Supporting Material

Trehalose-rich, degradable hydrogels designed for trehalose release under physiologically relevant conditions

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1. Materials and general methods

Acrylamide (AM), acryloyl chloride, amberlyst-15, ammonia aqueous, ammonium persulphate (APS), 2,2'-Azobis[2-(2-imidazolin-2-yl)propane] dihydrochloride (V-044), Celite 501, hydrochloric acid, hydroquinone, *p*-hydroxybenzaldehyde, lithium tetrafluoroborate, methanesulfonyl chloride, phosphate buffered saline (PBS) tablets, polyethylene glycol (PEG₄₀₀), pyridine, silver(I) oxide (Ag₂O), sodium hydroxide, *N*,*N*,*N'*,*N'*-tetramethylethylenediamine (TMEDA), *p*-toluenesulfonic acid, tosyl chloride, anhydrous trehalose, triethylamine (TEA), and trimethyl orthoformate, trimethylsilyl chloride were purchased from Sigma Aldrich, Acros Organics, TCI or Fluka and used directly without any purification. *N*,*N*-dimethylacrylamide (Sigma Aldrich) were purified by passing through a column filled with basic aluminum oxide to remove inhibitor. Anhydrous solvents were purchased from Acros Organics and stored over molecular sieves under inert atmosphere. All other solvents and inorganic salts were purchased from Avantor Performance Materials Poland S.A. Trehalose Assay Kit was purchased from Megazyme International Ireland.

p-(2-Hydroxyethoxy)benzaldehyde was synthesized according to the published procedure¹.

Purifications by flash column chromatography were performed on silica gel 60 (40-63 μ m, Merck) using automated system (Isolera, Biotage). Freeze-drying was carried out under 0.035 mbar at -50 °C (ALPHA 1-2 LDplus, CHRIST). Mass spectra were recorded using Electrospray Ionisation Mass Spectrometry (QTRAP 4000, AB Sciex). NMR spectra were recorded in deuterated solvents (Deutero GmbH) with internal standards using NMR spectrometer operating at 400 or 600 MHz (Varian).

2. Synthesis and characterization of trehalose monomers

2.1. Synthetic procedure for MT1 and DT1



Scheme S1 Synthetic pathway to trehalose monomers DT1 and MT1.

p-(Hydroxy(polyethylenoxy))benzaldehyde



p-(2-Hydroxy(polyethylenoxy))benzaldehyde was synthesized in two step reaction starting from PEG₄₀₀, based on the slightly modified literature procedures^{2,3}.

Briefly, PEG₄₀₀ (10.3 mmol) was dried by repeated evaporation with toluene under vacuum and then dissolved in 125 mL of anhydrous toluene. KI (2.1 mmol) and Ag₂O (15.4 mmol) were added and magnetic stirring was set to 1000 rpm before tosyl chloride (10.8 mmol) was quickly introduced in one portion. The reaction was left overnight and resulting suspension was then filtrated through the pad of Celite 501. Toluene was evaporated under vacuum to afford tosylated PEG₄₀₀ as a viscous, colorless oil, which was subjected to the second step without further purification. After dissolving in 25 mL of anhydrous DMF, *p*-hydroxybenzaldehyde (15.5 mmol) and K₂CO₃ (20.6 mmol) were added and the reaction was left overnight under stirring at 90 °C. Afterwards, DMF was evaporated under vacuum, 50 mL of 5% Na₂CO₃ aqueous solution was added to the residue and the mixture was extracted with DCM (2 x 50 mL). The combined organic layers were washed with 5% Na₂CO₃ aqueous solution (2 x 50 mL) and brine (50 mL), dried over MgSO₄ and concentrated under vacuum to afford product as a yellow viscous oil (4.45 g). As calculated from ¹H NMR spectrum, the product was found to contain 76.3 mol%

and 23.7 mol% of mono- and disubstituted PEG_{400} derivatives, respectively. The product was taken forward without further purification. Monosubstituted PEG_{400} derivative was separated from disubstituted PEG_{400} derivative after next step (acetalization with trimethylorthoformate) by silica gel column chromatography.

¹H NMR* (DMSO-d₆, 600 MHz,) δ [ppm]: 3.39-3.42 (m, 2H, H_h); 3.44-3.63 (m, ~29 H, H_g, PEG backbone); 3.74-3.80 (m, 2H, H_f); 4.19-4.24 (m, 2H, H_e); 4.53-4.60 (m, 1H, –OH); 7.10-7.17 (m, 2H, H_c); 7.83-7.89 (m, 2H, H_b); 9.86 (br s, 1H, –CHO). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 60.20 (C_h); 67.68, 68.69, (C_e, C_f); 69.66-69.96 (PEG backbone); 72.33 (C_g); 114.95 (C_c);; 129.64 (C_a); 131.78 (C_b) 163.48 (C_d). 191.29 (– CHO).

*with respect to monosubstituted PEG_{400} derivative. Molar ratio of mono- to disubstituted PEG_{400} derivative: (1 : 0.31)

p-(Hydroxy(polyethylenoxy))benzaldehyde dimethyl acetal



p-(Hydroxy(polyethylenoxy))benzaldehyde dimethyl acetal was synthetized based on the reported procedure.⁴ To a solution of p-(hydroxy(polyethylenoxy))benzaldehyde (4.40 g) and lithium tetrafluoroborate (0.9 mmol) in methanol (20 mL), trimethyl orthoformate (17.6 mmol) was introduced, and the solution was stirred under reflux for 21h. After cooling, pyridine (0.19 mmol) and saturated aqueous solution of NaHCO₃ (50 mL) was added, and the mixture was extracted twice with DCM (70 mL). Organic layer was then washed with saturated aqueous solution of NaHCO₃ (40 mL), dried over MgSO₄ and concentrated under vacuum. The crude product was purified by silica gel flash chromatography (CHCl₃ : TEA 99:1) to afford product as a yellowish oil (3.68 g, 65%*).

