

Review

Extraction by Subcritical and Supercritical Water, Methanol, Ethanol and Their Mixtures

Yizhak Marcus

Institute of Chemistry, the Hebrew University of Jerusalem, Jerusalem 91904, Israel; ymarcus@vms.huji.ac.il

Received: 5 November 2017; Accepted: 27 December 2017; Published: 1 January 2018

Abstract: Hot, subcritical and supercritical water, methanol, ethanol and their binary mixtures have been employed to treat fuels (desulfurize coal and recover liquid fuels from coal and oil shales) and to extract valuable solutes from biomass. The properties of these solvents that are relevant to their extraction abilities are presented. Various extraction methods: accelerated solvent extraction (ASE), pressurized liquid extraction (PLE), supercritical fluid extraction (SFE, but excluding supercritical carbon dioxide) with these solvents, including microwave- and ultrasound-assisted extraction, are dealt with. The extraction systems are extensively illustrated and discussed.

Keywords: ethanol; methanol; water; pressurized liquid; supercritical extraction; fuels; biomass

1. Introduction

The concept of “green solvents” pertains to the wider area of “green chemistry”, for which several principles have been established. These include waste prevention, safety—no toxic or hazardous materials should be used or produced—maximization of energy efficiency, and minimization of the potential for accidents—explosion, fire, and pollution possibilities must be kept in mind. In view of these principles, the chemical community is proceeding in recent years towards sustainable industrial processes. Water is the “greenest solvent” imaginable: it is readily available at the required purity, it is cheap, it is readily recycled, non-toxic, non-flammable, and environmentally friendly. In its supercritical form, water is also considered as a “green solvent” [1]. Supercritical ethanol has also been mentioned as a “green solvent” and even supercritical methanol (although much less often). Therefore, the use of such fluids, also in their subcritical forms, is nowadays considered to be at the forefront of efforts to apply neoteric solvents to chemical processes. On the other hand, supercritical water in the presence of an oxidant is used for total oxidation of hazardous substances [1], and this aspect of its use should be kept in mind when its use for the extraction of useful products is considered.

Extraction by subcritical and supercritical water, methanol, and ethanol and their mixtures has been applied to many materials for the recovery of useful products from them. The treatment of coals and oil shales with these solvents, after possible desulfurization and denitrification, may yield liquid fuels of better environmental impact. Many plant and some animal biomass materials are sources of bio-active products, such as anti-oxidants, biofuels, and bio-oils, when subjected to extraction with these solvents at the named conditions. Some of the biomass is conventionally extracted by soxhlet devices, but accelerated solvent extraction (ASE) is faster and economically superior; the term ASE means the use of pressurized liquid extraction (PLE) or supercritical fluid extraction (SFE). However, SFE is generally understood as extraction with supercritical carbon dioxide (scCO₂), possibly modified by co-solvents, but extraction with this particular method is outside the scope of this review, which is limited to neat supercritical water, methanol, ethanol, or their mixtures.

In order to be able to understand and follow the many publications that deal with the extraction by subcritical and supercritical water, methanol, ethanol and their mixtures and apply them in the future it is necessary to know their physical and chemical properties. These are summarized in

Section 2 of this review and in several tables. Water, methanol, and ethanol at ambient conditions have highly hydrogen-bonded structures, which are destroyed to an appreciable extent as the temperature is increased. This phenomenon reduces their dielectric permittivities and makes them less polar and tight, and facilitates the dissolution of non-polar substances in them. However, the increased temperatures, at liquid-like densities, do not detract from the dipolarity and hydrogen bond donation and acceptance abilities of these solvents so that they also dissolve and solvate polar solutes.

An extensive body of publications deals with the application of subcritical and supercritical water, methanol, and ethanol and their mixtures to the production of liquid and gaseous fuels from coals and oil sands. These are reviewed in Section 3, with the understanding that the reports dealt with are illustrative rather than comprehensive regarding this topic. Extraction with the title solvents of biomass of various origins has also been extensively dealt with in the literature. Pressurized liquid extraction with hot-but-subcritical water, methanol, and ethanol and their mixtures is dealt with in Section 4. The extraction of phenolic bio-active compounds, useful as antioxidants, as well as other kinds of useful products, is reviewed in this section, again with the restriction that the reports are merely illustrative. Extraction with the supercritical solvents of cellulosic and lignin biomass is reviewed in Section 5, where their application to algae and other plant materials is also dealt with. Finally, in Section 6, some methods, such as ultrasound- and microwave-assisted extraction combined with the pressurized liquid extraction are shown, leading to optimization of the extraction processes.

2. Properties of Subcritical and Supercritical Water, Methanol, and Ethanol and Their Mixtures

The critical properties: critical temperature, T_c , critical pressure, P_c , critical density, ρ_c , and critical molar volume, $V_c = M/\rho_c$ (M is the molar mass) of the three pure fluids water, methanol, and ethanol are presented in Table 1. The critical compressibility factor, $Z_c = P_c V_c / RT_c$, is also shown there, being nearly the same for these three fluids.

Table 1. The properties of pure water, methanol, and ethanol at the critical point.

Property	Water [1]	Methanol [2]	Ethanol [2]
Critical temperature, T_c/K	647.096	512.6	514.1
Critical pressure, P_c/MPa	22.064	8.092	6.137
Critical density, $\rho_c/kg\ m^{-3}$	322	272	276
Critical molar volume, $V_c/cm^3\ mol^{-1}$	55.95	117.7	166.8
Critical compressibility factor	0.2294	0.2234	0.2397
Permittivity, ϵ	8.63	-	-

The properties of hot, subcritical water, methanol, and ethanol along the saturation line are shown in Tables 2–4, respectively [1,3–13]. These properties include the vapor pressures p , the molar enthalpies of vaporization $\Delta_V H$, the density ρ , the isobaric expansibility α_P , the isothermal compressibility κ_T , the internal pressure $P_{int} = T\alpha_P/\kappa_T - p$, the isobaric molar heat capacity C_P , and the static permittivity ϵ . In the case of water (Table 2) data on the surface tension, the dynamic viscosity η , and the specific electrical conductivity κ could also be found and are included. The surface tension, σ , of methanol and ethanol is available only for the liquids below their normal boiling points, diminishing linearly with the temperature. Up to 333.15 K the values are $\sigma/mN\ m^{-1} = 19.41 - 0.0774 (T/K)$ for methanol and $19.06 - 0.0832 (T/K)$ for ethanol [14].

For water and ethanol at elevated temperatures the solvatochromic indices, indicating the polarity/polarizability (Kamlet-Taft π^*), hydrogen bond donation ability (Kamlet-Taft α) and hydrogen bond acceptance ability (Kamlet-Taft β) are shown in Tables 2 and 4. No such data were found for methanol.

It should be noted that the polarities and permittivities of the solvents considered here diminish with increasing temperatures, and do so drastically as the critical point is approached. Supercritical water, for instance, is capable of dissolving hydrocarbons such as hexane. Such behavior is connected

with the destruction of the three-dimensional hydrogen bond structure of water and the chain-like and ring hydrogen bonded structures of the alkanols [1,10,13].

Table 2. Properties of hot, subcritical water ($M = 0.018015 \text{ kg mol}^{-1}$), along the saturation line [1].

T/K	423	473	523	573	623
Reduced temperature, $T_r = T/T_c$	0.654	0.731	0.808	0.885	0.963
Vapor pressure, p/MPa	0.476	1.555	3.978	8.592	16.537
Molar enthalpy of vaporization, $\Delta_v H/\text{kJ mol}^{-1}$	38.41	34.40	30.95	26.63	15.97
Density, $\rho/\text{kg m}^{-3}$	917.0	864.9	799.1	712.2	575.0
Molar volume, $V/\text{m}^3 \text{ mol}^{-1}$	0.01965	0.02083	0.02255	0.02530	0.03133
Isobaric expansibility, $1000\alpha_p/\text{K}$	0.93	1.47	2.11	2.95	4.38
Isothermal compressibility, κ_T/GPa^{-1}	0.630	0.864	1.193	1.617	2.136
Internal pressure, $P_{\text{int}}/\text{MPa}$	742	717	616	538	482
Isobaric heat capacity, $C_p/\text{J K}^{-1} \text{ mol}^{-1}$	77.67	80.79	87.38	100.87	126.07
Surface tension, $\sigma/\text{mN m}^{-1}$	48.70	37.81	26.19	14.39	3.79
Viscosity, $\eta/\text{mPa s}$	0.182	0.134	0.106	0.086	0.066
Specific conductivity, $\kappa/\text{mS m}^{-1}$	0.169	0.298	0.328	0.229	0.063
Permittivity, ϵ	43.94	34.74	27.08	20.26	13.16
Kamlet-Taft π^* [3]	0.98	0.88	0.78	0.62 ^a	0.44 ^{a,b}
K-T H-bond donation α [3]	0.93	0.88	0.85	-	-
K-T H-bond acceptance β [3]	0.18	0.19	0.20	-	-

^a [4]; ^b at 634 K.

Table 3. Properties of hot, subcritical methanol ($M = 0.03204 \text{ kg mol}^{-1}$), along the saturation line.

