

Review

Blumea balsamifera—A Phytochemical and Pharmacological Review

Yuxin Pang ^{1,3,*}, Dan Wang ¹, Zuowang Fan ^{2,4}, Xiaolu Chen ¹, Fulai Yu ¹, Xuan Hu ², Kai Wang ³ and Lei Yuan ³

- ¹ Tropical Crops Genetic Resources Institute, Chinese Academy of Tropical Agricultural Sciences, Danzhou, Hainan 571737, China; E-Mails: wang_dan1414@163.com (D.W.); hillowchan@hotmail.com (X.C.); fulai.yu@163.com (F.Y.)
- ² Key Laboratory of Crop Gene Resources and Germplasm Enhancement in Southern China, Danzhou, Hainan 571737, China; E-Mails: fujianfanzuowang@126.com (Z.F.); mchuxuan@163.com (X.H.)
- ³ Hainan Provincial Engineering Research Center for *Blumea balsamifera*, Danzhou, Hainan 571737, China; E-Mails: jimojijie29@163.com (K.W.); ylei1216@126.com (L.Y.)
- School of Traditional Chinese Medicine, Guangdong Pharmaceutical University, Guangzhou, Guangdong 510006, China
- * Author to whom correspondence should be addressed; E-Mail: blumeachina@126.com; Tel.: +86-898-2330-0268; Fax: +86-898-2330-0246.

Received: 30 May 2014; in revised form: 27 June 2014 / Accepted: 2 July 2014 /

Published: 3 July 2014

Abstract: The main components of sambong (*Blumea balsamifera*) are listed in this article. The whole plant and its crude extracts, as well as its isolated constituents, display numerous biological activities, such as antitumor, hepatoprotective, superoxide radical scavenging, antioxidant, antimicrobial and anti-inflammation, anti-plasmodial, anti-tyrosinase, platelet aggregation, enhancing percutaneous penetration, wound healing, anti-obesity, along with disease and insect resistant activities. Although many experimental and biological studies have been carried out, some traditional uses such as rheumatism healing still need to be verified by scientific pharmacological studies, and further studies including phytochemical standardization and bioactivity authentication would be beneficial.

Keywords: Traditional Chinese Medicines; *Blumea balsamifera*; sambong; herbal authentication; phytochemistry; biological activities

1. Introduction

Nowadays, herbal medicines are widely consumed and their sales have been rising significantly all over the world. According to the reports of the World Health Organization (WHO), to treat diseases over 80% of the populations in developing countries mainly rely on herbs, which are considered to be safer and more effective than synthetic drugs [1–3].

Blumea balsamifera (L.) DC. (Asteraceae), also known as sambong, has been used as medicine for thousands of years in Southeast Asia countries, such as China, Malaysia, Thailand, Vietnam, and Philippines. Sambong is the most important member of the genus *Blumea* and is an indigenous herb of tropical and subtropical Asia, especially in China. This plant grows on forest edges, under forests, river beds, valleys and grasses [4,5]. In China, it is generally a common used herb in the areas south of the Yangtze River, such as Hainan, Guizhou, Yunnan, and Guangdong provinces and Taiwan [6–8]. B. balsamifera is commonly called "Ainaxiang" and "Dafeng'ai" in Chinese and used as incense because it has a high level of essential oils [9]. It was originally recorded in "Bei Ji Qian Jin Yao Fang" in 652 by Sun Simiao. The whole plant or its leaves were used as a crude Chinese traditional medicinal material to treat eczema, dermatitis, beriberi, lumbago, menorrhagia, rheumatism, skin injury, and as an insecticide [10]. Bing Pian and Aipian are two important traditional Chinese medicines (TCMs) extracted from plants and have been used as one in prescriptions for centuries in China. Both of them mainly contain borneol and are similar in efficacy [11]. They are synonymous in the Chinese pharmaceutical industry nowadays. Before 2010, sambong was one of the most important plant sources for Bing Pian, but since 2010, the Pharmacopoeia of the People's Republic of China records B. balsamifera as the only plant source for Aipian [11], with a consistent efficacy with B. balsamifera medicinal materials, which could induce resuscitation, clear heat, and relieve pain. Recently, extracts of its leaves have been verified do display various new physiological activities, such as antitumor [12–14], antifungal [13,15], radical-scavenging [16], and anti-obesity properties [17]. The main active compound is L-borneol, which was characterized by a high volatility. Besides, essential oils, flavonoids, and terpenoids with several different biological activities were also reported [18]. These studies could explain why this plant has multiple pharmacological effects.

In this review, botanical descriptions, herbal authentications, and phytochemical constituents of *B. balsamifera* are covered. In addition, the previous *in vitro* and *in vivo* studies conducted on its biological activities are reviewed, concentrating on antitumor, hepatoprotective, superoxide radical scavenging, antioxidant, antimicrobial, anti-inflammation, antiplasmodial, antityrosinase, platelet aggregation, wound healing, anti-obesity, disease and insect resistant activities as well as enhancing percutaneous penetration.

General Botanical Description

According to the description of Flora Republicae Popularis Sinicae and Chinese Materia Medica [19,20], B. balsamifera is a perennial herb or subshrub, which rises about 1–3 meters in height. Its stem is strong and taupe, and erects with taupe, longitudinal edges. Its upper internodes are covered by dense tawny nonglandular hair. Its leaves, when triturated, send out a unique, cool aroma, which can make people feel refreshed. The leaves are wide ovoid or oblong-lanceolate in shape at the bottom, 22–25 cm in length, 8–10 cm in width. Its base is attenuated with petiole, narrow linear appendants of 3–5 pairs on both sides, pubescenced above, slight brown or thick yellow-white silky-villous, highlighted below midrib, with lateral veins of 10–15 pairs. The leaves at the top are oblong-lanceolate or ovate-lanceolate in shape, 7-12 cm in length, 1.5-3.5 cm in width, with an acuminate apex, a slightly acuminate base, without petiole or with a short petiole with narrow linear appendants of 1-3 pairs, entire or with thin serration or pinnatopectinate. Capitulum is arranged in much more branched leafy panicles; a peduncle with yellow and dense pubescence; an involucre campanulate, and a dense pubescence at the back. Its flower is yellow, with numerous female parts; tubular corolla, receptacle honeycomb, and corolla thin tubulous. Akene is cylindrical, with five edges, a dense pubescence, and a red-brown rough hairy pappus. The flowering period almost covers the whole year. B. balsamifera often grows in forest edges, under forests, river beds, valleys, or grasslands, and the altitude is 600-1000 m. In addition to its various Chinese locations it is also distributed in India, Pakistan, Burma, Indo-China Peninsula, Malaysia, Indonesia and Philippines.

2. Phytochemistry

There have been more than 100 volatile or non-volatile constituents isolated from sambong, including monoterpenes, sesquiterpenes, diterpenes, flavonoids, organic acids, esters, alcohols, dihydroflavone, and sterols. The study of the plant mainly focused on the volatile oils and flavonoids, which possessed various bioactivities *in vivo* and *in vitro*. The chemical constituents of *B. balsamifera* have been reviewed earlier [8,18].

2.1. Volatile Constituents

The volatile constituents account for the largest amount of the constituents in *B. balsamifera*, which are the major active constituents containing terpenoids, fatty acids, phenols, alcohols, aldehydes, ethers, ketones, pyridines, furans, and alkanes (Table 1, Appendix A). The most important constituent is L-borneol. To address the need for a natural borneol, different extraction methods have been used to extract the volatile constituents from *B. balsamifera*. Steam distillation (SD), simultaneous distillation and extraction (SDE), and CO₂ supercritical extraction were the most common methods in the extraction of volatile oils [21–24]. Wang *et al.* have adopted SD, SDE, and headspace solid-phase micro-extraction (HS-SPME) in order to obtain the volatile compounds of *B. balsamifera* leaves [23]. The extracts were then isolated and were identified by gas chromatography mass spectrometry (GC-MS). They found that alcohols and terpenoids were the main volatile compounds and the terpenoids accounted for a considerable proportion. Fifty, twenty-four, and forty-nine kinds of compounds were extracted by SD, SDE, and HS-SPME, respectively.

Table 1. The known volatile constituents of *B. balsamifera*.