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.21 (s, 6H, $-CH_3$); 3.39-3.43 (m, 2H, H_h); 3.45-3.60 (m, ~29 H, H_g, PEG backbone); 3.72-3.75 (m, 2H, H_f); 4.06-4.10 (m, 2H, H_e); 4.54-4.58 (m, 1H, -OH); 5.31 (s, 1H, -OCHO-); 6.91-6.95 (m, 2H, H_c); 7.26-7.31 (m, 2H, H_b). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 52.31 ($-CH_3$); 60.20 (C_h); 67.09, 68.90, (C_e, C_f); 69.70-69.98 (PEG backbone); 72.33 (C_g); 102.47 (-OCHO-); 113.94 (C_c); 127.77 (C_b); 130.35 (C_a); 158.40 (C_d).*after 3 steps

p-(Acryloyloxy(polyethylenoxy))benzaldehyde dimethyl acetal



A solution of p-(hydroxy(polyethylenoxy))benzaldehyde dimethyl acetal (6.5 mmol) and TEA (14.6 mmol) in THF (80 mL) was cooled on ice water bath, and then a solution of acryloyl chloride (9.8 mmol) in THF (5 mL) was added dropwise. The mixture was left to warm up to room temperature and the reaction was finished after 3 h. Precipitated triethylamine hydrochloride was separated on a pad of Celite 501, and filtrate was concentrated under vacuum. The crude product was purified by silica gel flash

chromatography (CHCl₃ : TEA 99:1) to afford p-(acryloyloxy(polyethylenoxy))benzaldehyde dimethyl acetal as a yellowish syrup (3.61 g, 92%).

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.21 (s, 6H, –CH₃); 3.48-3.60 (m, ~27 H, PEG backbone); 3.62-3.66 (m, 2H, H_g); 3.72-3.75 (m, 2H, H_f); 4.06-4.10 (m, 2H, H_e); 4.19-4.23 (m, 2H, H_h); 5.31 (s, 1H, –OCHO–); 5.93-5.97 (m, 1H, –CH=CH₂ *cis*); 6.16-6.22 (m, 1H, –CH=CH₂); 6.30-6.36 (m, 1H, –CH=CH₂ *trans*); 6.90-6.95 (m, 2H, H_c); 7.26-7.30 (m, 2H, H_b). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 52.31 (–CH₃); 63.45 (C_g); 67.09, 68.91, (C_e, C_f); 68.21 (C_h); 69.65-69.95 (PEG backbone); 102.46 (–OCHO–); 113.93 (C_c); 127.77 (C_b); 128.21 (–CH=CH₂); 130.35 (C_a); 131.66 (–CH=CH₂); 158.41 (C_d); 165.47 (–OC(O)–).

$4,6-O-[p-(acryloyloxy(polyethylenoxy))benzylidene]-\alpha,\alpha'-D-trehalose$ (MT1) and $4,6:4',6'-di-O-[p-(acryloyloxy(polyethylenoxy))benzylidene]-\alpha,\alpha'-D-trehalose$ (DT1)



To a suspension of anhydrous trehalose (2.4 mmol) in anhydrous DMF (5 mL), p-(acryloyloxy(polyethylenoxy))benzaldehyde dimethyl acetal (5.5 mmol), hydroquinone (0.05 mmol) and a catalytic amount of *p*-toluenesulfonic acid were added. The solution was stirred at 80 °C for 3 h and then reaction was quenched with TEA. DMF was evaporated under reduced pressure and the sticky residue was treated with diethyl ether (3 x 50 mL). **DT1** and **MT1** were separated by silica gel flash chromatography (CHCl₃ : MeOH : TEA 97:2 :1 \rightarrow 88:11:1) as yellowish oils (1.54 g, 45% and 0.28 g, 13%, respectively).

4,6-O-[p-(acryloyloxy(polyethylenoxy))benzylidene]- α , α '-D-trehalose (MT1)

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.15 (dd, 1H, J = 9.6, 5.2 Hz, H-4'); 3.26 (ddd, 1H, J = 9.6, 5.8, 3.7 Hz, H-2'); 3.34 (~t, 1H, J = 9.6 Hz, H-4); 3.39 (ddd, 1H, J = 9.8, 6.4, 3.8 Hz, H-2); 3.45-3.67 (m, ~33 H, H-3', H-6'a, H-6'b, H-6a, PEG backbone, H_g); 3.70 (ddd, 1H, J = 10.9, 4.8, 2.3 Hz, H-5'); 3.72-3.76 (m, 3H, H-3, H_f); 3.98 (~td, 1H, J = 9.9, 5.0 Hz, H-5); 4.04-4.10 (m, 3H, H-6b, H_e); 4.19-4.24 (m, 2H, H_h); 4.36 (~t, 1H, J = 6.0 Hz, C-6'-OH'); 4.75-4.79 (m, 2H, 2x –OH); 4.87 (d, 1H, J = 3.7 Hz, H-1'); 4.90-4.95 (m, 3H, H-1, 2x –OH); 5.16 (d, 1H, J = 5.1 Hz, –OH); 5.49 (s, 1H, –OCHO–); 5.92-5.98 (m, 1H, –CH=CH₂ *cis*); 6.15-6.23 (m, 1H, –CH=CH₂); 6.30-6.36 (m, 1H, –CH=CH₂, *trans*); 6.88-6.95 (m, 2H, H_c); 7.31-7.38 (m, 2H, H_b). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 63.44 (C_g); 67.14, 68.89 (C_e, C_f); 68.21 (C_h); 69.60-69.94 (PEG backbone); 60.69, 62.40, 68.21, 69.52, 70.00, 71.47, 72.13, 72.62, 72.71, 81.47 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5', C-6, C-6'); 93.81, 94.21 (C-1, C-1'); 100.72 (–OCHO–); 113.81 (C_c); 127.67 (C_b); 128.20 (–CH=CH₂); 130.31 (C_a); 131.63 (–CH=CH₂); 158.69 (C_d); 165.45 (–C(O)O–).