T/K	373	398	423	448	473	498
Reduced temperature, $T_r = T/T_c$	0.728	0.776	0.825	0.874	0.923	0.971
Vapor pressure, p/MPa	0.3524	0.7175	1.3843	2.4618	4.0137	6.184
Molar enthalpy of vaporization, $\Delta_v H/\text{kJ mol}^{-1}$ [5]	32.70	30.53	28.07	24.99	20.63	13.58
Density, $\rho/\text{kg m}^{-3}$ [6]	771.1	678.2	646.9	598.8	551.1	514.3
Molar volume, $V/\text{cm}^3 \text{ mol}^{-1}$	45.07	47.24	49.53	53.51	58.14	62.30
Isobaric expansibility, $1000\alpha_p/\text{K}$ [6]	1.611	1.907	2.543	4.084	9.718	18.918
Isothermal compressibility, κ_T/GPa^{-1} [6]	2.28	2.96	4.47	8.52	28.34	61.08
Internal pressure, $P_{\text{int}}/\text{MPa}$	263	256	239	221	158	61
Isobaric heat capacity, $C_p/\text{J K}^{-1} \text{ mol}^{-1}$	101.6	111.1	-	-	-	-
Viscosity, $\eta/\text{mPa s}$ [7]	0.247	0.191	0.141	0.098	0.063	0.034
Permittivity, ϵ [8]	21.01	17.52	14.40	11.65	9.28	7.29

Table 4. Properties of hot, subcritical ethanol ($M = 0.04607 \text{ kg mol}^{-1}$), along the saturation line.

T/K	373	398	423	448	473	498
Reduced temperature, $T_r = T/T_c$	0.726	0.775	0.823	0.872	0.920	0.969
Vapor pressure, p/MPa	0.226	0.498	0.983	1.734	2.945	4.715
Molar enthalpy of vaporization, $\Delta_v H/\text{kJ mol}^{-1}$ [9]	37.12	34.41	30.92	26.66	21.61	15.79
Density, $\rho/\text{kg m}^{-3}$ [10]	715.8	690.5	664.4	637.5	610.1	574.6
Molar volume, $V/\text{cm}^3 \text{ mol}^{-1}$	64.3	66.7	69.3	72.3	75.5	80.2
Isobaric expansibility, $1000\alpha_p/\text{K}$ [10]	1.555	1.612	1.675	1.746	1.824	1.937
Isothermal compressibility, κ_T/GPa^{-1} [11]	1.820	2.113	2.438	2.794	3.180	3.597
Internal pressure, $P_{\text{int}}/\text{MPa}$	319	291	291	280	271	268
Isobaric heat capacity, $C_p/\text{J K}^{-1} \text{ mol}^{-1}$ [12]	152.1	170.2	180.2	-	-	-
Viscosity, $\eta/\text{mPa s}$	-	-	-	-	-	-
Permittivity, ϵ [8]	14.76	12.13	9.88	8.00	6.50	5.37
Kamlet-Taft dipolarity/polarizability, π^* [13]	0.50	0.41	0.27	0.26	0.18	0.03
Reichardt polarity, $E_T(30)/\text{kJ mol}^{-1}$ [13]	198	189	185	177	171	165
Kamlet-Taft H-bond donation α [13]	0.67	0.63	0.57	0.55	0.53	0.53
Kamlet-Taft H-bond acceptance β [13]	0.63	0.60	0.56	0.53	0.50	0.47

The density of supercritical water is shown in Table 5 at several pressures ($P_r = P/P_c$) and temperatures and the densities of supercritical methanol and ethanol at similar reduced pressures and temperatures are shown in Table 6.

Table 5. The density $\rho/\text{kg m}^{-3}$ of supercritical water at several pressures and temperatures [1].

P/Mpa	T/K	673	698	723	748	773	798	823	848	873
	T_r	1.040	1.079	1.117	1.156	1.195	1.233	1.272	1.310	1.349
25, $P_r = 1.14$		166	135	109	97	90	86	77	73	71
30, $P_r = 1.36$		353	241	149	123	115	103	98	93	87
35, $P_r = 1.58$		474	301	202	158	144	131	120	115	105
40, $P_r = 1.81$		524	382	272	213	178	158	143	135	124
50, $P_r = 2.26$		578	477	401	322	258	226	196	174	166

Table 6. The densities $\rho/\text{kg m}^{-3}$ of supercritical methanol and ethanol at several pressures and temperatures.

P/Mpa	T/K	523	548	573	598	623	648
Methanol [6]							
	T_r	1.019	1.068	1.117	1.166	1.215	1.263
10, $P_r = 1.23$		312	133	95	72	64	-
15, $P_r = 1.85$		457	315	209	138	102	-
20, $P_r = 2.47$		-	-	360	267	204	171
25, $P_r = 3.09$		-	497	412	341	276	218
Ethanol [11,15]							
	T_r	1.018	1.066	1.115	1.164	1.212	1.261
10, $P_r = 1.63$		-	270	241	208	-	-
15, $P_r = 2.44$		-	436	337	262	210	182
20, $P_r = 4.07$		-	492	420	357	304	259
25, $P_r = 4.89$		587	524	466	413	364	321

The critical data (temperatures, pressures, densities, and molar volumes) of mixtures of water with the co-solvents methanol and ethanol are shown in Table 7 as functions of their mole fractions x . The densities, $\rho/\text{kg m}^{-3}$ of supercritical aqueous mixtures of methanol and ethanol at specified mole fractions of the alkanol, temperatures, and pressures are shown in Table 8. The values were interpolated from detailed T - P - ρ - x tables of data and are deemed accurate to $\pm 2\%$.

Table 7. The critical temperatures, T_c/K , pressures, P_c/MPa , densities, $\rho_c/\text{kg m}^{-3}$, and molar volumes, $V_c/\text{cm}^3 \text{ mol}^{-1}$, of aqueous mixtures of methanol and ethanol at specified mole fractions x of the co-solvent and expressions representing these quantities.

x	0.20	0.35	0.50	0.65	0.80
Methanol [10], T_c	609.9	585.4	563.8	545.1	529.3
P_c	17.72	14.95	12.61	10.68	9.17
ρ_c	313.1	304.2	296.6	288.9	281.3
V_c	68.1	77.3	86.6	95.9	105.1
Ethanol [15], T_c	599.3	571.6	549.5	532.1	521.4
P_c	16.01	12.44	9.69	7.77	6.66
ρ_c	306.9	296.9	288.9	283.0	279.2
V_c	78.3	95.3	112.3	129.2	146.2

Methanol: $T_c/\text{K} = 647.1 - 200.6x + 65.8x^2$; $P_c/\text{MPa} = 22.06 - 23.58x + 9.34x^2$; $\rho_c/\text{kg m}^{-3} = 323.3 - 50.89x$; $V_c/\text{cm}^3 \text{ mol}^{-1} = 55.7 + 61.8x$; Ethanol: $T_c/\text{K} = 647.1 - 258.8x + 127.0x^2$; $P_c/\text{MPa} = 22.06 - 33.88x + 18.29x^2$; $\rho_c/\text{kg m}^{-3} = 323.3 - 91.4x + 45.34x^2$; $V_c/\text{cm}^3 \text{ mol}^{-1} = 55.7 + 113.1x$.

Table 8. The densities, $\rho_c/\text{kg m}^{-3}$ of supercritical aqueous mixtures of methanol and ethanol at specified mole fractions of the alkanol, temperatures, and pressures.

Aqueous Methanol [16,17]					
		$x = 0.2$	$x = 0.36$	$x = 0.5$	$x = 0.8$
$T = 573 \text{ K}$	$P = 10 \text{ MPa}$	-	74	204	-
	$P = 20 \text{ MPa}$	-	-	501	376
$T = 623 \text{ K}$	$P = 10 \text{ MPa}$	-	50	-	75
	$P = 20 \text{ MPa}$	186	200	177	187
$T = 673 \text{ K}$	$P = 10 \text{ MPa}$	-	47	-	-
	$P = 20 \text{ MPa}$	-	120	-	-
Aqueous Ethanol [11]					
$T = 573 \text{ K}$	$P = 10 \text{ MPa}$	68	-	107	145
	$P = 20 \text{ MPa}$	-	-	465	441
$T = 623 \text{ K}$	$P = 10 \text{ MPa}$	-	-	106	102
	$P = 20 \text{ MPa}$	265	-	355	270
$T = 673 \text{ K}$	$P = 10 \text{ MPa}$	-	-	65	83
	$P = 20 \text{ MPa}$	125	-	163	198

3. Supercritical Extraction Applied to Fuels

One of the aspects of the application of supercritical fluids to the extraction of coals is their desulfurization, involving the removal of both inorganic and organic sulfur compounds. As an example, Ohio coals contain 3.3 wt % total sulfur, of which some 60% are organic. When extracted with supercritical methanol, aqueous methanol, or water (at 560–712 K and 10.5–28.6 MPa) some 20% to 40% of the sulfur is removed. Water is most effective for the removal of pyritic sulfur, whereas methanol is most effective for the removal of organic sulfur [18]. Illinois coals may contain less sulfur, 1.4 wt %, of which up to 60% could be removed by supercritical methanol at 673 K [19]. Both supercritical aqueous methanol and aqueous ethanol were applied to Midwestern bituminous coals, where again the removal of the inorganic (pyritic and sulfate) forms of sulfur was more efficient than that of the organic sulfur [20]. Chinese coals are richer in sulfur, and a high-ranking coal with 4.0 wt % sulfur was treated with supercritical ethanol (which may contain a small amount of water) at 673 K and 12 MPa. Some 30% of the sulfur could be removed, most effectively with 95% ethanol [21]. Supercritical methanol at 673 to 723 K removed about 45% of the organic sulfur from Canadian coal [22]. Desulfurization of low-ranking coal from Thailand appears to require the addition of potassium hydroxide to the supercritical ethanol. Up to 48% of the sulfur could be removed with 5 g dm⁻³ of KOH at 623 K and 8.3 MPa of supercritical ethanol [23]. Desulfurization of coal with supercritical methanol or ethanol leads to partial transformation of the pyrite to pyrrhotite and to eventual coke formation. For a given level of sulfur removal, methanol produces a lower coke yield than ethanol [24]. In analogy with desulfurization may be considered the removal of oxygen from sub-bituminous coals on supercritical water extraction. When the temperature was increased from 633 to 673 K the oxygen removal increased from 53% to 71% [25].