Table 1. The known volatile constituents of B. balsamijera.						
Classification		Compound name	Molecular formula	References		
	1	L-borneol	$C_{10}H_{18}O$	[22,25,26]		
	_ 2	Isoborneol	$C_{10}H_{18}O$	[22,23,27]		
	3	(+)-Limonene	$C_{10}H_{16}$	[21,25,28]		
	4	(–)-Limonene	$C_{10}H_{16}$	[23]		
	5	(E) Ocimene	$C_{10}H_{16}$	[23,27,28]		
	6	(Z)-β-Ocimene	$C_{10}H_{16}$	[23]		
	7	β-Myrcene	$C_{10}H_{16}$	[28]		
	8	Camphene	$C_{10}H_{16}$	[21,23,25,27,28]		
	9	α-Pinene	$C_{10}H_{16}$	[21,23,25,27,28]		
	10	β -Pinene	$C_{10}H_{16}$	[21,23,25,27,28]		
Monoterpenes	11	Terpinen-4-ol	$C_{10}H_{18}O$	[21,22,27]		
F	12	Perillyl alcohol	C ₁₀ H ₁₆ O	[21,28]		
	13	Chrysanthenone	C ₁₀ H1 ₄ O	[21,23,27]		
	14	α -Terpineol	C ₁₀ H ₁₈ O	[22,23,26,27]		
	15	Bornyl acetate	$C_{12}H_{20}O_2$	[23]		
	16	Sabinene	$C_{10}H_{16}$	[27]		
	17	α -Thujene	$C_{10}H_{16}$	[27]		
	18	Trans-linalool oxide	$C_{10}H_{18}O_2$	[27]		
	19	Linalooloxide	$C_{10}H_{18}O_2$ $C_{10}H_{18}O_2$	[21]		
	20	Camphor	$C_{10}H_{18}O_2$ $C_{10}H_{16}O$	[21,22,25–28]		
	21	1,8-Cineole		[26]		
	22	Perilla aldehyde	$C_{10}H_{18}O$			
Overgonated		j	$C_{10}H_{14}O$	[21,25,28]		
Oxygenated	23	Cuminaldehyde	$C_{10}H_{12}O$	[27,28]		
monoterpenes	24	Myrtenal	$C_{10}H_{14}O$	[21,27]		
	25	Thymohydroquinone dimethyl ether	$C_{12}H_{18}O_2$	[21]		
	26	α-Gurjunene	$C_{15}H_{24}$	[21,23,25,27]		
	27	Alloaromadendrene	$C_{15}H_{24}$	[23,25]		
	28	(+)-Aromadendrene	$C_{15}H_{24}$	[23]		
	29	Aromadendrene	$C_{15}H_{24}$	[21,23,27,28]		
	30	Aromadendrene oxide	C ₁₅ H ₂₄ O	[28]		
	31	Aromadendrene, dehydro	$C_{15}H_{22}$	[28]		
	32	Longifolene	$C_{15}H_{24}$	[23,27]		
	33	α-Caryophyllene	$C_{15}H_{24}$	[21,23,25–28]		
	34	β -Caryophyllene	$C_{15}H_{24}$	[21,25,27]		
	35	Caryophyllene oxide	$C_{15}H_{24}O$	[21,22,26–28]		
	36	Guaia-3,9-diene	$C_{15}H_{24}$	[26,28]		
	37	γ-Cadinene	$C_{15}H_{24}$	[21,27]		
	38	δ -Cadinene	$C_{15}H_{24}$	[21,27,28]		
	39	β -Selinene	$C_{15}H_{24}$	[27]		
Casquitarnanas	40	(+)- β -Selinene(β -Selinene)	$C_{15}H_{24}$	[26]		
Sesquiterpenes	41	β -Gurjunene	$C_{15}H_{24}$	[26]		
	42	(+)-γ-Gurjunene	$C_{15}H_{24}$	[23]		
	43	Thujopsene-13	$C_{15}H_{24}$	[23,28]		
	44	β -Elemene	$C_{15}H_{24}$	[28]		
	45	(–)-β-Elemene	$C_{15}H_{24}$	[23]		
	46	(−)-γ-Cadinene	$C_{15}H_{24}$	[23]		
	47	(−)-δ-Cadinene	$C_{15}H_{24}$	[23]		
	48	10-Epi-γ-Eudesmol	C ₁₅ H ₂₆ O	[21,23,25,27]		
	49	Globulol	C ₁₅ H ₂₆ O	[27,28]		
	50	(–)-Guaiol	C ₁₅ H ₂₆ O	[21,22,25,28]		
	51	Ledol	$C_{15}H_{26}O$	[21,23,25,28]		
	52	γ-Muurolene	$C_{15}H_{24}$	[26,28]		
	53	Elemol	$C_{15}H_{26}O$	[21,23,27]		
	54	α-Eudesmol	$C_{15}H_{26}O$	[21,25]		
	55	β -Eudesmol		[21,23,27,28]		
	33	ρ -Eudesiiioi	$C_{15}H_{26}O$	[41,43,41,40]		

Table 1. Cont.

Classification		Compound name	Molecular formula	References
	56	γ-Eudesmol	$C_{15}H_{26}O$	[21,23,25,28]
Sesquiterpenes	57	Carotol	$C_{15}H_{26}O$	[23,28]
	58	Cubenol	$C_{15}H_{26}O$	[23,28]
	59	16-Kaurene	$C_{20}H_{32}$	[23]
	60	1-Ang-4,7-dihydroxyeudesmane	$C_{20}H_{34}O_4$	[9]
	61	Phytol	$C_{20}H_{40}O$	[26,28]
	62	Blumeaene A	$C_{20}H_{30}O_{5}$	[9]
Diterpenes	63	Blumeaene B	$C_{20}H_{30}O_{5}$	[9]
	64	Blumeaene C	$C_{19}H_{30}O_5$	[9]
	65	Blumeaene D	$C_{21}H_{32}O_5$	[9]
_	66	Blumeaene E	$C_{20}H_{30}O_{6}$	[9]
	67	Blumeaene F	$C_{20}H_{30}O_{6}$	[9]
	68	Blumeaene G	$C_{20}H_{30}O_5$	[9]
	69	Blumeaene H	$C_{19}H_{30}O_5$	[9]
	70	Blumeaene I		
	71	Blumeaene J		[9]
Fatty acids	72	(11Z)-11-hexadecenoic acid		[23]
	73	Trans-2-undecenoic acid		[22]
	74	9-Hexadecenoic acid		[27]
	75	Capric acid		[27]
	76	Palmitic acid		[23]
Phenols	77	Xanthoxylin		[23,29,30]
	78	Eugenol		[21,22,26]
	79	Dimethoxydurene		[25]
A1 1 1	80	1-Octen-3-ol		[21,23,25,27]
Alcohols	81	3-Octanol	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	[21,23,25]
A111 1	82	3-Propyl benzaldehyde	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	[25,31]
Aldehydes	83	4-Isopropylbenzaldehyde		[21,22]
TZ 4	84	3-Octanone	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	[21,23]
Ketones	85	2-Hydroxy-4,6-dimethoxyacetophenone		[27]
Pyridines	86	3-Fluoro-5-amino-pyridine		[22]
F	87	Furan,4,5-diethy-2,3-dihydro-2,3-		[22]
Furans		dimethyl	$C_{10}H_{18}O$	[22]
	88	1,3,4,5,6,7-hexahydro-2,5,5-trimete-hyl-	CII	[22.25]
		2H-2,4A-et hanonaphthalene	$C_{10}\Pi_{14}$	[23,23]
	89	Paracymene (4-Isopropyltoluene)	$C_{10}H_{14}$	[27]
	90	3-Nitrophthalic acid		[23]
	91	α-Butenoic acid, 3-methoxy-4-nitro		[22]
Others	92	O-acetyl-1-serine		[22]
	93	4,4-		
		dimethyltetracyclo[6.3.2.0 ^{2,5} .0 ^{1.8}]tridecan-	$C_{13}H_{20}O$	[28]
		9-01	-	

Several volatile oils are contained in the leaves and branchlets of *B. balsamifera*, which are the key crude materials of refined borneol [28]. Volatile oil in *B. balsamifera* is a yellow oily liquid with a unique aroma [32]. The oil yield of this plant is at least 2.5 mg/g (D·W) in Guizhou, China. If the plants are fertilized in certain way, it can be much higher, may be up to 7.72 mg/g (D·W) [33]. In Hainan (China), Pang *et al.* also got an oil yield of 3.2–4.3 mg/g (unpublished data). Some previous studies have reported that the volatile oils of *B. balsamifera* mainly contained monoterpenes and sesquiterpenes, such as L-borneol, 10-epi- γ -eudesmol, γ -eudesmol, β -eudesmol, α -eudesmol, limonene, L-camphor, palmitic acid, and D-camphor [21]. Hao *et al.* have qualitatively and quantitatively analyzed the volatile oil of *B. balsamifera* growing in Guizhou Province by GC-MS [25]. A total of 28

constituents were identified in this study, including: L-borneol, β-caryophyllene, camphor, γ-eudesmol, 1-octen-3-ol, trans-β-ocimene, and 1,3,4,5,6,7-hexahydro-2,5,5-trimethyl-2*H*-2,4a-ethanonaphthalene. Fifty-six compounds were separated and identified in the volatile oils of B. balsamifera by GC-MS according to the studies of Du et al. [22]. According to their studies, the main constituents are borneol and camphor. Others are isoborneol, terpineol, caryophyllene, eugenol, guaiol, and cubenol. Bhuiyan et al. also studied the volatile oils of B. balsamifera leaves to isolate fifty constituents, which contributed to 99.07% of the oil [28]. The main constituents were borneol (33.22%), caryophyllene (8.24%), ledol (7.12%), and 4.4-dimethyltetracyclo $[6.3.2.0^{2.5}.0^{1.8}]$ tridecan-9-ol, (5.18%). It was difficult to give a trivial name to or classify 4,4-dimethyltetracyclo-[6.3.2.0^{2,5}.0^{1,8}]tridecan-9-ol, because it had a non-typical structure, which made it seem rather a fragment of some other compound(s). The contents of other constituents such as phytol, caryophyllene oxide, thujopsene-13, guaiol, dimethoxydurene, and y-eudesmol were 3%-5%. Wen et al. adopted a RP-HPLC method to measure the content of xanthoxylin in different parts of B. balsamifera [34]. Xia et al. selected fourteen different regions of B. balsamifera to establish a GC chromatographic fingerprint [35]. The sample plants were selected from Hongshuihe village (II), Luodian, and Guizhou with the highest concentration of L-borneol. Chu et al. reported 1,8-Cineole contribute 20.98% to B. balsamifera oil in Nanning, Guangxi Zhuang Autonomous Region, China, while Pang et al., didn't find it when they analyzed sambong oil with GC-MS (unpublished data), as well as some other research [22,25]. Sun, Shirota, and Xu also determined the contents of constituents in B. balsamifera oil [36–38]. The Aifen, B. balsamifera powder, is a crude product during the production of Aipian. Jiang et al. found that the yield and extraction rate of Aifen increased significantly if the plant materials were harvested from October to December [39].