Fig. S1 ¹H and ¹³C NMR spectra of 4,6-*O*-[*p*-(acryloyloxy(polyethylenoxy))benzylidene]- α , α '-D-trehalose (MT1)

4,6:4',6'-di-O-[p-(acryloyloxy(polyethylenoxy))benzylidene]- α , α '-D-trehalose (DT1)

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.36 (~t, 2H, J = 9.4 Hz, H-4, H-4'); 3.42 (ddd, 2H, J = 9.6, 6.1, 3.8 Hz, H-2, H-2'); 3.47-3.60 (m, ~54 H, PEG backbone); 3.60-3.66 (m, 6H, H-6a, H-6'a, H_g, H_g'); 3.71-3.79 (m, 6H, H-3, H-3', H_f, H_f'); 4.00-4.10 (m, 8H, H-5, H-5', H-6b, H-6'b, H_e, H_e'); 4.19-4.23 (m, 4H, H_h, H_h'); 4.93 (d, 2H, J = 3.8 Hz, H-1, H-1'); 5.18 (d, 2H, J = 5.1 Hz, C-3-OH, C-3'-OH'); 5.25 (d, 2H, J = 6.0 Hz, C-2-OH, C-2'-OH'); 5.50 (s, 2H, –OCHO–, –OC'H'O–); 5.91-5.98 (m, 2H, –CH=CH₂, –C'H'=C'H'₂ *cis*); 6.14-6.23 (m, 2H, –CH=CH₂, –C'H'=C'H'₂); 6.29-6.36 (m, 2H, –CH=CH'₂, –C'H'=C'H'₂ *trans*); 6.87-6.95 (m, 4H, H_c, H_c'); 7.31-7.39 (m, 4H, H_b, H_b'). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 63.44 (C_g, C_g'); 67.13, 68.89 (C_e, C_e', C_f, C_f'); 68.20 (C_h, C_h'); 69.65-69.95 (PEG backbone); 62.57, 68.20, 69.42, 72.00, 81.42 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5', C-6, C-6'); 94.92 (C-1, C-1'); 100.75 (–OCHO–,–OC'H'O–); 113.81 (C_c, C_c'); 127.67 (C_b, C_b'); 128.20 (–**C**H=CH₂, –**C**'H'=C'H'₂); 130.29 (C_a, C_a'); 131.63 (–CH=**C**H₂, –C'H'=**C**'H'₂); 158.69 (C_d, C_d'); 165.45 (–C(0)O–, –C'(O)O–).



Fig. S2 ¹H and ¹³C NMR spectra of 4,6:4',6'-di-*O*-[*p*-(acryloyloxy(polyethylenoxy))benzylidene]- α , α '-D-trehalose (DT1)

2.2. Synthetic procedure for MT2 and DT2



Scheme S2 Synthetic pathway to trehalose monomers DT2 and MT2.

p-(2-Hydroxyethoxy)benzaldehyde dimethyl acetal



p-(2-Hydroxyethoxy)benzaldehyde dimethyl acetal was synthetized based on the reported procedure.⁴ To a solution of *p*-(2-hydroxyethoxy)benzaldehyde (78.0 mmol) and lithium tetrafluoroborate (4 mmol) in methanol (40 mL), trimethyl orthoformate (117.0 mmol) was added, and the solution was stirred under reflux for 3h. After cooling, saturated aqueous solution of NaHCO₃ (250 mL) was added, and the mixture was extracted twice with ethyl acetate (360 mL, 120 mL). The combined organic layers were washed with saturated aqueous solution of NaHCO₃ (120 mL), dried over MgSO₄ and concentrated under vacuum to afford product as an orange syrup (15.92 g, 96%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 2.28 (t, 1H, J = 6.2 Hz, –OH); 3.31 (s, 6H, –OCH₃); 3.93-3.97 (m, 2H, – CH₂OH); 4.06-4.10 (m, 2H, –OCH₂–); 5.35 (s, 1H, –OCHO–); 6.89-6.93 (m, 2H, H_c); 7.35-7.39 (m, 2H, H_b). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 52.74 (–OCH₃); 61.59 (-CH₂OH); 69.36 (–OCH₂–); 103.12 (–OCHO–); 114.33 (C_c); 128.14 (C_b); 131.04 (C_a); 158.86 (C_d). LR ESI-MS: m/z calcd for C₁₁H₁₆O₄Na [M+Na]⁺ 235.1, found 235.2

p-(2-(Methanesulfonyloxy)ethoxy)benzaldehyde dimethyl acetal



A solution of *p*-(2-hydroxyethoxy)benzaldehyde dimethyl acetal (30.0 mmol) and TEA (51.0 mmol) in anhydrous DCM (100 mL) was cooled on ice water bath, and then methanesulfonyl chloride (45.0 mmol) was added dropwise, while stirring. The mixture was left to warm up to room temperature and left overnight. Afterwards, the suspension was diluted with DCM (100 mL) and extracted with an aqueous solution of NaHCO₃ (100 mL). The organic layer was dried over MgSO₄ and concentrated under vacuum to afford product as an yellow syrup (7.99 g, 92%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 3.09 (s, 3H, $-SO_2CH_3$); 3.31 (s, 6H, $-OCH_3$); 4.23-4.27 (m, $-OCH_2-$); 4.55-4.59 (m, 2H, 2H, $-CH_2SO_2-$); 5.35 (s, 1H, -OCHO-); 6.87-6.92 (m, 2H, H_c); 7.36-7.40 (m, 2H, H_b). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 37.92 ($-SO_2CH_3$); 52.76 ($-OCH_3$); 65.96 ($-CH_2SO_2-$); 68.11 ($-OCH_2-$); 102.99 (-OCHO-); 114.32 (C_c); 128.26 (C_b); 131.60 (C_a); 158.17 (C_d). LR ESI-MS: m/z calcd for $C_{12}H_{18}O_6SNa$ [M+Na]⁺ 313.1, found 313.2