Another aspect of the application of supercritical fluids to coal is the winning of liquid fuels from it. The conversion of South African coals to liquid and gaseous products by supercritical fluid extraction was studied with 18 fluids, among which were water, methanol, and ethanol. At 723 K and 20 MPa extraction conditions these supercritical fluids resulted in 18.4%, 18.9%, and 25.5% conversion, rather less than obtained with some other supercritical fluids (tetralin, *m*-cresol, aniline, and cyclohexanone) at the same conditions [26]. Illinois No. 6 bituminous coal was subjected to supercritical fluid extraction at 673 K with water (at 21.2–24.7 MPa), methanol (at 8.6 MPa), and ethanol (at 11.9 MPa), among other solvents, but in this study the conversion yield with the water (34% and 39%) was larger than with methanol (8.4%) and ethanol (17.5%) [27]. Supercritical ethanol at 673 K and 20.5 MPa

yielded 35.7% conversion to liquid products, but only 78.6% ethanol recovery, which was increased to 84.3% recovery on lowering the pressure at 673 K to 12.7 MPa, but at the expense of a much lower conversion to liquid products, 22.2% [28]. Brazilian coals were extracted by supercritical methanol and ethanol (also propanols, butanols, and pentanols) with recovery of >40% of liquid and gaseous products [29]. High-volatile bituminous coal from the Saar region in Germany was subjected to supercritical extraction with methanol and ethanol and the dependence of the extraction yields on the conditions employed was studied [30]. Both pure ethanol and its aqueous mixtures (also isopropanol) were applied as supercritical fluids at 598–698 K and ≤ 12.5 MPa to high-ash Brazilian coal, with increased temperatures and pressures increasing the conversion yield [31].

Supercritical water was found to act as both an extracting solvent as well as a reactant, converting coal to liquid and gaseous products [32]. Supercritical water at 633–673 K applied to sub-bituminous coals resulted in 36% to 44% total conversion [25]. Supercritical water at 673–698 K and 14–24.5 MPa was applied to Western Canadian lignite and high-volatile bituminous coals, with 50% and 35% maximal conversion (on a dry, ash-free basis) to liquid and gaseous products [33]. Supercritical, but also subcritical, water was applied to lignite coal for the extraction of liquid products, applied to Turkish [34] and Dayan lignite coals [35]. Their yield increased with increasing pressure and temperatures of 673 to 723 K were effective, but with increasing temperatures mainly gas and light oil production was enhanced. A similar study with sub- and supercritical water was conducted on hard coals at 573 to 673 K. Admixture in a 1:1 mass ratio of methanol and more so ethanol to the water increased the yield of liquid products significantly (at 623 K) [36]. Sub- and supercritical water were also applied to some Chinese coals, the maximal extraction rate occurred near 670 K and the gas (CO, H₂, CH₄) and extract yields increased with the pressure [37]. Higher temperatures 673 to 1033 K, and pressures, 30 MPa, were used for the supercritical extraction of brown coal from Russia, with conversion of 48% to 63% to liquid and gaseous products [38]. Russian brown coal was treated with supercritical water at 673 to 873 K for the production of combustible volatile products in a continuously supplied reactor [39,40]. Bitumen has been subjected to sub- and supercritical water extraction for the production of gaseous fuel, mainly hydrogen and carbon monoxide at supercritical conditions [41]. The catalytic gasification of bituminous Chinese coal by supercritical water in the presence of potassium carbonate catalyst was studied in [42] and the rates of production of hydrogen and methane are shown in Figure 1. The methane yield reaches a plateau at an early stage, but that of hydrogen increases with time and is more sensitive to the temperature. The co-liquefaction of coal and *Dunaliella tertiolecta* algae for the production of liquid fuel by 60 vol % ethanol in water mixtures at 633 K (supercritical for the mixture) was studied in [43].

Several studies dealt with some aspects of the application of the supercritical fluids considered here to coals that were not the winning of liquid fuels per se. The interaction of two model compounds, quinoline and isoquinoline with supercritical water was examined. It was found that these compounds were more reactive in the presence of supercritical water than when undergoing inert pyrolysis. Some other compounds (benzonitrile, aniline, tetralin, dihydroanthracene, and ethylbenzene) were studied less intensively with regard to their interaction with supercritical water, and alkyl side chains on aromatics were found to be somewhat reactive and that carbon atoms reacted with the water forming hydrogen [44]. Alkylation of C atoms, but also of O, S, and N atoms, was found spectroscopically to occur on extraction of Spanish lignite coal with supercritical methanol and ethanol at 523 to 623 K and up to 15.2 MPa [45]. Coal char is constituted mainly by polycyclic aromatic hydrocarbons (PAH), of which anthracene is the model compound. Its conversion to hydrogen by supercritical water at 873 to 1023 K was studied in [46].

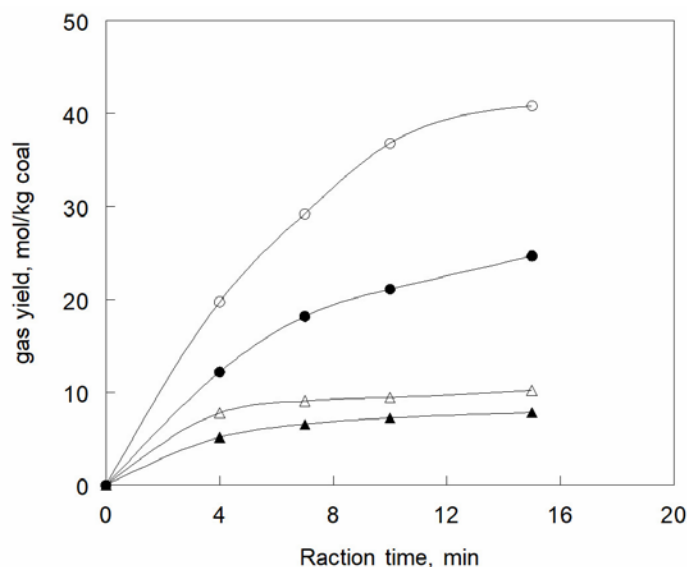


Figure 1. The yields of hydrogen (circles) and methane (triangles) on the catalytic treatment of bituminous coal with supercritical water at 923 K (filled) and 1023 K (empty) [42].

Another kind of material treated by supercritical fluids in order to obtain liquid (eventually also gaseous) fuels is oil shales. Extraction of Green River oil shale with supercritical water, methanol, and their mixtures at 473 K showed that part of the methanol was consumed by yielding esterification products, but it produced liquid organic materials beyond these [47]. Use of a 50 vol % mixture of water and methanol yielded 90% of the organic material in the shale on extraction at 673 K for 34 min [48]. Supercritical water was applied to Chinese Maoming oil shales at 673 K with 45% oil recovery when heated at this temperature for 1 h [49]. Extraction of Chinese (Huadian) oil shales with water for 30 min at 633 K (subcritical) to 723 K showed a maximal yield of 81.2% at 673 K, of which one half was oil and the rest asphaltene and pre-asphaltene [50]. Supercritical water was applied at a lower temperature, 648 K, to extract oil from Turkish oil shale, with 75% yield after 1 h treatment [51]. Turkish (Beypazari) oil shales were extracted with water at 623 (subcritical) to 698 K, with the asphaltene fraction diminishing but the polar fraction of the produced oil increasing in this temperature range [52]. Supercritical water applied to Moroccan oil shales at 653 K to 673 K yielded oils with increasing yields and diminishing fractions of asphaltenes [53]. Jordanian (El-Lajjun) oil shales were subjected to retorting and supercritical extraction with CO₂ and with water. Supercritical water at 712 K and 24.9 MPa produced the highest oil yield, nearly 47% [54]. Russian oil shales (Bazhenov formation) were treated with water at temperatures from 573 K (subcritical) to 753 K (supercritical) and liquid products were recovered at 673 K and 30 MPa, whereas at the highest temperature secondary hydrocarbon formation was observed [55]. Supercritical water extraction was applied to Russian (Krassava) and Bulgarian (Syssola) oil shales at 623 K, and admixtures of methanol and ethanol at 50 wt % to the water enhanced the oil recovery from 18% to 37% and 44% for the Russian shale, but less (from 13% to 18% and 35%) for the Bulgarian shale [36]. Spanish oil shales (from Puertollano) were treated by supercritical methanol at 553 to 623 K and the kerogen produced was mainly constituted by aliphatic hydrocarbon, contrary to the nature of the kerogen produced by supercritical toluene, which was mostly aromatic [37].