2.2. Non-Volatile Constituents

2.2.1. Flavonoids Constituents

Flavonoids, including flavonoid, flavanone and chalcone constituents, are the major non-volatile constituents of *B. balsamifera* (Table 2, Appendix B). The ultrasonic extraction methods of total flavonoids of *B. balsamifera* were studied by Pang *et al.* [40]. It was found that 30% ethanol was the suitable extraction solvent with the solid/liquid ratio of 1/300. It was then extracted by ultrasonic frequency (85 Hz), twice, each time for 30 min, where the extraction yield was 208.6 mg/g. Previous phytochemical studies have shown that the leaves of *B. balsamifera* included a number of flavonoids, such as blumeatin, velutin, tamarixetin, dihydroquercetin-7,4'-dimethyl ether, ombuine, rhamnetin, luteolin-7-methyl ether, luteolin, quercetin, 5,7,3',5'-tetrahydroxyflavanone, and dihydroquercetin-4'-methyl ether [16,41–44]. With the advances in phytochemical studies of *B. balsamifera*, flavonoids have been recognized for their medicinal properties and the investigation, validation, standardization of the local plant have been developed as an herbal medicine. Ali *et al.* extracted 3,4',5-trihydroxy-3',7-dimethoxyflavanone from the ligroin extract of *B. balsamifera* leaves [31]. The constituents of 3',4',5-trihydroxy-7-methoxyflavanone and a new bioflavonoid, determined as 3-O-7"-biluteolin, were extracted by acetone. Zhu *et al.* isolated three constituents, including: xanthoxylin, blumeatin, dihydroquercetin-7, and 4'-dimethylethe [29]. Chen *et al.* also obtained three flavonoid compounds

from aerial parts of *B. balsamifera* [45]. The compounds of 5,7-dihydroxy-3,3',4'-trim ethoxy flavone, davidigenin, catechin, ayanin, and davidioside were originally extracted from *B. balsamifera*. In the same year, Huang *et al.* isolated and identified seven constituents from this plant [46]. The constituents of 5,4'-dihydroxy-7-methoxyflavone and 5,4'-dihydroxy-3,3',7-trimethoxy flavanone were first found in the plant. Deng *et al.*, Zhu, Saewan *et al.*, Yan *et al.*, and Tan *et al.* also studied the flavonoid constituents [14,29,47–49]. Liang *et al.* used the methods of silica gel column chromatography, TLC, and Sephadex LH-20 column chromatography in order to isolate the chemical constituents of the plant, the structures of which were determined by physico-chemical constants and spectral analysis [39]. As a result, seven constituents were isolated. Besides, the contents of the flavonoids were determined by different methods. The chromatographic methods of RP-HPLC (reversed-phase high-performance liquid chromatographic), and HPLC were the most widely used to determine the contents of flavonoids of *B. balsamifera* [44,50]. Huang *et al.* determined the total amount of flavonoids in different parts of *B. balsamifera* [51]. The results exhibited that the total flavonoid content was 2.94%, 1.21%, and 1.36% in the leaves, branch, and stem, respectively.

Table 2. The known non-volatile constituents of *B. balsamifera*.

Classification		Compound Name	Molecular Formula	References
Flavones	1	4',5-Dihydroxy-7-methyletherflavanone	$C_{16}H_{12}O_5$	[30,46]
	2	Luteolin	$C_{15}H_{10}O_6$	[9,14,30,39,46]
	3	Luteolin-7-methyl-ether	$C_{16}H_{12}O_6$	[14,30,46]
	4	Diosmetin (Luteolin 4'-methyl ether)	$C_{16}H_{12}O_6$	[48]
	5	Chrysoeriol (Luteolin 3'-methyl ether)	$C_{16}H_{12}O_{6}$	[48]
	6	Quercetin	$C_{15}H_{10}O_7$	[9,14,39,44]
	7	3,5,3',4'-Tetrahydroxy-7-methoxyflavone	$C_{16}H_{12}O_7$	[9,31,47]
	8	3,5,3'-Trihydroxy-7,4-dimethoxyflavone	$C_{17}H_{14}O_7$	[9,45,47]
	9	Rhamnetin (7-Methoxyquercetin)	$C_{16}H_{12}O_7$	[14,30,46]
	10	Tamarixetin	$C_{16}H_{12}O_7$	[14]
	11	Ombuine	$C_{17}H_{14}O_7$	[49]
	12	3,5,7-Trihydroxy-3'4'-dimethoxyflavone	$C_{17}H_{14}O_7$	[48]
F11-	13	3,3',4',5-Tetrahydroxy-7-methoxyflavone	$C_{16}H_{15}O_{6}$	[48]
Flavonols	14	3,5-Dihydroxy-3',4',7-trimethoxyflavone	$C_{18}H_{16}O_{7}$	[48]
	15	4',5-Dihydroxy-3,3',7-trimethoxy flavanone	$C_{18}H_{16}O_{7}$	[30,46]
	16	5,7-Dihydroxy-3,3',4',-trimethoxyflavone	$C_{18}H_{16}O_{7}$	[9,45]
	17	Ayanin	$C_{18}H_{16}O_{7}$	[9,45]
	18	Chrysosplenol C	$C_{18}H_{16}O_{8}$	[45]
	19	4',5,7-Trihydroxy-3,3'-dimethoxyflavone	$C_{17}H_{14}O_7$	[48]
	20	Hyperoside	$C_{21}H_{20}O_{12}$	[48]
	21	Isoquercitrin	$C_{21}H_{20}O_{12}$	[48]
Flavanones	22	Blumeatin (5,3',5'-trihydroxy-methoxydihydro-	C II O	
		flavone)	$C_{16}H_{14}O_{6}$	[9,14,29,30,45,52]
	23	Eriodictyol	$C_{15}H_{12}O_6$	[30,46]
	24	5,7,3',5'-Tetrahydroxyflavanone	$C_{15}H_{12}O_6$	[9,44,45]
	25	3',4',5-Trihydroxy-7-metoxyflavanone	$C_{16}H_{15}O_{8}$	[45]

Table 2. Cont.

Classification		Compound Name	Molecular Formula	References
Flavanonols	26	Dihydroquercetin-4'-methylether	$C_{16}H_{14}O_{7}$	[9,12,14,30,39,45,52,53]
	27	Dihydroquercetin-7,4'-dimethylether	$C_{17}H_{16}O_{7}$	[9,14,29,30,39,45,52,53]
	28	3,4'5-Trihydroxy-3'7-dimethoxyflavanone	$C_{17}H_{18}O_9$	[45]
	29	3,3',5,5',7-Pentahydroxyflavanone	$C_{15}H_{12}O_7$	[48]
	30	3,3',4',5-Tetrahydroxy-7-methoxyflavanone	$C_{16}H_{15}O_{8}$	[48]
	31	3,3',5-Trihydroxy-4',7-dimethoxyflavanone	$C_{17}H_{18}O_9$	[48]
	32	3,3',5,7-Tetrahydroxy-4'-methoxyflavanone	$C_{16}H_{15}O_{8}$	[48]
_	33	3',4',5-Trihydroxy-3,7-dimetoxyflavanone	$C_{17}H_{18}O_9$	[48]
F11-	34	Catechin	$C_{15}H_{14}O_{6}$	[9,45]
Flavanols	35	(2R,3R)-(+)-7-O-Methyldihydroquercetin	$C_{16}H_{14}O_{7}$	[53]
Chalanna	36	Davidioside	$C_{21}H_{24}O_9$	[9,45]
Chalcones	37	Davidigenin	$C_{15}H_{14}O_4$	[9,45]
C : t	38	Blumealactone A	$C_{19}H_{26}O_{6}$	[54]
Sesquiterpene lactone	39	Blumealactone B	$C_{20}H_{28}O_{6}$	[54]
	40	Blumealactone C	$C_{16}H_{22}O_{6}$	[54]
Sterides	41	β -Sitosterol	$C_{29}H_{50}O$	[9]
	42	5α , 8α -Epidioxyergosta-6,22-dien- 3β -ol	$C_{28}H_{44}O_3$	[9]
	43	Daucosterol	$C_{35}H_{60}O_{6}$	[9,39]
Diterpenes	44	Cryptomeridiol	$C_{15}H_{28}O_2$	[55]
Tuitamaanaa	45	3,13-Clerodadiene-6,15-diol	$C_{20}H_{34}O_2$	[9]
Triterpenes	46	Austroinulin	$C_{20}H_{34}O_{3}$	[55]
Lignans	47	Syringaresinol	$C_{22}H_{26}O_{8}$ [9]	
Commonis	48	Hydranngetin	$C_{10}H_8O_4$	[9]
Coumarin	49	Umberlliferone (7-hydroxycoumarin)	$C_9H_6O_3$	[9]
Naphthalenone	50	5,7-Dihydroxychromone	$C_9H_6O_4$	[49]

2.2.2. Sterols

Apart from the above constituents, a small number of sterols were also isolated from *B. balsamifera*. Zhao *et al.* obtained colorless acicular as well as sheet crystals from *B. balsamifera* by silica gel column chromatography [56], where the crystals were identified to be stigmasterol and β -sitosterol by TLC and melting point measurement. Chen isolated β -sitosterol, daucosterol, and 5α ,8 α -epidioxyergosta-6,22-dien-3 β -ol from the aerial sections of *B. balsamifera* collected from Mengla, Yunnan by MS determination [9]. Liang *et al.* yielded seven compounds, which were isolated from ethyl acetate and chloroform extract, including daucosterol [39].