p-(2-Aminoethoxy)benzaldehyde dimethyl acetal



Aqueous ammonia (285 mL) was added to p-(2-(methanesulfonyloxy)ethoxy)benzaldehyde dimethyl acetal (25.0 mmol) and the solution was stirred vigorously for 2 days. The mixture was then extracted with DCM (250 mL), and afterwards the organic layer was washed twice with an aqueous solution of NaHCO₃ (250 mL), dried over MgSO₄ and concentrated under vacuum to afford product as an yellow syrup (5.11 g, 96%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 1.65 (br s, 2H, $-NH_2$); 3.08 (~t, 2H, J = 5.2, 5.2 Hz, $-CH_2NH_2$); 3.30 (s, 6H, $-OCH_3$); 3.99 (~t, 2H, J = 5.2, 5.2 Hz, $-OCH_2-$); 5.34 (s, 1H, -OCHO-); 6.86-6.92 (m, 2H, H_c); 7.32-7.38 (m, 2H, H_b). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 41.66 ($-CH_2NH_2$); 52.72 ($-OCH_3$); 70.20 ($-OCH_2-$); 103.16 (-OCHO-); 114.27 (C_c); 128.08 (C_b); 130.71 (C_a); 159.12 (C_d). LR ESI-MS: m/z calcd for $C_{11}H_{18}O_3N$ [M+H]⁺ 212.1, found 212.2

p-(2-acrylamidoethoxy)benzaldehyde dimethyl acetal



A solution of *p*-(2-aminoethoxy)benzaldehyde dimethyl acetal (24.0 mmol) and TEA (54.0 mmol) in THF (120 mL) was cooled on ice water bath, and then a solution of acryloyl chloride (36.0 mmol) in THF (10 mL) was added dropwise. The mixture was left to warm up to room temperature and the reaction was finished after 3 h. Precipitated triethylamine hydrochloride was separated on a pad of Celite 501, and

filtrate was concentrated under vacuum. The crude product was purified by silica gel flash chromatography (CHCl₃: TEA 99:1) to afford p-(2-acrylamidoethoxy)benzaldehyde dimethyl acetal as a yellowish syrup (3.65 g, 57%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 3.30 (s, 6H, $-OCH_3$); 3.74 (~q, 2H, J = 5.6 Hz, $-CH_2NH-$); 4.07 (~t, 2H, J = 5.1, 1.2 Hz, $-OCH_2-$); 5.33 (s, 1H, -OCHO-); 5.64 (dd, 1H, J = 10.2, 1.5 Hz, $-CH=CH_2$, *cis*); 6.12 (1H, J = 17.0, 10.2 Hz, $-CH=CH_2$); 6.13-6.25 (br s, 1H, -NHC(O)-); 6.29 (dd, 1H, J = 17.1, 2.2 Hz, $-CH=CH_2$, *trans*); 6.84-6.91 (m, 2H, H_c); 7.32-7.38 (m, 2H, H_b). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 39.13 ($-CH_2NH-$); 52.73 ($-OCH_3$); 66.83 ($-OCH_2-$); 103.07 (-OCHO-); 114.19 (C_c); 126.88 ($-CH=CH_2$); 128.17 (C_b); 130.75 ($-CH=CH_2$); 131.11 (C_a); 158.64 (C_d); 165.78 (-OC(O)-). LR ESI-MS: m/z calcd for $C_{11}H_{18}O_3Na$ [M+Na]⁺ 288.1, found 288.3

4,6:4',6'-di-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (DT2) and 4,6-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (MT2)



To a suspension of anhydrous trehalose (5.65 mmol) in anhydrous DMF (10 mL), *p*-(2-acrylamidoethoxy)benzaldehyde dimethyl acetal (13.0 mmol), hydroquinone (0.1 mmol) and a catalytic amount of *p*-toluenesulfonic acid were added. The solution was stirred for 3 h at 80 °C and then reaction was quenched with TEA. The solvent was evaporated under reduced pressure and the sticky residue was treated with diethyl ether (3 x 50 mL). **DT2** and **MT2** were separated by silica gel flash chromatography (CHCl₃ : MeOH : TEA 97:2 : 1 \rightarrow 85:14:1) as white solids (1.85 g, 44% and 0.36 g, 12%, respectively).

4,6-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (MT2)

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.15 (~t, 1H, J = 9.4, Hz, H-4'); 3.24-3.29 (m, 1H, H-2'); 3.34-3.53 (m, 5H, H-4, H-2, H-6'a, $-CH_2NH-$); 3.53-3.60 (m, 2H, H-3', H-6'b); 3.63 (~t, 1H, J = 10.2 Hz, H-6a); 3.70 (ddd, 1H, J = 10.0, 4.8, 2.3 Hz, H-5'); 3.74 (~t, 1H, J = 9.9 Hz, H-3); 3.98 (~td, 1H, J = 10.0, 5.0 Hz, H-5); 4.03 (~t, 2H, J = 5.63 Hz, $-OCH_2-$); 4.07 (dd, 1H, J = 10.0, 4.9 Hz, H-6b); 4.36 (br s, 1H, -OH); 4.60-5.10 (m, 4H, 4x -OH) 4.87 (d, 1H, J = 3.6 Hz, H-1'); 4.94 (d, 1H, J = 3.7 Hz, H-1); 5.20 (br s, 1H, J = 5.1 Hz, -OH); 5.50 (s, 1H, -OCHO-); 5.59 (dd, 1H, J = 10.2, 2.2 Hz, $-CH=CH_2$ *cis*); 6.10 (dd, 1H, J = 17.1, 2.2 Hz, $-CH=CH_2$, *trans*); 6.26 (dd, 1H, J = 17.1, 10.2 Hz, $-CH=CH_2$); 6.93 (d, 2H, J = 8.8 Hz, H_c, H_c'); 7.36 (d, 2H, J = 8.7 Hz, H_b, H_b'); 8.35 (t, 2H, J = 5.7 Hz, -NHC(O)-); ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 38.27 ($-CH_2NH-$); 66.26 ($-OCH_2-$, $-OC'H'_2-$); 60.70, 62.41, 68.22, 69.51, 70.01, 71.47, 72.15, 72.64, 72.71, 81.46 (C-2, C-2', C-3), C-3', C-4, C-4', C-5, C-5', C-6, C-6'); 93.84, 94.24 (C-1, C-1'); 100.70 (-OCHO-); 113.88 (C_c); 125.30