All these reports are summarized in Table 9.

Table 9. Application of supercritical water, methanol, ethanol and their mixtures to fuels.

Supercritical Solvent	References
Desulfurization of coal	
Water	[18,25]
Methanol	[19,22,24]
Ethanol	[21,23,24]
Water + methanol	[18,20]
Water + ethanol	[20]
Recovery of liquid fuels from coal	
Water	[25–27,32–35,38–40]
Methanol	[26,27,29,30,45]
Ethanol	[26–31,45]
Water + methanol	[36]
Water + ethanol	[31,36]
Recovery of liquid fuels from oil shales	
Water	[36,49–55]
Methanol	[47]
Water + methanol	[48]

4. Pressurized Subcritical Liquid Extraction from Bio-Materials

Pressurized liquid extraction (PLE) applies subcritical hot solvents at elevated pressures for the efficient extraction of useful products from bio-materials. This technique has also been called accelerated solvent extraction (ASE) [56]. Bio-active products are apt to decompose at very high temperatures, hence supercritical extraction may not be applicable to them. The application of near- and supercritical water to biorefineries has been summarized in a book [57]. Pressurized liquid extraction is preferable to extraction with supercritical water, methanol, ethanol and their mixtures when materials are to be recovered that are sensitive to the higher temperatures needed for the supercritical conditions.

A class of products that has been obtained by the PLE technique is anti-oxidants, such as carotenoids and polyphenols, and a fairly large set of studies has been devoted to this topic in recent years. Microalgae are a source of carotenoid antioxidants. Pressurized water and ethanol at 383 and 443 K were applied to *Spirulina platensis* microalgae to extract antioxidants, the activity of which was immediately ascertained [58], ethanol being the optimal solvent [59]. The rate of the extraction is shown in Figure 2 [60]. Pressurized water and ethanol were applied at 313, 373 and 433 K to *Dunaliella salina* microalgae to extract antioxidants, the best results being obtained with ethanol at 433 K [61]. Pressurized ethanol has again been applied to a microalga, *Haematococcus fluviialis*, at temperatures of 323, 373, 423, and 473 K, and although an increase in the temperature enhances the yield, this was deleterious to the antioxidant activity [62].

Pressurized water, methanol and ethanol were applied at 373 to 473 K for the extraction of the antioxidants catechin and epicatechin from tea leaves and grape seeds. Methanol was most efficient and temperatures <403 K led to $\geq 95\%$ yields [63]. Polyphenolic antioxidants were obtained by the application of pressurized water and ethanol to olive leaves. Water at 473 K and ethanol at 423 K were most effective [64]. A pressurized 50 mass% ethanol + water mixture at 473 K produced the best yield of phenolic antioxidants from black bamboo leaves [65]. Pressurized 80 mass% ethanol in water at 433 K was effective for the extraction of antioxidants from bitter melon fruits (*Momordica charantia*). Pressurized water and ethanol were used to extract phenolic antioxidants from spent coffee grounds and a temperature of 468 K was found to be most useful [66].

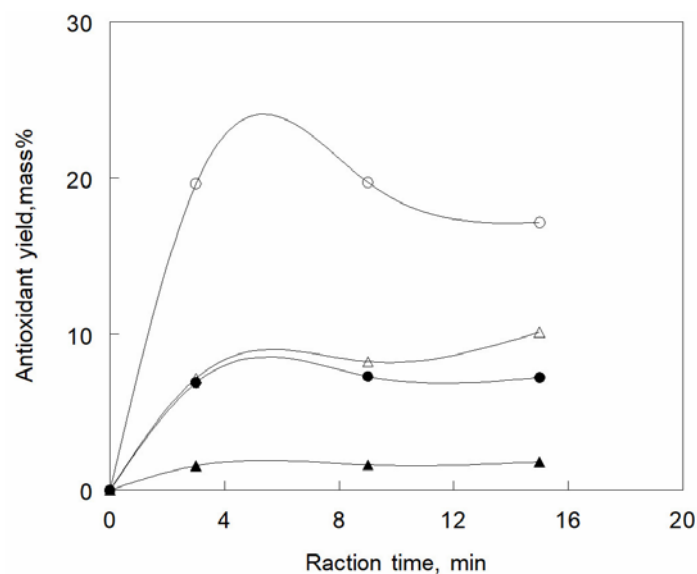


Figure 2. Antioxidant yields from pressurized liquid extraction of *Spirulina platensis* microalgae with ethanol (circles) and water (triangles) at 333K (filled) and 388 K (empty) [60].

Red grape skins appear to be a good source for phenolic and flavonoid antioxidants. Pressurized methanol at 423 K was effective for the extraction of phenolic antioxidants from grape skins, but flavonoids tended to degrade after prolonged treatment [67]. On the other hand, pressurized water at 383 to 433 K was effective for the extraction of anthocyanin antioxidants from red grape skins, with the yield increasing with the temperature in this range, but the anti-oxidative effect decreased [68]. Temperatures of up to 393 K were most effective for pressurized water extraction from grape skins, and this solvent was more efficient than pressurized methanol or ethanol [69]. A 50 mass% ethanol + water mixture, pressurized to 9 MPa at 393 K, was the most efficient for the extraction of phenolic antioxidants from red grape pomace, compare with neat water or ethanol in these conditions [70]. Pressurized ethanol at 5 to 15 MPa and 313 to 393 K was effective for the extraction of anthocyanin antioxidants from Brazilian jaboticaba skins [71].

Further studies of the pressurized liquid extraction of phenolic compounds from bio-materials, without stressing their antioxidant properties have also been reported. PLE was applied to methanol, ethanol, and their mixtures with water for the extraction of phenolic compounds from parsley (*Petroselinum crispum*) flakes, and 50 vol % ethanol+water appeared to be the most effective [72]. A 60 vol % methanol+water mixture at 363 K was best for PLE of phenolic acids from black cohosh (*Cimicifuga racemosa*) [73]. Water and 70 mass% ethanol in water mixtures were used for PLE at 323 to 463 K of spinach to yield flavonoids, the mixture being more effective than neat water [74].

Phenolic bio-active compounds were extracted from virgin olive oil filter cake by pressurized mixtures of water and ethanol, the best yield was with 50 vol % mixtures at 393 K [75]. Pressurized water and ethanol were also used to extract phenolic compounds from olive leaves, and neat ethanol at 463 K proved to be optimal on-the-whole, but a 43:53 mixture at this temperature increased the yield of oleuropein [76]. Pressurized water and ethanol mixtures were also used for extraction from defatted sesame seeds, in this case a 63.5% ethanol mixture at 493 K led to maximal yields of flavones and other phenolic compounds with radical scavenging properties [77]. Brazilian pepper fruits were submitted to PLE with slightly acidified 54.2% aqueous ethanol at 348 K to yield the maximal amount of phenolic compounds [78]. Phenolic compounds and anthocyanines were extracted from juçara (*Euterpe edulis* Mrt.) residues using PLE with water, ethanol, their mixtures, and their acidified liquids. The highest yield of phenolic compounds was obtained with the acidified mixtures at 353 K [79].

Non-phenolic compounds were also obtained from natural materials by pressurized liquid extraction (PLE). Ketones of polycyclic aromatic hydrocarbons were obtained from contaminated

soil by PLE with ethanol at 373 K and 15 MPa. Precipitates formed after cooling the extract, so that dichloromethane was added to prevent this [80]. Polar and non-polar lipids were obtained by PLE from corn and oats with several solvents including ethanol. Only for the polar compounds were extractions with ethanol at 373 K more effective than with the other solvents (hexane, methylene chloride, and isopropanol) [81]. Capsaicinoids were extracted from peppers with pressurized methanol and ethanol containing up to 20% water at 323 to 473 K. The presence of the water had relatively little effect on the yields of the different capsaicinoids, whereas increasing temperatures increased the yields [82]. Charantin was extracted from *Momordica charantia* fruits by PLE with several solvents, including water and ethanol at 323 to 423 K and 10 MPa. Ethanol (also acetone) was found to be more effective than dichloromethane and water for this extraction [83]. A mixture of lactose and lactulose was separated (to obtain the lactulose component) by PLE with mixtures of water and ethanol at 313 to 403 K and 10 MPa. Optimal extractions were obtained at the lowest temperature with 70% ethanol [84]. Pressurized hot water at 373 and 393 K was used for the extraction of thermally labile bioactive compounds (gastrodin and vanillyl alcohol) from *Gastrodia elata* Blume, with results similar to those with reflux, but at a somewhat shorter time [85]. Pressurized extraction with at 403 K with 86% ethanol with water was found to be optimal for recovery of spicatoside A from *Liriope platyphyllia* [86]. The active constituents from ginger, *Zingiber cassumunar* Roxb rhizome, were recovered by PLE with methanol, ethanol, or water at 373 K for the alkanols and at 413 K for the water [87]. Another kind of ginger, *Zingiber officinale* Roscoe, was submitted to PLE with bio-ethanol or 70% ethanol in water at 373 K and 10 MPa with high efficiency [88]. Fucoxanthin was extracted most effectively from brown algae, *Eisenia bicyclis* (Kjellman), by pressurized 90% aqueous ethanol at 383 K [89]. A green microalga, *Chlorella ellipsoidea*, was subjected to PLE with ethanol at 389 K for the recovery zeaxanthin, this being more efficient than PLE with hexane or isopropanol [90]. Constituents, such as 1,8-cineol and α -terpinenyl, were effectively recovered from cardamom seeds by PLE with 75% aqueous ethanol at 363 K [91]. Olive pomace was the raw material for the recovery of sugars and lignin, the former of which being eventually converted to bio-ethanol, by PLE with water at 453 to 493 K [92].