2.2.3. Sesquiterpene Lactone (SLs)

Sesquiterpene lactones (SLs) are a group of common chemicals in many Asteraceae plants. They were famous because they had cytotoxic and potential to be tumor inhibitors [57,58]. In sambong, a member of Asteraceae family, there were three sesquiterpene lactones, Blumealactone A, Blumealactone B, and Blumealactone C. Fujimoto *et al.* isolated them by extracting its dried leaves with 90% ethanol [54].

2.2.4. Other Constituents

There were some other constituents in this plant. Chen found two coumarin constituents, such as umberlliferone and hydranngetin, in *B. balsamifera*. He also found a lignans constituent, which was syringaresinol [9].

3. Biological Activities

3.1. Antitumor Activity

Hasegawa et al. extracted a dihydroflavonol from B. balsamifera as a result of screening among more than 150 plant materials [12]. The dihydroflavonol components showed the most significant synergism with tumor related apoptosis inducing ligand (TRAIL). It enhanced the level of TRAIL-R2 promoter activity and promoted the expression of surface protein in a p53-independent manner. The ethanol extract of B. balsamifera leaves was tested on male mice to investigate its hepatoxicity. The results exhibited that the hepatic cells, sitplasm, nucleus, and sinusoid of the mice liver were damaged through some changes in the liver color and texture [59]. The methanol extract of B. balsamifera inhibited the growth in rat and showed no cytotoxicity on human hepatocellular carcinoma cells. The methanol extract decreased the expression of cyclin-E and phosphorylation of retinoblastoma (Rb) protein resulting in cell cycle arrest. Likewise, it decreased the level of the proliferation related ligand (APRIL) [60,61]. Moreover, the methanol extract of B. balsamifera was used to determine its cytotoxicity on a panel of human cancer cell lines by MTT assay. There was no regular or acute cytotoxicity on the cells of HepG2, HCT-116, T-47D, NCl-H23 and CCD-18Co [62]. Saewan et al. found six compounds out of nine isolated flavonoids to have cytotoxicity against KB, MCF-7, and NCI-H187 cancer cell lines [14]. These six compounds were evaluated for cytotoxicity against KB, MCF-7, and NCI-H187 cancer cell lines. Three compounds were active against the KB cells with the IC₅₀ values of 17.09, 47.72, and 17.83 µg/mL, respectively. Another three compounds exhibited a moderate activity against the NCI-H187 cells with the IC₅₀ values of 16.29, 29.97, and 20.59 µg/mL. Luteolin-7-methyl ether showed a strong cytotoxicity against human lung cancer (NCI-H187) cell lines with an IC₅₀ of 1.29 µg/mL and a moderate toxicity against oral cavity cancer (KB) cell lines with an IC₅₀ of 17.83 μg/mL. Li et al. studied the antitumor activity determined by means of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay [13]. The three endophytic streptomycetes strains of B. balsamifera, including: YIM 56092, YIM 56093, and YIM 56099 exhibited anticancer activity. Yet, different strains displayed different antitumor activities. The YIM 56092 strain displayed a cytotoxic activity on polyketide synthases I (PKS-I) nonribosomal peptide synthetases (NRPS) and P388D1. The YIM 56093 strain displayed a cytotoxic activity on PKS-II, NRPS, and P388D1. The YIM 56099 was on the PKS-I, PKS-II, and NRPS. Fuijimoto et al., extracted blumealactone A, B, and C from sambong's dried leaves and found them could inhibit the growth of Yoshida sarcoma at the concentration of 5–10 µg/ml [54]. Lee disclosed a medication combination including sambong (Ainaxiang) and found it could enhance the efficiency of curing hepatoma and pancreatic cancer treatments [63].

3.2. Hepatoprotective Effects

Xu *et al.* demonstrated that oral blumeatin (5,3',5-trihydroxy-7-methoxydihydroflavone) exhibited a significant protective activity against the liver injury caused by paracetamol and prednisolone [64]. Furthermore, Xu and Zhao have shown that five other *blumea* flavanones possessed protective activity for acute experimental liver injury [65]. Pu *et al.* further verified the five *blumea* flavanones protecting the hepatocytes against lipid peroxidation, which was induced by CCl₄ or FeSO₄+cysteine. Certain concentration of the five compounds (10–100 μmol/L) inhibited the malonaldehyde production, GSH depletion, and GPT leakage of hepatocytes [66]. Furthermore, *blumea* flavanone II showed the strongest activity. They also reported that the *blumea* flavones had protective effects against acute liver injury induced by different chemicals [67].

3.3. Superoxide Radical Scavenging Activity

The methanol extracts of *B. balsamifera* leaves showed a higher radical scavenging activity than the chloroform extracts. However, the pet-ether extracts had less activity against nonenzymatically generated superoxide radicals. The capacity of nine kinds of flavonoids (100 mmol/L) of the plant was decreased as follows: quercetin > luteolin > 5.7.3'.5'-tetrahydroxyflavanone > blumeatin > rhamnetin > tamarixetin > luteolin-7-methyl ether > dihydroquercetin-4'-methyl ether > dihydroquercetin-4',7-dimethyl ether. The flavonoids showed more activity than methylated compounds [16]. Zhao and Xu studied antiperoxidant activities of five compounds of *blumea* flavanones [68]. As a result, five compounds with the concentration of $10^{-5}-10^{-4}$ mol/L of blocked malondialdehyde formation in homogenates and in liver mitochondria of rats *in vitro*.

3.4. Antioxidant Activity

Nguyen *et al.* demonstrated the methanol extracts of *B. balsamifera* (collected in Lam Dong province) with strong xanthine oxidase inhibitory activity with an IC₅₀ value of 6.0 μg/mL [69]. They also verified the seven compounds of *B. balsamifera* methanol extract in Vietnam, which exhibited significant xanthine oxidase inhibitory activity. Three compounds among them, such as (2*R*,3*S*)-(-)-4'-*O*-methyldihydroquercetin, quercetin, and quercetin-3,3',4', showed a higher potent inhibitory activity, with their IC₅₀ values ranging from 0.23 to 1.91 mmol/L, as compared to that of the positive control allopurinol (IC₅₀ of 2.50 mmol/L) [70]. Furthermore, Nessa *et al.* have found that the methanol extract of *B. balsamifera* exhibited a higher xanthine oxidase inhibitory activity as compared to that of the chloroform and pet-ether extracts [71]. The activity of the isolated flavonoids' order was as follow: allopurinol > luteolin > quercetin > tamarixetin > 5,7,3',5'-tetrahydroxyflavanone > rhamnetin > luteolin-7-methylether > blumeatin > dihydroquercetin-4'-methylether > dihydroquercetin-7,4'-dimethyl ether > L-ascorbic acid. The plant of *B. balsamifera* was collected in Taiwan and the dry powder of the whole plant was extracted twice with methanol. The methanol extracts showed an activity on scavenging 1,1-diphenyl-2-picrylhydrazyl (DPPH) radicals with an IC₅₀ value of 72 g/mL. The superoxide anion scavenging activity was over 200 g/mL [72].

3.5. Anti-Microbial and Anti-Inflammation Activity

Ongsakul *et al.* claimed that the crude aqueous and ethanolic extracts of *B. balsamifera* displayed no significant antibacterial activity against the strains of *Staphylococcus aureus* and *Escherichia coli* [73]. However, the stain of *B. balsamifera*, including YIM 56092 and YIM 56093, displayed a significant activity against *S. epidermidis*, such that YIM 56099 was active against *E. coli*. There seems to be no antimicrobial activity against *S. aureus*, *Klebsiella pneumonia*, and *Candida albicans* [13]. Chen isolated twelve new compounds [9], four of which displayed inhibitory activities against LPS-induced NO production in RAW 264.7 with the IC₅₀ values of 40.06, 46.35, 57.80, and 59.44 μg/mL, respectively. Sakee *et al.* reported the essential oil of *B. balsamifera* to have a minimum inhibitory concentration (MIC) of 150 μg/mL and 1.2 mg/mL against *Bacillus cereus*, *S. aureus* and *Candida albicans*, respectively [74]. Furthermore, the hexane extract inhibited *Enterobacter cloacae* and *S. aureus*. These results suggested that the extracts of *B. balsamifera* possessed an activity against certain kinds of infectious and toxin-producing microorganisms. It could potentially be utilized to prevent and treat microbial diseases.

3.6. Antiplasmodial Activities

According to the traditional efficacy of relieving fever, the methanol extract of *B. balsamifera* from Forest Research Institute Malaysia was investigated for any potential antiplasmodial activity. The extracts of roots and stems exhibited some activity against *Plasmodium falciparum* D10 strain (sensitive strain) with an IC₅₀ value of $(26.25 \pm 2.47) \,\mu\text{g/mL}$ and $(7.75 \pm 0.35) \,\mu\text{g/mL}$, respectively [75].