 $(-CH=CH_2)$; 127.71 (C_b); 130.46 (C_a) 131.57 ($-CH=CH_2$) 158.58 (C_d) 164.88 (-C(O)-). LR ESI-MS: m/z calcd for C₂₄H₃₃O₁₅N₂ [M+H]⁺ 544.2, found 544.2

Fig. S3 ¹H and ¹³C NMR spectra of 4,6-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (MT2)

4,6:4',6'-di-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (DT2)

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.36 (~t, 2H, J = 9.5 Hz, H-4, H-4');); 3.42 (ddd, 2H, J = 9.6, 6.0, 3.7 Hz, H-2, H-2'); 3.50 (~q, 2H, J = 5.6 Hz, $-CH_2NH-$, $-C'H'_2NH'-$); 3.63 (~t, 2H, J = 9.9 Hz, H-6a, H-6'a); 3.76 (~td, 2H, J = 9.3, 5.1 Hz, H-3, H-3'); 4.00-4.10 (m, 8H, H-5, H-5', H-6b, H-6'b, $-OCH_2-$, $-OC'H'_2-$); 4.93 (d, 2H, J = 3.8 Hz, H-1, H-1'); 5.19 (d, 2H, J = 5.1 Hz, C-3-OH, C-3'-OH'); 5.26 (d, 2H, J = 6.0 Hz, C-2-OH, C-2'-OH'); 5.51 (s, 2H, -OCHO-, -OCH'O-); 5.59 (dd, 2H, J = 10.2, 2.1 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$ *cis*); 6.10 (dd, 2H, J = 17.1, 2.1 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$ *trans*); 6.26 (dd, 2H, J = 17.1, 10.2 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$, 6.10 (dd, 2H, J = 17.1, 2.1 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$ *trans*); 6.26 (dd, 2H, J = 17.1, 10.2 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$); 6.93 (d, 4H, J = 8.7 Hz, H_c, H_c'); 7.36 (d, 4H, J = 8.7 Hz, H_b, H_b'); 8.36 (t, 2H, J = 5.7 Hz, -CHC(O)-, -NH'C'(O)-); ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 38.27 ($-CH_2NH-$, $-C'H'_2NH'-$); 66.26 ($-OCH_2-$, $-OC'H'_2-$); 62.56, 66.26, 68.19, 69.43, 72.01, 81.39 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5', C-6, C-6'); 94.94 (C-1, C-1'); 100.71(-OCHO-, -OC'H'O-); 113.88 (C_c, C_c'); 125.28 ($-CH=CH_2$, $-C'H'=C'H'_2$); 127.71 (C_b, C_b'); 130.44 (C_a, C_a'); 131.56 ($-CH=CH_2$, $-C'H'=C'H'_2$); 158.59 (C_d, C_d'); 164.88 (-C(O)-, -C'(O)-). **LR ESI-MS:** m/z calcd for C₃₆H₄₅O₁₃N [M+H]⁺745.3, found 745.5



Fig. S4 ¹H and ¹³C NMR spectra of 4,6:4',6'-di-O-[p-(2-acrylamidoethoxy)benzylidene]- α , α '-D-trehalose (DT2)

2.3. Synthetic procedure for MT3 and DT3



Scheme S3 Synthetic pathway of trehalose monomers DT3 and MT3.

2,3,4,6,2',3',4',6'-okta-O-trimethylsilyl- α , α '-D-trehalose



Anhydrous trehalose (21.9 mmol) was dissolved in 80 mL of anhydrous pyridine and the solution was cooled on ice-water bath before trimethylsilyl chloride (362.8 mmol) was added dropwise, while stirring. The reaction was allowed to warm up to room temperature and left overnight. The resulting suspension was diluted with 100 mL of ethyl acetate and extracted with 80 mL of DI water. The organic phase was washed two more times with 80 mL of DI water, dried over MgSO₄ and concentrated under vacuum. The residue was evaporated twice with toluene to afford product as white crystals (19.70 g, 98%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 0.10, 0.12, 0.14, 0.14 (4x s, 72H, $-Si(CH_3)_3$); 3.38 (dd, 2H, J = 9.3, 3.2 Hz, H-2, H-2'); 3.44 (~t, 2H, J = 9.1 Hz, H-4, H-4'); 3.64-3.71 (m, 4H, H-6a, H-6'a, H-6b, H-6'b); 3.76-3.81 (ddd, 2H, J = 9.5, 4.1, 2.3 Hz, H-5, H-5'); 3.89 (~t, 2H, J = 9.0 Hz, H-3, H-3'); 4.91 (d, 2H, J = 3.2 Hz, H-1, H-1'). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: -0.11, 0.32, 1.10, 1.24 (8x $-Si(CH_3)_3$); 62.31 (C-6, C-6'); 71.92, 73.04, 73.41, 73.77 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 94.53(C-1, C-1'). LR ESI-MS: m/z calcd for C₃₆H₈₆O₁₁Si₈Na [M+Na]⁺941.4, found 941.7

2,3,4,2',3',4',6'-hepta-O-trimethylsilyl- α , α '-D-trehalose



2,3,4,6,2',3',4',6'-okta-*O*-trimethylsilyl- α , α '-D-trehalose (4.3 mmol) was dissolved in 270 mL of methanol and cooled on ice-water bath before K₂CO₃ (4.1 mmol) was added, while intensively stirring. The reaction was quenched after 18 min by transferring into separating funnel containing 200 mL of hexane and extracting twice with 100 mL of DI water. The organic phase was dried over MgSO₄ and concentrated under vacuum. Crude product containing starting material, 2,3,4,2',3',4',6'-hepta-*O*-trimethylsilyl- α , α '-D-trehalose and 2,3,4,2',3',4'-hexa-*O*-trimethylsilyl- α , α '-D-trehalose was separated by silica gel flash chromatography (hexane: ethyl acetate 98:2) to afford 2,3,4,2',3',4',6'-hepta-*O*-trimethylsilyl- α , α '-Dtrehalose as a colorless oil (1.71 g, 47%). 1.02 g (25%) of starting material was recovered as a first fraction.