Pressurized 88% aqueous ethanol at 513 K was more effective than at lower temperatures, down to 453 K, and more than pressurized water at 453 K for the PLE of carbohydrates from barley hulls, but phenolic compounds were best recovered by pressurized 59% aqueous *N*-methyl-2-hydroxyethyl-ammonium acetate at 433 K and 15 MPa [93]. Proteins and amino acids were extracted from porcine placenta by pressurized water at 443 K and 1MPa. The addition of ethanol to the extracting fluid was deleterious [94].

All these reports are summarized in Table 10.

Table 10. Pressurized liquid extraction of biomass with water, methanol, ethanol and their mixtures.

Pressurized Solvent	References
Antioxydants	
Water	[58,59,63,64,69,72]
Methanol	[63,67]
Ethanol	[58,59,61–64,66]
Water + ethanol	[65,70]
Phenolics	
Water	[74,76,77,79]
Methanol	[72]
Ethanol	[72,76,77,79]
Water + methanol	[72,73]
Water + ethanol	[72,74,75,78,79]
Non-Phenolic Compounds	
Water	[63,85,87,92,94]
Methanol	[82,87]
Ethanol	[80–83,87,88,90]
Water + ethanol	[84,86,88,89,91,93]

5. Supercritical Fluid Extraction from Bio-Materials

An extensive amount of work has been reported regarding the extraction with supercritical water, methanol, and ethanol, but the term “supercritical fluid extraction” generally pertains to extraction with supercritical carbon dioxide, possibly with modifiers, such as methanol and ethanol. This latter field of study is outside the scope of the present paper.

Cellulosic materials, including lignin, have been subjected to extraction with supercritical water, methanol, and ethanol. Japanese sugi wood (*Cryptomeria japonica*) was treated with supercritical water and various products could be identified in the water-soluble, methanol-soluble, and methanol-insoluble fractions [95]. Japanese beech (*Fagus crenata* Blume) was treated with supercritical methanol containing some water, and 10 vol % water at 623 K was found to be optimal for obtaining products from the lignocelluloses [96]. Japanese beech as well as oil palm trunks were treated with supercritical water at 653 K and 100 MPa to obtain low molecular size products [97]. German beech wood was treated with supercritical ethanol, optimally at 589 K, to produce bio-oil [98]. Holm oak was treated by water at 453 to 623 K to hydrolyze hemicellulose, and then supercritical water at 669 K at 24.5 MPa was used for extraction of low molecular mass products, e.g., lactic acid [99].

Non-wood cellulosic and lignin materials have also been subjected to supercritical fluid extraction, for example bagasse of sugar cane stalks, treated by supercritical ethanol at 603 K to yield bio-oil [100]. Supercritical water can deal directly with wet biomass (olive husk, cotton cocoon shell, and tea waste), without need for drying, to produce hydrogen and methane beside carbon oxides. The yields increase with temperature in the range 650 to 700 K and with pressure in the range 23 to 38 MPa [101]. Two recent reviews [102,103] dealt with the hydrolysis, fractionation, and extraction of biomass containing cellulose, hemicellulose, and lignin by sub- and supercritical water at 623 and 673 K. The yields of phenolics, for example, increased with increasing density (pressure) and temperature of the water. Lignin has been effectively converted to biofuel by treatment with supercritical methanol catalyzed by the layered (hydrotalcite-like) copper-doped magnesium-aluminium porous metal oxide [104].

Algae are a source of useful products when treated with supercritical fluids, e.g., supercritical methanol at 528 K that yields biodiesel from wet algae [105]. Direct transesterification of algal biomass under supercritical methanol at 533 K and microwave irradiation was compared [106]. Supercritical ethanol yields ethyl esters by transesterification when wet algae are treated by it at 533 K and microwave irradiation [107]. Supercritical water at 24 MPa and 936 K was used for the gasification of *Nannochloropsis gaditana* microalgae, producing mainly hydrogen, methane, and carbon dioxide [108]. The same microalgae as wet biomass were subjected to hydrothermal liquefaction with subcritical water at 623 K and subsequent extraction with supercritical water at 673 K in the presence of hydrogen gas to produce water-soluble biocrudes [109].

Other plant materials were also treated by supercritical water, methanol, or ethanol in order to recover valuable products. Supercritical water at 650 to 750 K and 23 to 48 MPa was used to produce hydrogen-rich gas from almond, hazelnut, walnut, sunflower shells and cotton cocoon as wet biomass [110]. Bullrush plants, *Typha latifolia*, were subjected to supercritical methanol and ethanol (also acetone and 2-butanol) for supercritical fluid extraction and liquefaction at 518 to 558 K [111]. Stinging nettle was subjected to supercritical ethanol extraction at 608 K, yielding various esters, phenols, indoles and nitrogen-containing compounds [112]. Another kind of thistle stalks, *Onopordum heteracanthum*, was subjected to liquefaction to produce bio-oils by means of supercritical methanol or ethanol (also acetone) at 523 to 563 K [113]. Red grape stems were used for the extraction of antioxidants by supercritical denatured ethanol up to 573 K [114]. Supercritical water at 923 K and 25 MPa was used for the gasification of glucose to produce hydrogen [115], consisting mainly of fatty acid esters [116]. Sea mango oil (*Cerbera odollam*) was subjected to transesterification by supercritical methanol, and the rate of the formation of the methyl esters of the fatty acid contents is shown in Figure 3 [117].

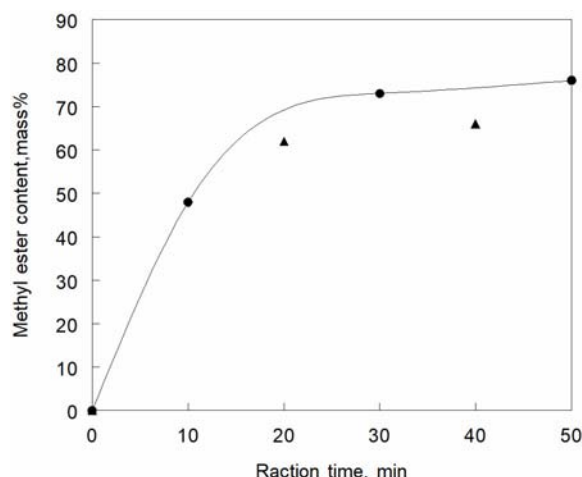


Figure 3. The formation of fatty acid methyl esters from sea mango oil by supercritical methanol at 633 K and methanol:oil ratio of 40 (circles) and at 653 K and methanol:oil ratio of 30 (triangles) [117].

All these reports are summarized in Table 11.

Table 11. Supercritical extraction of biomass with water, methanol, ethanol and their mixtures.

Pressurized Solvent	References
Cellulose, lignin	
Water	[95,97,99,101–103]
Ethanol	[98,100]
Water + methanol	[96]
Algae	
Water	[108,109]
Methanol	[105,106]
Ethanol	[107]
Other plant material	
Water	[108,110,111]
Methanol	[111,113]
Ethanol	[112–114,116]

6. Methods Used with Sub- and Supercritical Fluid Extraction

Several methods are being employed to enhance the yields and qualities of the products of solvent extraction processes, among which are solid micro-extraction and ultrasound- and microwave-assisted extraction. However, pressurized liquid extraction used with subcritical water, methanol, ethanol, and their mixtures as well as supercritical fluid extraction with these fluids are sufficiently powerful, so that these enhancement methods have only seldom been needed. Nevertheless, there are situations, for instance with temperature-sensitive products, for which lowering the extraction temperature and enhancement of the process with either one of these methods has proved useful.