3.7. Antityrosinase Activities

The ethylacetate extract consisting of nine flavonoids were isolated from the leaves of *B. balsamifera*. Their antityrosinase activities were surveyed by Saewan *et al.* [14]. According to their reports, compared with arbutin, two dihydroflavonols, dihydroquercetin-4'-methylether and dihydroquercetin-7,4'-dimethylether, and three flavonols, quercetin, rhamnetin and tamarixetin, showed a significantly higher inhibitory activity, but another two flavanones, 5,7,3',5'-tetrahydroxyflavanone and blumeatin, and two flavones, luteolin and luteolin-7-methyl ether, showed a lower inhibitory activity. The possible mechanism of the antityrosinase activity might be the cause of chelating with copper in the active center of tyrosinase.

3.8. Platelet Aggregation Activities

The concentration of 1.26 µmol/L blumeatin displayed a significant promoting activity on the rat and human platelet aggregation caused by arachidonic acid, 5-hydotypamice, and epinephrine. However, concentrations of 0.315 and 2.52 µmol/L inhibited platelet aggregation. It suggested that the effects of blumeatin on the platelet aggregation were dependent upon the concentration used. The injection of *B. balsamifera* extracts decreased the blood pressure, expanded the blood vessels, and inhibited the sympathetic nervous system in order to address the high pressure and insomnia. The infusion of the plant also had the function of diuresis [67].

3.9. Enhancing Percutaneous Penetration Activity

The L-borneol, as the main effective compound of *B. balsamifera*, showed a percutaneous penetration enhancer effect. The essential oil camphor and 1-menthol of the plant specifically promoted the percutaneous absorption of nicotinamide [9]. Fu *et al.* further verified the 0.5%, 1.0%, and 2.0% *B. balsamifera* oil enhancing albuterol sulfate transdermal absorption, respectively [76]. The percutaneous penetration of a combination of 1.0% *B. balsamifera* oil and 1.0% azone was less than that of their separate uses.

3.10. Wound Healing Activity

Wang *et al.* discovered that the external application of *B. balsamifera* oil on the intact and damaged skin of rats exhibited no acute toxicity [77]. The rats with pure *B. balsamifera* oil exposure at dosage of 2000 mg/kg for 24 h showed no allergic reaction or acute toxicity reaction, but a better wound recovery activity as compared to the one treated with non-*B. balsamifera* oil formulations. The results were consistent with the traditional use in ethnic minority, Li and Miao, in China in order to heal the skin wound and itch [19].

3.11. Anti-Obesity Activity

Kubota *et al.* reported that the extracts of *B. balsamifera* inhibited the lipid accumulation and glycerol-3-phosphate dehydrogenase (GPDH) activities [17], which mainly decreased the expressions of key adipogenic transcription factors, such as peroxisome proligerator-activated receptor γ , CCAAT element binding protein, and leptin in the 3T3-L1 adipocytes. Therefore, the extracts of *B. balsamifera* might possess antidiabetic, antiatherogenic, and anti-inflammatory functions. The methanol extracts of *B. balsamifera* (100 µg/mL) were used to investigate the ability of inhibiting blood vessel's formation by the method of rat aortic ring. The results exhibited that there was no remarkable differences between *B. balsamifera* extracts and vehicle control treatment [62].

3.12. Disease and Insect Resistant Activity

Luo et al. reported that the acetone extracts of B. balsamifera possessed an activity against Pryicutaria oryzae, Fusarium oxysporum sp., Colletorichum musae, C. gloeosporioides, C. capsici, and F. oxysporum f. sp. in vitro, with an inhibition rate of over 90.0% [78]. The volatile oil of B. balsamifera inhibited Aeromonas hydrophila, F. graminearum, and Magnaporthe grisea [27]. Wang et al. also demonstrated that the extract of B. balsamifera leaves showed a 60.8% insecticidal activity against the adult Aleurodicus disperses [23]. Furthermore, the essential oil of B. balsamifera showed fumigant toxicity against the maize weevils, such as Sitophilus zeamais [26]. The crude oil also induced the death in the S. zeamais adults. The results suggested that the extracts of B. balsamifera possessed significant disease and insect resistant activities, and could be used as new potential plant pesticides.

4. Conclusions

In traditional medicine, the source of the plant should be primarily clear and definite. Thus, the general botanical description should be reviewed according to the Flora Republicae Popularis Sinicae and Chinese Materia Medica [19,20]. The herbal authentication of B. balsamifera was then further carried out to verify the plant source and medicinal efficacy [19]. These studies confirmed the accuracy and uniqueness of the source of B. balsamifera. In the same way, as a folk medicine, B. balsamifera has been widely used in South Asian countries, especially in China. Due to the chemical constituents contained in this plant for the different effects, the phytochemical research has been reviewed in this article. The chemical constituents were mostly volatile and non-volatile. The former constituents occupied the largest amount, which mainly included: terpenoids, fatty acids, phenols, alcohols, aldehydes, ethers, ketones, pyridines, furans, and alkanes. L-borneol was the most abundant and active constituent in this plant, while other constituents included flavonoids, sterols, sesquiterpene lactones, and other constituents [8,18]. The chemical composition of B. balsamifera oils varies from different plant populations [22,25,26,28], indicating more studies should be done on individual's structural development, the cultivation, geographical, and climate conditions and essential oil standardization. The survey and summary of the extensive studies revealed that B. balsamifera was an essential and valuable medicinal plant used for folk treatments such as treating eczema, dermatitis, beriberi, lumbago, menorrhagia, rheumatism, skin injury, or used as insecticide [45]. As a traditional medicine, the biological and pharmacological studies of the plant materials, crude extracts, and isolated chemical constituents of B. balsamifera offered experimental and scientific proofs for its various traditional uses. The pharmacological studies focused on studying the anti-microbial and anti-inflammatory effects [13,15], antiplasmodial effects [75], platelet aggregation [79], wound healing [77], and disease and insect resistant activities [26], all of which confirmed the plant's traditional uses. Moreover, some new pharmacological uses were discovered, such as antitumor [12–14], hepatoprotective [66], superoxide radical scavenging [16], antioxidant [69], antityrosinase [14], enhancing percutaneous penetration [76], and anti-obesity activities [17]. However, there was no experimental and pharmacological evidence to prove the traditional uses of this plant in rheumatism and lumbago. Besides, the isolated chemical constituent and its correspondent pharmacological effects were rarely simultaneously carried out in one study. Hasegawa et al. extracted a dihydroflavonol from B. balsamifera, which exhibited a significant synergism with TRAIL by pharmacological experiment [12]. Nevertheless, more studies should be done in abundance to understand the pharmacodynamic chemical constituents. The outcome from this study could establish the basis for its future clinical utilization in modern science.

On the basis of the above review, several prospects were revealed. In order to further define the effective chemical compounds, the biological activities of monomeric compounds, the plant material, and its crude extraction further studies were proposed. In addition, some traditional effects of *B. balsamifera*, such as rheumatism still need to be testified by more modern methods and further pharmacological trials. Few more aspects such as pharmacokinetics, molecular biology, and naturel medicinal chemistry should be utilized to study its phytochemical standardization and bioactivity identification according to its bioactive metabolism. Moreover, in the production process of TCM Aipian, *B. balsamifera* oil and Aifen could be important accessory substances yielded, which also

possess many chemical and biological activities to be further studied in the future. Therefore, we concluded that the area of *B. balsamifera* research should be significantly expanded.

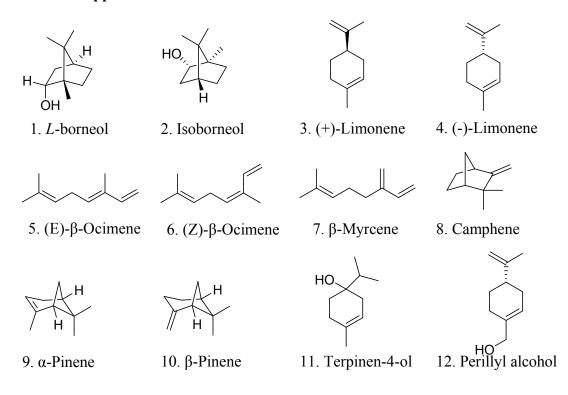
Acknowledgments

We thank Dr. Lingliang Guan for grammar correction, Mr. Yingbo Zhang for valuable comments, Qing Wen and all other sixteen master degree candidates for modifying the spelling and grammar mistakes in this review. This project was supported by the Natural Science Fund of Hainan Province, China (Grant No. 312022) and the Natural Science Fund of China (Grant No. 81374065, 81303171). We are grateful to Professor Chen Song-bi (Tropical Crops Genetic Resources Institute, Chinese Academy of Tropical Agricultural Sciences) for critical reviewing and revision of the manuscript.

Author Contributions

Dan Wang: Acquisition, interpretation of data and wrote the manuscript. References management. Obtained funding; Zuowang Fan: Draw the structural formulas. Classified the chemical components. Xiaolu Chen: References management. Revising the review critically for important intellectual content. Fulai Yu: Obtained funding. Xuan Hu: References management. Participate in drafting the article. Kai Wang: Drafted the structural formulas. Lei Yuan: Drafted the structural formulas. Yuxin Pang: Contributed to conception and design of the review. Obtained funding. Overall responsibility.

Appendix 1. The structures of the volatile chemical constituents.



Appendix 1. Cont.