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 0.10, 0.11, 0.12, 0.14, 0.14, 0.14, 0.16 (7x s, 63H, $-Si(CH_3)_3$); 1.76 (br s, 1H, -OH); 3.38-3.44 (m, 3H, H-2, H-2', H-4'); 3.46 (~t, 2H, J = 9.1 Hz, H-4); 3.63-3.74 (m, 4H, H-6a, H-6b, H-6'a, H-6'b); 3.79 (ddd, 1H, J = 9.5, 4.7 Hz, 2.0 Hz, H-5'); 3.84 (~dt, 1H J = 9.5, 3.5 Hz, H-5); 3.89, 3.92 (2 t, 2H, J = 9.0 Hz, H-3, H-3'); 4.89 (d, 1H J = 3.1 Hz, H-1); 4.94 (d, 1H J = 3.1 Hz, H-1'). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: -0.15, 0.25, 0.27, 1.01, 1.09, 1.17, 1.20 (7x –Si(CH₃)₃); 61.88, 62.29 (C-6, C-6'); 71.62, 71.89, 72.87, 72.90, 72.94, 73.54, 73.55, 73.63 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 94.40, 94.57 (C-1, C-1'). LR ESI-MS: m/z calcd for $C_{33}H_{78}O_{11}Si_7Na$ [M+Na]⁺869.4, found 869.6

2,3,4,2',3',4'-hexa-O-trimethylsilyl- α , α '-D-trehalose



2,3,4,2',3',4'-hexa-O-trimethylsilyl- α,α' -D-trehalose was synthetized according to the procedure described for 2,3,4,2',3',4',6'-hepta-O-trimethylsilyl- α,α' -D-trehalose, except that alcoholysis was left to proceed for 120 min. Pure product was separated from the crude by silica gel flash chromatography (hexane: ethyl acetate 98:2 \rightarrow 90:10) as white crystals (2.76 g, 83%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 0.12, 0.15, 0.16 (3x s, 54H, $-Si(CH_3)_3$); 1.85 (br s, 2H, -OH) 3.42 (dd, 2H, J = 9.3, 3.1 Hz, H-2, H-2'); 3.48 (~t, 2H, J = 9.1 Hz, H-4, H-4'); 3.65-3.74 (m, 4H, H-6a, H-6'a, H-6b, H-6'b); 3.83-3.87 (~td, 2H, J = 9.5, 3.4 Hz, H-5, H-5'); 3.89 (~t, 2H, J = 9.0 Hz, H-3, H-3'); 4.91 (d, 2H, J = 3.1 Hz, H-1, H-1'). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 0.23, 1.00, 1.15 (6x $-Si(CH_3)_3$); 61.76 (C-6, C-6'); 71.53, 72.91, 73.16,73.49 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 94.80 (C-1, C-1'). LR ESI-MS: m/z calcd for C₃₀H₇₀O₁₁Si₆Na [M+Na]⁺ 797.3, found 797.5

6-O-acryloyl-2,3,4,2',3',4',6'-hepta-O-trimethylsilyl- α , α '-D-trehalose



A solution of 2,3,4,2',3',4',6'-hepta-*O*-trimethylsilyl- α , α '-D-trehalose (7.9 mmol) and TEA (17.8 mmol) in 30 mL of anhydrous DCM was cooled on ice water bath, before a solution of acryloyl chloride (11.9 mmol) in 20 mL of anhydrous DCM was added dropwise. The mixture was left to warm up to room temperature. After 2 h the solution was diluted with 160 mL of DCM and extracted twice with 220 mL of brine. The organic phase was dried over MgSO₄ and concentrated under vacuum. Crude product was purified by silica gel flash chromatography (hexane: ethyl acetate 98:2) to afford product as colorless oil (5.26 g, 74%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 0.08, 0.12, 0.13, 0.14, 0.15 (5x s, 63H, $-Si(CH_3)_3$); 3.40 (dd, 1H, J = 9.3, 3.1 Hz, H-2'); 3.42-3.47 (m, 2H, H-2, H-4'); 3.51 (dd, 1H, J = 9.5, 8.6 Hz, H-4); 3.64-3.70 (m, 2H, H-6'a), H-6'b); 3.77 (m, 2H,H-5'); 3.86-3.94 (m, 2H, H-3, H-3'); 4.05 (ddd, 1H J = 9.6, 4.4, 2.4 Hz, H-5); 4.16 (dd, 1H, J = 12.0, 4.4 Hz, H-6a); 4.37 (dd, 1H, J = 12.0, 2.4 Hz, H-6b); 4.90 (d,1H, J = 3.1 Hz, H-1'); 4.96 (d, 1H, J = 3.1 Hz, H-1); 5.84 (dd, 1H, J = 10.4, 1.4 Hz, $-CH=CH_2 cis$); 6.18 (dd, 1H, J = 17.3, 10.4 Hz, $-CH=CH_2$); 6.44 (dd, 1H, J = 17.3, 1.4 Hz, $-CH=CH_2 trans$). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: -0.12, 0.30, 0.35, 1.03, 1.07, 1.21, 1.23 (7x $-Si(CH_3)_3$); 62.09, 63.83 (C-6, C-6'); 70.74, 71.72, 72.14, 72.84, 72.97, 73.55, 73.64, 73.73 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 94.39, 94.70 (C-1, C-1'); 128.31 ($-CH=CH_2$); 131.28 ($-CH=CH_2$); 166.31 (-OC(O)-). LR ESI-MS: m/z calcd for C₃₆H₈₀O₁₂Si₇Na [M+Na]⁺923.4, found 923.8

6,6'-di-O-acryloyl-2,3,4,2',3',4'-hexa-O-trimethylsilyl-α,α'-D-trehalose



6,6'-di-O-acryloyl-2,3,4,2',3',4'-hexa-O-trimethylsilyl- α,α' -D-trehalose was synthetized according to the same procedure as described for 6-O-acryloyl-2,3,4,2',3',4',6'-hepta-O-trimethylsilyl- α,α' -D-trehalose, except that of TEA and acryloyl chloride were used in higher amounts (35.6 and 23.7 mmol respectively). Crude product was purified by silica gel flash chromatography (hexane: ethyl acetate 96:4) to afford the product as white crystals (5.41 g, 78%).