Microwave assistance has been employed for the PLE with water of thermally labile bioactive compounds from *Gastrodia elata* Blume, shortening their residence time at the high temperature, 373 to 393 K [85]. Transesterification of algal biomass with supercritical methanol [106] and ethanol [107] at 533 K benefited from microwave irradiation. Phenolic compounds were extracted from *Equisetum arvense* L. herb with pressurized hot methanol and aqueous methanol, and ultrasound-assisted extraction with 80% methanol proved to be the optimal procedure [118]. The recovery of anthocyanins from various plant materials (blueberries, blackberries, grumixama

residues) by means of PLE at 353 K with water and aqueous ethanol alone and combined with ultrasound assistance was examined. The use of pressurized 70% ethanol combined with ultrasound assistance provided the largest total phenolics yield, but ultrasound-assisted extraction with this solvent at ambient conditions yielded the most anthocyanin product [119]. Ultrasound assistance combined with PLE with 80% aqueous methanol at 403 K and 13 MPa was used for the pretreatment of four varieties of algae for the recovery of phenolics from them [120].

In some cases, the efficiency of extraction by pressurized liquids or supercritical fluids is enhanced by the use of catalysts. Supercritical methanol and ethanol (also acetone) at 523 to 563 K, used for the liquefaction of *Onopordum heteracanthum* stalks for the production of bio-oils, was enhanced by the use of potassium hydroxide or zinc chloride as catalysts [106]. Catalytic amounts of potassium hydroxide were also needed for the desulfurization of low rank coal with supercritical ethanol at 623 K and 8.3 MPa of [23].

The optimization of the extraction processes with pressurized subcritical and with supercritical water, methanol, and their mixtures has been studied in many cases, using a central composite design [121] or response surface methodology [76,112,122]. In all these methods, the optimal temperature, choice of solvent (or its composition, in the case of aqueous mixtures), and the duration of the extraction process were sought. The best results of these methods are described in the previous sections.

References

1. Marcus, Y. *Supercritical Water*; Wiley: New York, NY, USA, 2012.
2. Teja, A.S.; Lee, B.J.; Rosenthal, D.; Anselme, M. Correlation of the critical properties of alkanes and alkanols. *Fluid Phase Equilib.* **1990**, *56*, 153–169. [[CrossRef](#)]
3. Lu, J.; Brown, J.S.; Liotta, C.L.; Eckert, C.A. Polarity and hydrogen bonding of ambient to near-critical water: Kamlet-Taft solvent parameters. *Chem. Commun.* **2001**, *7*, 665–666. [[CrossRef](#)]
4. Minami, K.; Mizura, M.; Suzuki, M.; Aizawa, T.; Aray, K. Determination of Kamlet-Taft solvent parameter π^* of high pressure and supercritical water by the UV-vis absorption spectral shift of 4-nitroanisole. *Phys. Chem. Chem. Phys.* **2006**, *8*, 2257–2264. [[CrossRef](#)] [[PubMed](#)]
5. Yerlett, T.K.; Wormald, C.J. The enthalpy of methanol. *J. Chem. Thermodyn.* **1986**, *18*, 719–726. [[CrossRef](#)]
6. Bazaev, A.R.; Abdulagatov, I.M.; Bazaev, E.A.; Abdurashidova, A.A.; Ramazanov, A.E. PVT measurements for pure methanol in the near-critical and supercritical regions. *J. Supercrit. Fluids* **2007**, *41*, 217–226. [[CrossRef](#)]
7. Thompson, J.D.; Carr, P.W. High-speed liquid chromatography by simultaneous optimization of temperature and eluent composition. *Anal. Chem.* **2002**, *74*, 4150–4159. [[CrossRef](#)] [[PubMed](#)]
8. Teutenberg, T.; Wiese, S.; Wagner, P.; Gmehling, J. High temperature liquid chromatography. Part III. Determination of the static permittivities of pure solvents and binary solvent mixtures—Implications for liquid chromatographic separations. *J. Chromatogr. A* **2009**, *1216*, 8480–8487. [[CrossRef](#)] [[PubMed](#)]
9. Vine, M.D.; Wormland, C.J. The enthalpy of ethanol. *J. Chem. Thermodyn.* **1989**, *21*, 1151–1157. [[CrossRef](#)]
10. Marcus, Y. Some thermophysical properties of methanol and aqueous methanol mixtures at sub- and supercritical conditions. *J. Mol. Liq.* **2017**, *239*, 10–17. [[CrossRef](#)]
11. Bazaev, A.R.; Abdulagatov, I.M.; Bazaev, E.A.; Abdurashidova, A. (p, v, T, x) measurements of $\{(1 - x)\text{H}_2\text{O} + x\text{C}_2\text{H}_5\text{OH}\}$ mixtures in the near-critical and supercritical regions. *J. Chem. Thermodyn.* **2007**, *39*, 385–411. [[CrossRef](#)]
12. Dillon, H.E.; Penoncello, S.G. A fundamental equation for calculation of the thermodynamic properties of ethanol. *Int. J. Thermophys.* **2004**, *25*, 321–335. [[CrossRef](#)]
13. Lu, J.; Boughner, E.C.; Liotta, C.L.; Eckert, C.A. Near-critical and supercritical ethanol as a benign solvent: Polarity and hydrogen bonding. *Fluid Phase Equilib.* **2002**, *198*, 37–49. [[CrossRef](#)]
14. Jasper, J.J. The surface tension of pure liquid compounds. *J. Phys. Chem. Ref. Data* **1972**, *1*, 841–1009. [[CrossRef](#)]
15. Bazaev, A.R.; Abdulagatov, I.M.; Bazaev, E.A.; Abdurashidova, A. PVT measurements for pure ethanol in the near-critical and supercritical regions. *Int. J. Thermophys.* **2007**, *28*, 194–211. [[CrossRef](#)]

16. Bazaev, A.R.; Bazaev, E.A.; Abdurashidova, A.S. The thermal properties of a water-methanol system with a composition of 0.5 mass fraction at temperatures from 373.15 to 673.15 K and pressures up to 60 MPa. *High Temp.* **2004**, *42*, 895–900. [[CrossRef](#)]
17. Bazaev, A.R.; Karabekova, B.K.; Abdurashidova, A.A. p , ρ , T and x dependences for supercritical water—Aliphatic alcohol mixtures. *Russ. J. Phys. Chem. B* **2013**, *7*, 955–967. [[CrossRef](#)]
18. Lee, S.; Kesavan, S.K.; Ghosh, A.; Fullerton, K.L. Selective precombustion desulfurization of Ohio coals using supercritical fluids. *Fuel* **1989**, *68*, 1210–1213. [[CrossRef](#)]
19. Murdie, N.; Hippo, E.J.; Tao, W.; Muchmore, C.; Kent, A. Supercritical desulfurization kinetics of Illinois No. 5 coals. *Fuel Proc. Technol.* **1988**, *18*, 119–122. [[CrossRef](#)]
20. Azzam, F.O.; Lee, S. A comparison of alcohol-water blends in their supercritical desulfurization efficiency of Midwestern U.S. bituminous coals. *Fuel Sci. Technol. Int.* **1993**, *11*, 951–973. [[CrossRef](#)]
21. Li, W.; Guo, S. Supercritical desulfurization of high rank coal with alcohol/water and alcohol/KOH. *Fuel Proc. Technol.* **1996**, *46*, 143–155. [[CrossRef](#)]
22. Meffe, S.; Perkson, A.; Trass, O. Coal beneficiation and organic sulfur removal. *Fuel* **1996**, *75*, 25–30. [[CrossRef](#)]
23. Charutawai, K.; Ngamprasertsith, S.; Prasassarakich, P. Supercritical desulfurization of low rank coal with ethanol/KOH. *Fuel Proc. Technol.* **2003**, *84*, 207–216. [[CrossRef](#)]
24. Hippo, E.J.; Murdie, N.; Chen, J.W.; Muchmore, C.B.; Kent, A.C. Microscopic characterization of coal residues from supercritical desulfurization of Illinois coal. *Fuel Proc. Technol.* **1987**, *17*, 85–102. [[CrossRef](#)]
25. Li, L.; Egiebor, N.O. Oxygen removal from coal during supercritical water and toluene extraction. *Energy Fuels* **1992**, *6*, 35–40. [[CrossRef](#)]
26. Jezko, J.; Gray, D.; Kershaw, J.R. The effect of solvent properties on the supercritical extraction of coal. *Fuel Proc. Technol.* **1982**, *5*, 229–239. [[CrossRef](#)]
27. Vasilakos, N.P.; Dobbs, J.M.; Parisi, A.S. Solvent effects in supercritical extraction of coal. *Ind. Eng. Chem. Proc. Des. Dev.* **1985**, *24*, 121–128. [[CrossRef](#)]
28. Amestica, L.A.; Wolf, E.E. Supercritical toluene and ethanol extraction of Illinois No. 6 coal. *Fuel* **1984**, *63*, 227–230. [[CrossRef](#)]
29. Lanças, F.M.; Pereira, D.M. Supercritical fluid extraction of Brazilian coal with alcohols. *Energy Sources* **1999**, *21*, 355–365.
30. Goetz, G.; Hammer, H.; Wolf, M. Supercritical fluid extraction of high volatile bituminous coals from the Saar region, Germany. Influence of extraction parameters on extract yield and composition. *Coal Sci. Technol.* **1995**, *24*, 1403–1406.
31. Dariva, C.; de Oliveira, J.V.; Vale, M.G.R.; Caramao, E.B. Supercritical fluid extraction of a high-ash Brazilian coal. *Fuel* **1997**, *76*, 585–591. [[CrossRef](#)]
32. Deshpande, G.V.; Holder, G.D.; Bishop, A.A.; Gopal, J.; Wender, I. Extraction of coal using supercritical water. *Fuel* **1984**, *63*, 956–961. [[CrossRef](#)]
33. Berkowitz, N.; Calderon, J. On “partial” coal conversion by extraction with supercritical H₂O. *Fuel Proc. Technol.* **1987**, *16*, 245–256. [[CrossRef](#)]
34. Canel, M.; Missal, P. Extraction of solid fuels with sub- and supercritical water. *Fuel* **1994**, *73*, 1776–1780. [[CrossRef](#)]
35. Hu, H.; Guo, S.; Hedden, K. Extraction of lignite with water in sub- and supercritical states. *Fuel Proc. Technol.* **1998**, *53*, 269–277. [[CrossRef](#)]
36. Luik, H.; Luik, L. Extraction of fossil fuels with sub- and supercritical water. *Energy Sources* **2001**, *23*, 449–459. [[CrossRef](#)]
37. Ga-Hourcade, M.L.; Torrents, C.; Galan, M.A. Study of the solubility of kerogen from oil shales (Puertollano, Spain) in supercritical toluene and methanol. *Fuel* **2007**, *86*, 698–705. [[CrossRef](#)]
38. Wu, B.; Hu, H.; Huang, S.; Fang, Y.; Li, X.; Meng, M. Extraction of weakly reductive and reductive coals with sub- and supercritical water. *Energy Fuels* **2008**, *22*, 3944–3948. [[CrossRef](#)]
39. Vostrikov, A.A.; Fednaeva, O.N.; Dubov, D.Y.; Psarov, S.A.; Sokol, M.Y. Conversion of brown coal in supercritical water without and with addition of oxygen at continuous supply of coal-water slurry. *Energy* **2011**, *36*, 1948–1955. [[CrossRef](#)]
40. Vostrikov, A.A.; Shishkin, A.V.; Sokol, M.Y.; Dubov, D.Y.; Fedyaeva, O.N. Conversion of brown coal continuously supplied into the reactor as coal-water slurry in a supercritical water and water-oxygen mixture. *J. Supercrit. Fluids* **2016**, *107*, 707–714. [[CrossRef](#)]