13. Chrysanthenone

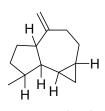


17. α-Thujene



21. 1,8-Cineole

25. Thymohydroquinone dimethyl ether



29. Aromadendrene

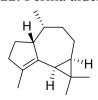
33. α-Caryophyllene

37. γ-Cadinene

14. α-Terpineol

18. Trans-linalool oxide

22. Perilla aldehyde



26. α-Gurjunene



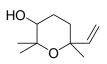
30. Aromadendrene oxide

34. β-Caryophylene

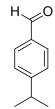
38. δ-Cadinene



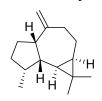
15. Bornyl acetate



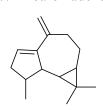
19. Linalool oxide



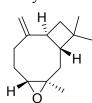
23. Cuminaldehyde



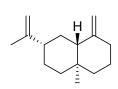
27. Alloaromadendrene



31. Aromadendrene, dehydro



35. Caryophyllene oxide



39. β-Selinene



16. Sabinene



20. Camphor



24. Myrtenal



28. (+)-Aromadendrene

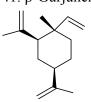


32. Longifolene

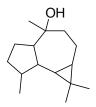
36. Guaia-3,9-diene

40. α-Selinene

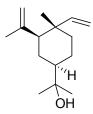
Appendix 1. Cont.



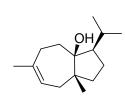
45. α-Elemene



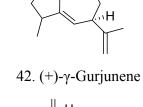
49. Globulol



53. Elemol

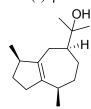


57. Carotol

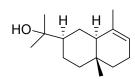




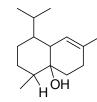
46. (-)-γ-Cadinene



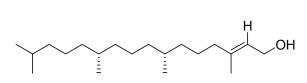
50. (-)-Guaiol



54. α-Eudesmol



58. Cubenol



61. Phytol

64. Blumeaene C

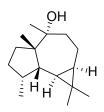
65. Blumeaene D



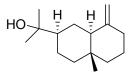
43. Thujopsene-13



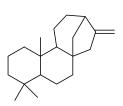
47. (-)-δ-Cadinene



51. Ledol



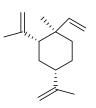
55. β-Eudesmol



59. 16-Kaurene

62. Blumeaene A

66. Blumeaene E



44. β-Elemene

48. 10-epi-γ-Eudesmol

52. γ-Murolene

56. γ-Eudesmol

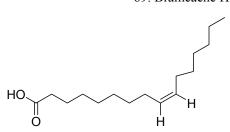
60. 1-Ang-4,7-dihydroxy eudesmane

63. Blumeaene B

67. Blumeaene F

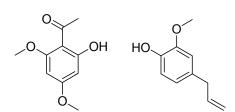
Appendix 1. Cont.

69. Blumeaene H



72. (11Z)-11-Hexadecenoic acid

75. Capric acid

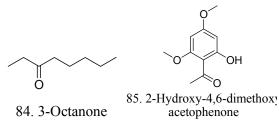


77. Xanthoxylin

78. Eugenol

80. 1-Octen-3-ol

81. 3-Octanol



84. 3-Octanone

88. 1,3,4,5,6,7-hexahydro-2,5,5-trimethyl-2H-2,4a -ethanonaphthalene

70. Blumeaene I

71. Blumeaene J

$$HO \longrightarrow HO \longrightarrow H$$

73. Trans-2-undecenoic acid

74. 9-Hexadecenoic acid

76. Palmitic acid

79. 1,4-Dimethoxy-2,3,5,6tetramethylbenzene

82. 3-Propyl benzaldehyde

86. 3-Fluoro-5-aminopyridine



89. 4-isopropyltoluene

83. 4-Isopropyl benzaldehyde

87. 4,5-diethy-2,3-dihydro-2,3-dimethylfuran

90. 3-Nitrophthalic acid

Appendix 1. Cont.

Appendix 2. The structures of the non-volatile chemical constituents.

A. 1–5 are flavones. B. 6–21 are flavonols.

$$R_3$$
 R_4 R_5 R_6 R_6 R_6

R1 R4 R5 R6 R2 R3 1. 4',5-Dihydroxy-7-methyletherflavanone -H -H -OMe -H -OH -H -H -H -OH -OH -H 2. Luteolin -OH 3. Luteolin-7-methyl-ether -H -H -ОН -OH -H -OMe -H -H 4. Diosmetin -H -OH -OH -OMe -H 5. Chrysoeriol (Luteolin 3'-methyl ether) -H -H -OMe -OH -OH -OH -H 6. Quercetin -Н -OH -OH -OH -H 7. 3,5,3',4'-Tetrahydroxy-7-methoxyflavone -ОН -H -OMe -OH -ОН -H 8. 3,5,3'-Trihydroxy-7,4'-dimethoxyflavone -H -OH -OMe -OH -OMe -H 9. Rhamnetin -OH -OH -H -OMe -OH 10. Tamarixetin -H -OH -H -OH -OH -OMe -H 11. Ombuine -ОН -H -OMe -OH -OMe 12. 3,5,7-Trihydroxy-3'4'-dimethoxyflavone -OH -Н -ОН -OMe -OMe -H 13. 3,3',4',5-Tetrahydroxy-7-methoxyflavone -OH -Н -OMe -OH -OH -Н 14. 3,5-Dihydroxy-3',4',7-trimethoxyflavone -OH -Н -OMe -OMe -OMe -H -H 15. 4',5-Dihydroxy-3,3',7-trimethoxy flavanone -OMe -H -OMe -OMe -ОН 16. 5,7-Dihydroxy-3,3',4',-trimethoxyflavone -OMe -Н -ОН -OMe -OMe -H -OMe -Н -OH -OMe -H 17. Ayanin -OMe -OMe -H 18. Chrysosplenol C -OMe -OH -OMe -OH 19. 4',5,7-Trihydroxy-3,3'-dimethoxyflavone -OMe -ОН -H -OMe -Н -ОН

20. Hyperoside

HO OH OH OH

21. Isoquercitrin

Appendix 2. Cont.

C. 22–25 are flavanones.

D. 26-33 are flavanonols.

$$R_2$$
 R_3
 R_4
 R_5
 R_1
 R_5

	R1	R2 R3	R4	R5
22. Bumeatin	-H	-OMe -OH	-H	-ОН
23. Eriodictyol	-H	-ОН -ОН	-ОН	-Н
24. 5,7,3',5'-Tetrahydroxyflavanone	- H	-ОН -ОН	-H	-ОН
25. 3',4',5-Trihydroxy-7-metoxyflavanone	-H	-OMe -OH	-ОН	-Н
26. Dihydroquercetin-4'-methylether	-OH	-ОН -ОН	-OMe	-Н
27. Dihydroquercetin-7,4'-dimethylether	-OH	-OMe -OH	-OMe	-Н
28. 3,5,4'-Trihydroxy-3',7-dimethoxyflavanone	-OH	-OMe -OMe	-OH	-Н
29. 3,3',5,5',7-Pentahydroxyflavanone	-OH	-ОН -ОН	- H	-ОН
30. 3,3',4',5-Tetrahydroxy-7-methoxyflavanone	-OH	-OMe -OH	-OH	-Н
31. 3,3',5-Trihydroxy-4',7-dimethoxyflavanone	-OH	-OMe -OH	-OMe	-Н
32. 3,3',5,7-Tetrahydroxy-4'-methoxyflavanone	-OH	-ОН -ОН	-OMe	-Н
33. 3',4',5-Trihydroxy-3,7-dimetoxyflavanone	-OMe	-OMe -OH	-OH	-Н

E. 34–35 are chalcones.

F. 36-37 are flavanols

37. (2R,3R)-(+)-7-*O*-methyl dihydroquercetin

Appendix 2. Cont.

H. 41–43 are steroids.

I. 44 is diterpene, 45–46 are triterpenes.

44. Cryptomeridiol

Appendix 2. Cont.

45. 3,13-Clerodadiene-6,15-diol

46. Austronulin

G. 47 is lignan; 48, 49 are coumarins; 50 is naphthalenone.

50. 5,7-Dihydroxychromone

Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Singh, A.; Singh, D.K. Molluscicidal activity of *Lawsonia inermis* and its binary and tertiary combinations with other plant derived molluscicides. *Indian J. Exp. Biol.* **2001**, *39*, 263–268.
- 2. Bhathena, S.J.; Velasquez, M.T. Beneficial role of dietary phytoestrogens in obesity and diabetes. *Am. J. Clin. Nutr.* **2002**, *76*, 1191–1201.
- 3. Canter, P.H.; Thomas, H.; Ernst, E. Bringing medicinal plants into cultivation: Opportunities and challenges for biotechnology. *Trends Biotechnol.* **2005**, *23*, 180–185.
- 4. Chinese Academy of Sciences editorial commission of the flora. *Flora Republicae Popularis Sinicae*; Sciences Press: Beijing, China, 1988.
- 5. Yuan, Y.; Pang, Y.X.; Wang, W.Q.; Zhang, Y.B.; Yu, J.B.; Zhu, M. Research advances in the genus of *Blumea* (inuleae) phylogenetic classification. *Chin. J. Trop. Agric.* **2011**, *31*, 81–86.
- 6. Pang, Y.X.; Wang, W.Q.; Zhang, Y.B.; Mo, T.H.; Yuan, Y. Clonal diversity and structure in natural populations of *Blumea balsamifera*. *Guihaia* **2010**, *30*, 209–214.