¹H NMR (CDCl₃, 600 MHz) δ [ppm]: 0.13, 0.14, 0.16 (3x s, 54H, $-Si(CH_3)_3$); 3.46 (dd, 2H, J = 9.3, 3.1 Hz, H-2, H-2'); 3.51 (dd, 2H, J = 9.6, 8.6 Hz, H-4, H-4'); 3.92 (~t, 2H, J = 9.0 Hz, H-3, H-3'); 4.05 (ddd, 2H, J = 9.6, 4.5, 2.4 Hz, H-5, H-5'); 4.16 (dd, 2H, J = 12.0, 4.5 Hz H-6a, H-6'a); 4.36 (dd, 2H, J = 12.0, 2.4 Hz, H-6b, H-6'b); 4.94 (d, 2H, J = 3.1 Hz, H-1, H-1'); 5.85 (dd, 2H, J = 10.4, 1.4 Hz, $-CH=CH_2$, $-C'H'=C'H'_2 cis$); 6.17 (dd, 2H, J = 17.3, 10.4 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$); 6.44 (dd, 2H, J = 17.3, 1.4 Hz, $-CH=CH'_2$, $-C'H'=C'H'_2 trans$). ¹³C NMR (CDCl₃, 150 MHz) δ [ppm]: 0.20, 0.86, 1.06 (6x $-Si(CH_3)_3$); 63.64 (C-6, C-6'); 70.76, 71.96, 72.68, 73.48 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 94.47 (C-1, C-1'), 128.12 ($-CH=CH_2$, $-C'H'=C'H'_2$); 131.14 ($-CH=CH_2$, $-C'H'=C'H'_2$); 166.10 (-OC(O)-, -OC'(O)-). LR ESI-MS: m/z calcd for C₃₆H₇₄O₁₃ Si₆Na [M+Na]⁺905.4, found 905.4

6-O-acryloyl- α , α '-D-trehalose (MT3)



Amberlyst 15* (8.8 g) was added to the solution of 6-*O*-acryloyl-2,3,4,2',3',4',6'-hepta-*O*-trimethylsilyl- α , α '-D-trehalose (2.5 mmol) in methanol and DI water (330 and 30 mL, respectively) and left at room temperature under stirring for 1 h. Amberlyst 15 was filtered off and the solution was then extracted with 110 mL of hexane. Aqueous phase was separated and concentrated under vacuum at 30 °C to ~15 mL. The residual solution was filtered through the syringe filter (0.045 µm) and freeze dried to afford **MT3** as a white solid (0.97 g, 98%).

*washed several times with methanol

¹H NMR (DMSO-d₆, 400 MHz) δ [ppm]: 3.21 – 3.08 (m, 2H, H-4, H-4'); 3.30 – 3.21 (m, 2H, H-2, H-2'); 3.47 (ddd, 1H, J = 11.4, 6.2, 5.0 Hz, H-6'a); 3.60 – 3.50 (m, 3H, H-3, H-3', H-6'b); 3.64 (ddd, 1H, J = 9.9, 4.9, 2.3 Hz, H-5'); 3.96 (ddd, 1H, J = 10.1, 5.6, 2.1 Hz, H-5); 4.16 (dd, 1H, J = 11.8, 5.7 Hz, H-6a); 4.29 (dd, 1H, J = 11.8, 2.1 Hz, H-6b); 4.34 (~t, 1H, J = 5.9 Hz, C-6-OH); 4.67 (d, 1H, J = 6.3 Hz, -OH); 4.69 (d, 1H, J = 6.0 Hz, -OH); 4.74 (d, 1H, J = 4.9 Hz, -OH); 4.76 (d, 1H, J = 5.2 Hz, -OH); 4.84 (d, 1H, J = 3.6 Hz, H-1); 4.85-4.89 (m, 2H, H-1', -OH); 5.09 (d, 1H, J = 5.3 Hz, -OH); 5.94 (dd, 1H J = 10.3, 1.7 Hz, 1H, -CH=CH₂ *cis*); 6.17 (dd, 1H, J = 17.3, 10.3 Hz, 1H, -CH=CH₂); 6.32 (dd, 1H, J = 17.3, 1.7 Hz, 1H, -CH=CH₂ *trans*). ¹³C NMR (DMSO-d₆, 100 MHz) δ [ppm]: 60.73, 63.56 (C-6, C-6'); 69.55, 70.06, 70.10, 71.45, 71.55, 72.59, 72.81 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 93.25, 93.36 (C-1, C-1'); 128.37 (-CH=CH₂); 131.48 (-CH=CH₂, -C'H'=C'H₂); 165.42 (-OC(O)-). LR ESI-MS: m/z calcd for C₁₅H₂₄O₁₂Na [M+Na]⁺ 419.1, found 419.3



Fig. S5 ¹H and ¹³C NMR spectra of 6-*O*-acryloyl- α , α '-D-trehalose (MT3)

6,6'-di-O-acryloyl- α , α '-D-trehalose (DT3)



DT3 was synthetized from 6,6'-di-*O*-acryloyl-2,3,4,2',3',4'-hexa-*O*-trimethylsilyl- α , α '-D-trehalose according to the same procedure as described for **MT3** and obtained as a white solid (1.09 g, 97%)

