 1h with 1, 5 i 10µM DSNN-P.



Figure S7 - C. K562 cells incubated1h with 1, 5 i 10µM DSNN-Mor.



Figure S7 - D. K562 cells incubated1h with 1, 5 i 10µM DSNN_DEA.



Figure S7 - E. K562 cells incubated1h with 1, 5 i 10µM DSNN-POK.



Figure S7 - F. K562 cells incubated1h with 1, 5 i 10µM DSNN-NH₂.



Figure S7 - G. K562 cells incubated1h with 1, 5 i 10µM DSNN-Py+



Figure S8 - A. MOLT4 cells incubated1h with 1, 5 i 10µM DSNN-NMe₃⁺.



Figure S8 - B. MOLT4 cells incubated 1h with 1, 5 i 10 μ M DSNN-P.



Figure S8 - C. MOLT4 cells incubated1h with 1, 5 i 10µM DSNN-Mor.



Figure S8_D. MOLT4 cells incubated1h with 1, 5 i 10µM DSNN_DEA.



Figure S8 - E. MOLT4 cells incubated1h with 1, 5 i 10µM DSNN-POK.



Figure S8 - F. MOLT4 cells incubated1h with 1, 5 i 10µM DSNN-NH₂.



Figure S8 - G. MOLT4 cells incubated1h with 1, 5 i 10 μM DSNN-Py^+