7. Yuan, Y.; Pang, Y.X.; Wang, W.Q.; Zhang, Y.B.; Yu, J.B. Investigation on the plants resources of *Blumea balsamifera* (L.) DC. in China. *J. Trop. Org.* **2011**, *2*, 78–82.

- 8. Guan, L.L.; Pang, Y.X.; Wang, D.; Zhang, Y.B.; Wu, K.Y. Research progress on Chinese Minority Medicine of *Blumea balsamifera* L. DC. *J. Plant Genet. Res.* **2012**, *13*, 695–698.
- 9. Chen, M. Studies on the Active Constituents of *Blumea balsamifera*. Master Degree, Shanghai Jiaotong University, Shanghai, China, 2009.
- 10. Chen, M.; Qin, J.J.; Fu, J.J.; Hu, X.J.; Liu, X.H.; Zhang, W.D.; Jin, H.Z. Blumeaenes A–J, sesquiterpenoid esters from *Blumea balsamifera* with NO inhibitory activity. *Planta Med.* **2010**, 76, 897–902.
- 11. Chinese Pharmacopeia Commission. *Pharmacopoeia of the People's Republic of China*, 2010 ed.; The medicine science and technology press of China: Beijing, China, 2010.
- 12. Hasegawa, H.; Yamada, Y.; Komiyama, K.; Hayashi, M.; Ishibashi, M.; Yoshida, T.; Sakai, T.; Koyano, T.; Kam, T.S.; Murata, K.; *et al.* Dihydroflavonol BB–1, an extract of natural plant *Blumea balsamifera*, abrogates TRAIL resistance in leukemia cells. *Blood* **2006**, *107*, 679–688.
- 13. Li, J.; Zhao, G.Z.; Chen, H.H.; Wang, H.B.; Qin, S.; Zhu, W.Y.; Xu, L.H.; Jiang, C.L.; Li, W.J. Antitumour and antimicrobial activities of endophytic streptomycetes from pharmaceutical plants in rainforest. *Lett. Appl. Microbiol.* **2008**, *47*, 574–580.
- 14. Saewan, N.; Koysomboon, S.; Chantrapromma, K. Anti-tyrosinase and anti-cancer activities of flavonoids from *Blumea balsamifera* DC. *J. Med. Plants. Res.* **2011**, *5*, 1018–1025.
- 15. Ragasa, C.Y.; Co, A.L.K.C.; Rideout, J.A. Antifungal metabolites from *Blumea balsamifera*. *Nat. Prod. Res.* **2005**, *19*, 231–237.
- 16. Nessa, F.; Ismail, Z.; Mohamed, N.; Haris, M.R.H.M. Free radical–scavenging activity of organic extracts and of pure flavonoids of *Blumea balsamifera* DC leaves. *Food Chem.* **2004**, *88*, 243–252.
- 17. Kubota, H.; Kojima-Yuasa, A.; Morii, R.; Huang, X.; Norikura, T.; Rho, S.N.; Matsui-Yuasa, I. Anti-obesity effect of *Blumea balsamifera* extract in 3T3–L1 preadipocytes and adipocytes. *Am. J. Chin. Med.* **2009**, *37*, 843–854.
- 18. Chen, M.; Jin, H.Z.; Zhang, W.D.; Yan, S.K.; Shen, Y.H. Chemical constituents of plants from the genus *Blumea*. *Chem. Biodiver.* **2009**, *6*, 809–817.
- 19. State Administration of Traditional Chinese Medicine. *Chinese Materia Medica*; Scientific and Technical Publishers: Shanghai, China, 1999.
- 20. Chinese Academy of Sciences editorial commission of the flora. *Flora Republicae Popularis Sinicae*; Sciences Press: Beijing, China, 1979.
- 21. Zhou, X.; Yang, X.S.; Zhao, C. Chemical components of volatile oil from folium *et* cacumen *Blumea balsamifera* originated from Guizhou. *J. Instrem. Anal.* **2001**, *20*, 76–78.
- 22. Du, P.; Zhang, X.J.; Sun, X.D. Chemical constituents of volatile oil from *Blumea balsamifera* (Linn.) DC. in Yunnan. *Chem. Ind. For. Prod.* **2009**, *29*, 115–118.
- 23. Wang, Y.H.; Tian, H.Y.; He, S.J.; Hu, Q.P.; Wang, H.X.; Zhou, C.L.; Wang, X. Analysis of volatile components from leaf in *Blumea balsamifera* (L.) DC. with different extraction methods by gas chromatography–mass spectrometry. *Sci. Tech. Food Ind.* **2012**, *33*, 97–105.
- 24. Liu, J.X.; Wang, K.X.; Li, C.P. The questions and solutions in extraction of volatile oils from Traditional Chinese Medicine by vapour distillation. *Lishizhen Med. Mater. Med. Res.* **2008**, *19*, 97–98.

25. Hao, X.Y.; Yu, Z.; Ding, Z.H. The study on chemical constituents of volatile oil on *Blumea balsamifera* growing in Guizhou. *Guiyang Yi Xue Yuan Xue Bao* **2000**, *25*, 121–122.

- 26. Chu, S.S.; Du, S.S.; Liu, Z.L. Fumigant compounds from the essential oil of Chinese *Blumea balsamifera* leaves against the maize weevil (*Sitophilus zeamais*). *J. Chem.* **2013**, *2013*, 1–7.
- 27. Gao, Y.Q. The Chemical Composition and Biological Activities of Volatile Oils of Nineteen Plants. PhD. Thesis, Guizhou University, Guiyang, China, 2007.
- 28. Bhuiyan, N.I.; Chowdhury, J.U.; Begum, J. Chemical components in volatile oil from *Blumea balsamifera* (L.) DC. *Bangladesh J. Botany* **2009**, *38*, 107–109.
- 29. Zhu, Y.C.; Wen, Y.X.; Wang, H.S.; Huang, Y.L. Chemical constituents in *Blumea balsamifera* (I). *Guihaia* **2008**, *28*, 139–141.
- 30. Zhu, T.C. Study on Chemical Constituents of the Extracts of Ethyl Acetate from *Blumea balsamifera* DC. Master Thesis, Guangxi Normal University, Guilin, China, 2007.
- 31. Ali, D.M.; Wong, K.C.; Lim, P.K. Flavonoids from *Blumea balsamifera*. *Fitoterapia* **2005**, *76*, 128–130.
- 32. Bai, Z.W.; Zhu, L.; Lu, Y.; Su, J.P. The research progress of Miao medicine *Blumea balsamifera* (L.) DC. *J. Med. Pharm. Chin. Minor.* **2012**, *18*, 65–67.
- 33. He, Y.; Chai, L.; Ding, Y.; Xian, F.; PAN, J. Effect of N nutrition on yield and active ingredient in *Blumea balsamifera*. *Guizhou Agri. Sci.* **2006**, *34*, 28–30.
- 34. Wen, Y.X.; Huang, Y.L.; Zhu, Y.C.; Zhao, Z.G. Analysis of Xanthoxylin in different section of *Blumea balsamifera* DC. by RP–HPLC. *Lishizhen Med. Mater. Med. Res.* **2007**, *18*, 2137–2138.
- 35. Xia, J.Z.; Zhao, Z.; An, J.; Deng, Y.H. GC fingerprint of herba *Blumea* from different habitats. *Zhong Guo Yao Shi* **2011**, *12*, 1191–1194.
- 36. Sun, Y.M. The methodology validation of the determination of *Blumea balsamifera* oil. *Guizhou Med. J.* **2011**, *35*, 757–759.
- 37. Shirota, O.; Oribello, J.M.; Sekita, S.; Satake, M. Sesquiterpenes from *Blumea balsamifera*. *J. Nat. Prod.* **2011**, *74*, 470–476.
- 38. Xu, J.; Jin, D.Q.; Liu, C.; Xie, C.; Guo, Y.; Fang, L. Isolation, characterization, and NO inhibitory activities of sesquiterpenes from *Blumea balsamifera*. *J. Agric. Food Chem.* **2012**, *60*, 8051–8058.
- 39. Liang, H.L.; Cao, P.X.; Qiu, J.Y.; Li, X.; Cai, L.; Liang, G.Y. Study on the chemical constituents of *Blumea balsamifera* DC. *Lishizhen Med. Mater. Med. Res.* **2011**, *22*, 308–309.
- 40. Pang, Y.X.; Wang, D.; Yuan, Y.; Yu, F.L.; Wu, K.Y.; Di, M. Study on the extraction technology of total flavonoids from *Blumea balsamifera* (L.) DC. *Chin. J. Trop. Crop.* **2013**, *34*, 168–170.
- 41. Ruangrungsi, N.; Tappayuthpijarn, P.; Tantivatana, P.; Borris, R.P.; Cordell, G.A. Traditional Medicinal Plants of Thailand. I. Isolation and structure elucidation of two new flavonoids, (2*R*,3*R*)-dihydroquercetin-4'-methyl ether and (2*R*,3*R*)-dihydroquercetin-4',7-dimethyl ether from *Blumea balsamifera*. *J. Nat. Prod.* **1981**, *44*, 541–545.
- 42. Lin, Y.C.; Long, K.H.; Deng, Y.J. Studies on the chemical constituents of the Chinese medicinal plant *Blumea balsamifera*. *Acta Sci. Natur. Univ. Sunyaatseni* **1988**, *2*, 77–81.
- 43. Nessa, F.; Mohamed, N.; Ismail, Z.; Haris, M.R.H.M. Phytochemical investigation on *Blumea balsamifera* DC. *J. Trop. Med. Plants* **2001**, *2*, 17–22.