¹H NMR (DMSO-d₆, 600 MHz) δ [ppm]: 3.17 (ddd, 2H, J = 10.1, 8.8, 5.4 Hz H-4, H-4'); 3.28 (ddd, 2H, J = 9.7, 6.0, 3.7 Hz, H-2, H-2'); 3.55 (~td, 2H, J = 9.3, 4.9 Hz, H-3, H-3'); 3.97 (ddd, 2H, J = 10.0, 5.5, 2.1 Hz, H-5, H-5'); 4.16 (dd, 2H J = 11.8, 5.5 Hz, H-6a, H-6'a); 4.28 (dd, 2H, J = 11.8, 2.1 Hz, H-6b, H-6'b); 4.80-4.85 (m, 4H, H-1, H-1', 2x -OH); 4.90 (d, 2H J = 5.0 Hz, C-3-OH, C-3'-OH); 5.11 (d, 2H J = 5.4 Hz, C-4-OH, C-4'-OH); 5.94 (dd, 2H J = 10.4, 1.5 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$ *cis*); 6.18 (dd, 2H, J = 17.3, 10.4 Hz, $-CH=CH_2$, $-C'H'=C'H'_2$); 6.32 (dd, 2H, J = 17.3, 1.5 Hz, $-CH=CH'_2$, $-C'H'=C'H'_2$ *trans*). ¹³C NMR (DMSO-d₆, 150 MHz) δ [ppm]: 63.49 (C-6, C-6'); 69.64, 70.00, 71.38, 72.72 (C-2, C-2', C-3, C-3', C-4, C-4', C-5, C-5'); 93.69 (C-1, C-1'); 128.34 ($-CH=CH_2$, $-C'H'=C'H'_2$); 131.48 ($-CH=CH_2$, $-C'H'=C'H'_2$); 165.42 (-OC(O)-, -OC'(O)-). LR ESI-MS: m/z calcd for C₁₈H₂₆O₁₃Na [M+Na]⁺ 473.1, found 473.2



Fig. S6 ¹H and ¹³C NMR spectra of 6,6'-di-*O*-acryloyl- α , α '-D-trehalose (DT3)

3. Synthesis and study on hydrogels

Sample code	DMAM / MTx (w/w)	DMAM [mg] ([mmol])	MT1 or MT2 [mg] ([mmol])	DT1 or DT2 [mg] ([mmol])	Crosslinker to monomers ratio [mmol/g]	Yield [%]
DMAM-MT1-0	100/0	140 (1.412)	-	15 (0.011)	0.08	88
DMAM-MT1-15	85/15	119 (1.200)	21 (0.024)	15 (0.011)	0.08	83
DMAM-MT1-30	70/30	98 (0.989)	42 (0.048)	15 (0.011)	0.08	81
DMAM-MT2-0	100/0	140 (1.412)	-	7.9 (0.011)	0.08	88
DMAM-MT2-15	85/15	119 (1.200)	21 (0.039)	7.9 (0.011)	0.08	85
DMAM-MT2-30	70/30	98 (0.989)	42 (0.077)	7.9 (0.011)	0.08	80

3.1. Feed composition for the synthesis of acid-labile hydrogels

Table S1 Monomer feed compositions and hydrogels yields

3.2. Feed composition for the synthesis of alkali-labile hydrogels

Table S2 Monomer feed compositions and hydrogels yields

Sample code	AM / MT3 (w/w)	AM [mg] ([mmol])	MT3 [mg] ([mmol])	DT3 [mg] ([mmol])	Crosslinker to monomers ratio [mmol/g]	Yield [%]
AM-MT3-0	100/0	108 (1.519)	-	8.7 (0.019)	0.18	96
AM-MT3-25	75/25	81 (1.140)	27 (0.068)	8.7 (0.019)	0.18	96
AM-MT3-50	50/50	54 (0.760)	54 (0.136)	8.7 (0.019)	0.18	91
AM-MT3-75	25/75	27 (0.380)	81 (0.204)	8.7 (0.019)	0.18	87
AM-MT3-50-1.5	50/50	81 (1.140)	81 (0.204)	8.7 (0.019)	0.12	84

3.3. Estimation of trehalose content originating from mono- and diacetal

Wt% of trehalose originating from diacetal (wt%_{TRE(DT)}) was estimated based on the enzymatically determined content of trehalose (wt%_{T=ΣTRE}) and mole fraction of diacetal (X_{DT}). X_{DT} was calculated from the integration of trehalose anomeric signal ($I_{TRE H-1,H-1'}$) and aldehyde aromatic signals ($I_{Ar(A)}$, $I_{Ar(B)}$) in ¹H NMR spectra of degradation products (Fig. S1-S4). The following equation was used:

$$wt\%_{TRE(DT)} = X_{DT} * wt\%_{\Sigma TRE} = \frac{\frac{I_{Ar(A)} + I_{Ar(B)}}{2}}{I_{TRE H-1,H-1'}} * wt\%_{\Sigma TRE}$$



Fig. S7 Signals in ¹H NMR spectra of degradation products of acid-labile hydrogels used for estimation of trehalose content originating from mono- and diacetal.

Sample code	wt% _{TRE}	X _{DT}	$X_{MT} = 1 - X_{DT}$	wt% _{TRE(DT)}	wt% _{TRE(MT)}
DMAM-MT1-0	1.7	1.00	-	1.7	-
DMAM-MT1-15	6.1	0.35	0.65	2.1	4.0
DMAM-MT1-30	11.4	0.22	0.78	2.5	8.9
DMAM-MT2-0	2.0	1.00	-	2.0	-
DMAM-MT2-15	8.5	0.27	0.73	2.3	6.2
DMAM-MT2-30	16.4	0.17	0.83	2.8	13.6



Fig. S8 1 H NMR spectra of degradation products of acid-labile hydrogels containing trehalose monomers set MT1/DT1



Fig. S9 1 H NMR spectra of degradation products of acid-labile hydrogels containing trehalose monomers set MT2/DT2



Fig. S10 1 H NMR spectra of degradation products of alkali-labile hydrogels containing trehalose monomers set MT3/DT3

4. References

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