41. Park, J.H.; Son, S.H. Extraction of bitumen with sub- and supercritical water. *Korean J. Chem. Eng.* **2011**, *28*, 455–460. [[CrossRef](#)]
42. Lan, R.; Jin, H.; Guo, L.; Ge, Z.; Guo, S.; Zhang, X. Hydrogen production by catalytic gasification of coal in supercritical water. *Energy Fuels* **2014**, *28*, 6911–6917. [[CrossRef](#)]
43. Yang, R.; Chen, Y.; Wu, Y.; Hua, D.; Yang, M.; Li, C.; Chen, Z.; Liu, J. Production of liquid fuel via coliquefaction of coal and *Dunaliella tertiolecta* in a sun-/supercritical water-ethanol system. *Energy Fuels* **2013**, *27*, 2619–2627. [[CrossRef](#)]
44. Houser, T.J.; Tiffany, D.M.; Li, Z.; McCarville, M.E.; Houghton, M.E. Reactivity of some organic compounds with supercritical water. *Fuel* **1986**, *65*, 827–832. [[CrossRef](#)]
45. Garcia, R.; Moineiro, S.R.; Snape, C.E. A spectroscopic study of structural changes occurring in supercritical gas extraction of a Spanish lignite with alcohols. *Fuel* **1993**, *72*, 427–433. [[CrossRef](#)]
46. Jin, H.; Liu, S.; Wei, W.; Zhang, D.; Cheng, Z.; Guo, L. Experimental investigation of hydrogen production by anthracene gasification in supercritical water. *Energy Fuels* **2015**, *29*, 6342–6346. [[CrossRef](#)]
47. Shing, S.-L.; McKay, J.F. Interaction of supercritical methanol and water with organic material in Green River oil shale. *Fuel Sci. Technol. Int.* **1987**, *5*, 513–541.
48. McKay, F.F.; Blanche, M.S. Effect of time and pressure on recovery of oil from Green River oil shale in extraction experiments. *Fuel Sci. Technol. Int.* **1987**, *4*, 413–431. [[CrossRef](#)]
49. Funazukuri, T.; Yokoi, S.; Wakao, N. Supercritical fluid extraction of Chinese Maoming oil shale with water and toluene. *Fuel* **1988**, *67*, 10–16. [[CrossRef](#)]
50. Hu, H.; Zhang, J.; Guo, S.; Chen, G. Extraction of Huadian oil shale with water at sub- and supercritical states. *Fuel* **1999**, *78*, 645–651. [[CrossRef](#)]
51. Yanik, J.; Yüksel, M.; Sağlam, M.; Olukçu, M.; Bartle, K.; Frere, B. Characterization of the oil fractions of shale oil obtained by pyrolysis and supercritical water extraction. *Fuel* **1995**, *74*, 46–52. [[CrossRef](#)]
52. Olukçu, M.; Yanik, J.; Sağlam, M.; Yüksel, M. Solvent effect on the extraction of Beypazari oil shale. *Energy Fuels* **1999**, *13*, 895–902. [[CrossRef](#)]
53. El harfi, K.; Bennouna, C.; Mokhlisse, A.; Ben chanaa, M.; Lemèe, L.; Joffre, J.; Ambles, A. Supercritical fluid extraction of Moroccan (Timahdit) oil shale with water. *J. Anal. Appl. Pyrolysis* **1999**, *50*, 163–174. [[CrossRef](#)]
54. Tucker, J.D.; Masri, B.; Lee, S. A comparison of retorting and supercritical extraction on El-Lajjun oil shale. *Energy Sources* **2000**, *22*, 453–463.
55. Popov, E.; Kalmykov, A.; Cheremisin, A.; Bychkov, A.; Bondarenkoi, T.; Morozov, N.; Karpov, I. Laboratory investigation of hydrous pyrolysis as ternary enhanced oil recovery method for Bazhenov formation. *J. Petrol. Sci. Eng.* **2017**, *156*, 852–857. [[CrossRef](#)]
56. Richter, B.E.; Jones, B.A.; Ezzell, J.L.; Porter, N.L. Accelerated solvent extraction: A technique for sample preparation. *Anal. Chem.* **1996**, *68*, 1033–1039. [[CrossRef](#)]
57. Fang, Z.; Xu, C. (Eds.) *Near-Critical and Supercritical Water and Their Applications for Biorefineries*; Springer: Dordrecht, the Netherlands, 2014.
58. Herrero, M.; Ibanez, E.; Senorans, J.; Cifuentes, A. Pressurized liquid extracts from *Spirulina platensis* microalga. Determination of their antioxidant activity and preliminary analysis by micellar electrokinetic chromatography. *J. Chromatogr. A* **2004**, *1047*, 195–203. [[CrossRef](#)]
59. Jaime, L.; Mendiola, J.A.; Herrero, M.; Soler-Rivas, C.; Santoyo, S.; Senorans, F.J.; Cifuentes, A.; Ibanez, E. Separation and characterization of antioxidants from *Spirulina platensis* microalga combining pressurized liquid extraction, TLC, and HPLC-DAD. *J. Sep. Sci.* **2005**, *28*, 2111–2119. [[CrossRef](#)] [[PubMed](#)]
60. Herrero, M.; Martin-Alvarez, P.J.; Senorans, F.J.; Cifuentes, A.; Ibanez, E. Optimization of accelerated solvent extraction of antioxidants from *Spirulina platensis* microalgae. *Food Chem.* **2005**, *93*, 417–423. [[CrossRef](#)]
61. Herrero, M.; Jaime, L.; Martinez-Alvarez, P.J.; Cifuentes, A.; Ibanez, E. Optimization of the extraction of antioxidants from *Dunaliella salina* microalga by pressurized liquids. *J. Agric. Food Chem.* **2006**, *54*, 5597–5603. [[CrossRef](#)] [[PubMed](#)]
62. Jaime, L.; Rodriguez-Meizoso, I.; Cifuentes, A.; Santoyo, S.; Suarez, S.; Ibanez, E.; Senorans, F.J. Pressurized liquids as an alternative process to antioxidant carotenoids' extraction from *Haematococcus pluvialis* microalgae. *LWT Food Sci. Technol.* **2010**, *43*, 105–112. [[CrossRef](#)]
63. Pineiro, Z.; Palma, M.; Barroso, C.G. Determination of catechins by means of extraction with pressurized liquids. *J. Chromatogr. A* **2004**, *1026*, 19–23. [[CrossRef](#)]