44. Nessa, F.; Ismail, Z.; Karupiah, S.; Mohamed, N. RP-HPLC method for the quantitative analysis of naturally occurring flavonoids in leaves of *Blumea balsamifera* DC. *J. Chromatogr. Sci.* **2005**, 43, 416-420.

- 45. Chen, M.; Jin, H.Z.; Yan, L.; Hu, X.J.; Qin, J.J.; Liu, J.H.; Yan, S.K.; Zhang, W.D. Flavonoids from *Blumea balsamifera*. *Nat. Prod. Res.* **2010**, 991–994.
- 46. Huang, Y.L.; Zhu, T.C.; Wen, Y.X.; Wang, H.S.; Chen, Y.Y. Isolation and identification of chemical constituents from *Blumea balsamifera*. *Guihaia* **2010**, *30*, 560–562.
- 47. Deng, Q.Y.; Ding, C.M.; Zhang, W.H.; Lin, Y.C. Studies on the flavonoid constituents in *Blumea balsamifera*. *Chin. J. Magn. Resonance* **1996**, *5*, 447–452.
- 48. Yan, Q.X.; Tan, D.P.; Kang, H.; Feng, H.L.; Zeng, W.Z. Study on flavonoids constituents of *Blumea balsamifera. Chin. J. Exp. Tradit. Med. Form.* **2012**, *18*, 86–89.
- 49. Tan, D.P.; Yan, Q.X.; Kang, H.; Zeng, W.Z.; Feng, H.L. Chemical constituents of *Blumea balsamifera* DC. *Nat. Prod. Res. Dev.* **2012**, *24*, 718–721.
- 50. Huang, Y.L.; Wen, Y.X.; Zhao, Z.G.; Zhu, T.C. Determination of flavanones and flavanonols in *Blumea balsamifera* DC. by RP–HPLC. *Guangxi Sci.* **2007**, *14*, 140–142.
- 51. Huang, Y.L.; Zhao, Z.G.; Wen, Y.X. Determination of total flavonoid in different sections of *Blumea balsamifera. Guihaia* **2006**, *26*, 453–455.
- 52. Nessa, F.; Mohamed, N.; Ismail, Z. Superoxide radical scavenging properties of extracts and flavonoids isolated from the leaves of *Blumea balsamifera*. *Pharm. Biol.* **2005**, *43*, 15–20.
- 53. Ishibashi, M.; Ohtsuki, T. Studies on search for bioactive natural products targeting TRAIL signaling leading to tumor cell apoptosis. *Med. Res. Rev.* **2008**, *28*, 688–714.
- 54. Fujimoto, Y.; Soemartono, A.; Sumatra, M. Sesquiterpene lactones from *Blumea balsamifera*. *Phytochemistry* **1988**, *27*, 1109–1111.
- 55. Osaki, N.; Koyano, T.; Kowithayakorn, T.; Hayashi, M.; Komiyama, K.; Ishibashi, M. Sesquiterpenoids and plasmin-inhibitory flavonoids from *Blumea balsamifera*. *J. Nat. Prod.* **2005**, *68*, 447–449.
- 56. Zhao, J.H.; Kang, H.; Yao, G.H.; Zeng, W.Z. The study of chemical constituents of *Blumea balsamifera* DC. *Chin. Trad. Herb. Drug.* **2007**, *38*, 350–352.
- 57. Chadwick, M.; Trewin, H.; Gawthrop, F.; Wagstaff, C. Sesquiterpenoids Lactones: Benefits to Plants and People. *Int. J. Mol. Sci.* **2013**, *14*, 12780–12805.
- 58. Ghantous, A.; Gali-Muhtasib, H.; Vuorela, H.; Saliba, N.A.; Darwiche, N. What made sesquiterpene lactones reach cancer clinical trials? *Drug Discov Today* **2010**, *15*, 668–678.
- 59. Arifin, H.; Widianingsih, I.; Marusin, N.; Andalas, J.F.F.M. Pengaruh pemberian akut ekstrak etanol daun Capo (*Blumea balsamifera* DC) terhadap gambaran morfolofis dan histologi hati mencit putih jantan. *J. Sains Tek. Far.* **2007**, *12*, 82–88.
- 60. Norikura, T.; Kojima-Yuasa, A.; Shimizu, M.; Huang, X.; Xu, S.; Kametani, S.; Rho, S.N.; Kennedy, D.O.; Matsui-Yuasa, I. Anticancer activities and mechanisms of *Blumea balsamifera* extract in hepatocellular carcinoma cells. *Am. J. Chin. Med.* **2008**, *36*, 411–424.
- 61. Norikura, T.; Kojima-Yuasa, A.; Shimizu, M.; Huang, X.; Xu, S.; Kametani, S.; Rho, S.; Kennedy, D.O.; Matsui-Yuasa, I. Mechanism of growth inhibitory effect of *Blumea balsamifera* extract in hepatocellular carcinoma. *Biosci. Biotechnol. Biochem.* **2008**, *72*, 1183–1189.

62. Ng, K.W.; Salhimi, S.M.; Majid, A.M.; Chan, K.L. Anti–angiogenic and cytotoxicity studies of some medicinal plants. *Planta Med.* **2010**, *76*, 935–940.

- 63. Lee, C.Y. Medication Combination for Hepatoma and Pancreatic Cancer and Preparation Protocol. U.S. Patent US6998141 B2, 14 February 2006.
- 64. Xu, S.Y.; Bian, R.L.; Chen, X. *Methodology of Pharmacological Experiments*, 2nd ed.; People's Medical Publishing House: Beijing, China, 1994.
- 65. Xu, S.B.; Zhao, J.H. Protective actions of *Blumea* flavanones on experimental liver injury. *Chin. Phar. Bull.* **1998**, *14*, 191–192.
- 66. Pu, H.L.; Zhao, J.H.; Xu, S.B.; Hu, Q. Protective actions of *Blumea* flavanones on primary cultured hepatocytes against lipid peroxidation. *Chin. Trad. Herb. Drug.* **2000**, *31*, 1113–1115.
- 67. Xu, S.B.; Hu, Y.; Lin, Y.C.; Yang, Z.B. Study on protection of blumeatin against experimental liver injury and aggregation of platelet. *Suppl. J. Sun Yatsen Univer.* **1994**, *1994*, 48–53.
- 68. Zhao, J.H.; Xu, S.B. Effects of *Blumea* flavanones on lipid peroxidation and active oxygen radicals. *Chin. Phar. Bull.* **1997**, *5*, 17.
- 69. Nguyen, M.T.; Awale, S.; Tezuka, Y.; Tran, Q.L.; Watanabe, H.; Kadota, S. Xanthine oxidase inhibitory activity of Vietnamese medicinal plants. *Biol. Pharm. Bull.* **2004**, *27*, 1414–1421.
- 70. Nguyen, M.T.; Nguyen, N.T. Xanthine oxidase inhibitors from Vietnamese *Blumea balsamifera* L. *Phytother. Res.* **2012**, *26*, 1178–1181.
- 71. Nessa, F.; Ismail, Z.; Mohamed, N. Xanthine oxidase inhibitory activities of extracts and flavonoids of the leaves of *Blumea balsamifera*. *Pharm. Biol.* **2010**, *48*, 1405–1412.
- 72. Shyur, L.; Tsung, J.; Chen, J.; Chiu, C.; Lo, C. Antioxidant properties of extracts from medicinal plants popularly used in Taiwan. *Int. J. Appl. Sci. Eng.* **2005**, *3*, 195–202.
- 73. Ongsakul, M.; Jindarat, A.; Rojanaworarit, C. Antibacterial effect of crude alcoholic and aqueous extracts of six medicinal plants against *Staphylococcus aureus* and *Escherichia coli*. *J. Health Res.* **2009**, *23*, 153–156.
- 74. Sakee, U.; Maneerat, S.; Cushnie, T.P.; De-Eknamkul, W. Antimicrobial activity of *Blumea balsamifera* (Lin.) DC. extracts and essential oil. *Nat. Prod. Res.* **2011**, *25*, 1849–1856.
- 75. Noor, R.A.; Khozirah, S.; Mohd, R.M.; Ong, B.K.; Rohaya, C.; Rosilawati, M.; Hamdino, I.; Badrul, A.; Zakiah, I. Antiplasmodial properties of some Malaysian medicinal plants. *Trop. Biomed.* **2007**, *24*, 29–35.
- Fu, W.J.; Wang, D.; Pang, Y.X.; Wang, H.; Wang, Z.; Nie, H.; Yu, F.L.; Zhang, Y.B. Effect of Blumea balsamifera oil on percutaneous absorption of salbutamol sulfate. Chin. J. Exp. Tradit. Med. Form. 2013, 19, 174–177.
- 77. Wang, D.; Fu, W.J.; Pang, Y.X.; Wang, H.; Hu, X.; Nie, H. The study of skin allergy and acute toxicity of *Blumea balsamifera* oil. *Chin. J. Trop. Crop.* **2013**, *34*, 2499–2502.
- 78. Luo, Y.; Zheng, F.; Yang, Y. Antifungal activities of 128 southern Chinese herbs against 6 pathogens. *Chin. J. Trop. Crop.* **2004**, *25*, 106–111.
- 79. Huang, T.K.; Ding, Z.Z.; Zhao, S.X. *Modern Compendium of Materia Medica*; Chinese Medical Science and Technology Press: Beijing, China, 2000.
- © 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).