64. Herrero, M.; Temirzoda, T.N.; Segura-Carretero, A.; Quirantes, R.; Plaza, M.; Ibanez, E. New possibilities for the valorization of olive oil by products. *J. Chromatogr. A* **2011**, *1218*, 7511–7520. [[CrossRef](#)] [[PubMed](#)]
65. Shang, Y.F.; Kim, S.M.; Um, B.-H. Optimization of pressurized liquid extraction of antioxidants from black bamboo leaves. *Food Chem.* **2014**, *154*, 164–170. [[CrossRef](#)] [[PubMed](#)]
66. Shang, Y.F.; Xu, J.-L.; Um, B.-H. Antioxidative polyphenolics obtained from spent coffee grounds by pressurized liquid extraction. *South Afr. J. Bot.* **2017**, *109*, 75–80. [[CrossRef](#)]
67. Casazza, A.A.; Aliakbarian, B.; Sannita, E.; Perego, P. High pressure high temperature extraction of phenolic compounds from grape skins. *Int. J. Food Sci. Technol.* **2012**, *47*, 399–405. [[CrossRef](#)]
68. Ju, Z.Y.; Howard, L.R. Subcritical water and sulfured water extraction of anthocyanins and other phenolics from dried red grape skin. *J. Food. Sci.* **2005**, *70*, S270–S276. [[CrossRef](#)]
69. Stavikova, L.; Polovka, M.; Hohnova, B.; Karasek, P.; Roth, M. Antioxidant activity of grape skin aqueous extracts from pressurized hot water extraction combined with electron paramagnetic resonance spectroscopy. *Talanta* **2011**, *85*, 2233–2240. [[CrossRef](#)] [[PubMed](#)]
70. Otero-Pareja, M.J.; Casas, L.; Fernandez-Ponce, M.T.; Mantell, C.; Martinez de la Ossa, E.J. Green extraction of antioxidants from different varieties of red grape pomace. *Molecules* **2015**, *20*, 9686–9702. [[CrossRef](#)] [[PubMed](#)]
71. Santos, D.T.; Veggi, P.C.; Meireles, M.A.A. Optimization and economic evaluation of pressurized liquid extraction of phenolic compounds from jaboticaba skins. *J. Food Eng.* **2012**, *108*, 444–452. [[CrossRef](#)]
72. Luthna, D.L.; Mukhopadhyay, S.; Kwansa, A.L. A systematic approach for extraction of phenolic compounds using parsley (*Petroselinum crispum*) flakes as a model substrate. *J. Sci. Food Agric.* **2006**, *86*, 1350–1358.
73. Mukhopadhyay, S.; Luthna, D.L.; Robbins, R.J. Optimization of extraction process for phenolic acids from black cohosh (*cimicifuga racemose*) by pressurized liquid extraction. *J. Sci. Food Agric.* **2006**, *86*, 156–162. [[CrossRef](#)]
74. Howard, L.; Pandjattan, N. Pressurized liquid extraction of flavonoids from spinach. *J. Food Sci.* **2008**, *73*, C151–C156. [[CrossRef](#)] [[PubMed](#)]
75. Lozano-Sanchez, J.; Castro-Puyana, M.; Mendiola, J.A.; Segura-Carretewro, A.; Cifuentes, A.; Ibanez, E. Recovering bioactive compounds from olive oil filter cake by advanced extraction techniques. *Int. J. Mol. Sci.* **2014**, *15*, 16270–16283. [[CrossRef](#)] [[PubMed](#)]
76. Xyntos, N.; Papaefstathiou, G.; Gikas, E.; Argyropoulou, A.; Aligiannis, N.; Skaltsounis, A.-L. Design optimization study of the extraction of olive leaves performed with pressurized liquid extraction using response surface methodology. *Sep. Purif. Technol.* **2014**, *122*, 323–330. [[CrossRef](#)]
77. Bodoira, R.; Velez, A.; Andreatta, A.E.; Martinez, M.; Maestri, D. Extraction of bioactive compounds from sesame (*Sesamum indicum* L.) defatted seeds using water and ethanol under sub-critical conditions. *Food Chem.* **2017**, *237*, 114–120. [[CrossRef](#)] [[PubMed](#)]
78. Feuereisen, M.M.; Barraza, M.G.; Zimmermann, B.F.; Schieber, A.; Schulze-Kaysers, N. Pressurized liquid extraction of anthocyanins and biflavonoids from *Schinus terebinthifolius*: A multivariate optimization. *Food Chem.* **2017**, *214*, 564–571. [[CrossRef](#)] [[PubMed](#)]
79. Del Pilar Garcia-Mendoza, M.; Espinosa-Pardo, F.; Baseggio, A.M.; Barbero, G.F.; Marostica, M.R., Jr.; Rostagno, M.A.; Martinez, J. Extraction of phenolic compounds and anthocyanins from juçara (*Euterpe edulis* Mrt.) residues using pressurized liquids and supercritical fluids. *J. Supercrit. Fluids* **2017**, *119*, 9–16. [[CrossRef](#)]
80. Zdrahal, Z.; Karasek, P.; Lojkova, L.; Buckova, M.; Vecera, Z.; Vejrosta, J. Pressurized extraction of ketones of polycyclic aromatic hydrocarbons from soil. *J. Chromatogr. A* **2000**, *893*, 201–206. [[CrossRef](#)]
81. Moreau, R.A.; Powell, M.J.; Singh, V. Pressurized liquid extraction of polar and nonpolar lipids in corn and oats with hexane, methylene chloride, isopropanol, and ethanol. *J. Am. Oil Chem. Soc.* **2003**, *80*, 1063–1067. [[CrossRef](#)]
82. Bebero, G.F.; Palma, M.; Barroso, C.G. Pressurized liquid extraction of capsaicinoids from peppers. *J. Agric. Food Chem.* **2006**, *54*, 3231–3236. [[CrossRef](#)] [[PubMed](#)]
83. Pitipanapong, J.; Chitprasert, S.; Goto, M.; Jiratchariyakul, W.; Sasaki, M.; Shotipruk, A. New approach for extraction of charantin from *Momordica charantia* with pressurized liquid extraction. *Sep. Purif. Technol.* **2007**, *52*, 416–422. [[CrossRef](#)]

84. Ruiz-Matute, A.I.; Sanz, M.L.; Corzo, N.; Martin-Alvarez, P.J.; Ibanez, E.; Martinez-Castro, I.; Olano, A. Purification of lactulose from mixtures with lactose using pressurized liquid extraction with ethanol-water at different temperatures. *J. Agric. Food Chem.* **2007**, *55*, 3346–3356. [[CrossRef](#)] [[PubMed](#)]
85. Teo, C.C.; Tan, S.N.; Yong, J.W.H.; Hew, C.S.; Ong, E.S. Evaluation of the extraction efficiency of thermally labile compounds in *Gastrodia elata* Blume by pressurized hot water extraction and microwave-assisted extraction. *J. Chromatogr. A* **2008**, *1182*, 34–40. [[CrossRef](#)] [[PubMed](#)]
86. Kim, S.H.; Kim, H.K.; Yang, E.S.; Lee, K.Y.; Kim, S.D.; Kim, Y.C.; Sung, S.H. Optimization of pressurized liquid extraction for spicatoside A in *Liriope platyphyllia*. *Sep. Purif. Technol.* **2010**, *71*, 168–172. [[CrossRef](#)]
87. Chienthavorn, O.; Poonsukcharoen, T.; Pathrakorn, T. Pressurized liquid and superheated water extraction of active constituents from *Zingiber cassumunar* Roxb rhizome. *Sep. Sci. Technol.* **2011**, *46*, 616–624. [[CrossRef](#)]
88. Hu, J.; Guo, Z.; Glasius, K.; Kristensen, K.; Xiao, L.; Xu, X. Pressurized liquid extraction of ginger (*Zingiber officinale* Roscoe) with bioethanol: An efficient sustainable approach. *J. Chromatogr. A* **2011**, *1218*, 5765–5773. [[CrossRef](#)] [[PubMed](#)]
89. Shang, Y.F.; Kim, S.M.; Lee, W.J.; Um, B.-H. Pressurized liquid method for fucoxanthin extraction from *Eisenia bicyclis* (Kjellman) setchell. *J. Biosci. Bioeng.* **2011**, *111*, 237–241. [[CrossRef](#)] [[PubMed](#)]
90. Koo, S.Y.; Cha, K.H.; Song, D.-G.; Chung, D.; Pan, C.-H. Optimization of pressurized liquid extraction of zeaxanthin from *Chlorella ellipsoidea*. *J. Appl. Phycol.* **2012**, *24*, 425–730. [[CrossRef](#)]
91. Eikani, M.H.; Golmohammad, F.; Amoli, H.S.; Sadr, Z.B. An experimental design approach for pressurized liquid extraction from cardamom seeds. *Sep. Sci. Technol.* **2013**, *48*, 1194–1200. [[CrossRef](#)]
92. Kazan, A.; Celiktas, M.S.; Sargin, S.; Yesil-Celiktas, O. Bio-based fractions by hydrothermal treatment of olive pomace: Process optimization and evaluation. *Energy Convers. Manag.* **2015**, *103*, 366–373. [[CrossRef](#)]
93. Sarkar, S.; Alvarez, V.H.; Saldana, M.D.A. Relevance of ions in pressurized fluid extraction of carbohydrates and phenolics from barley hull. *J. Supercrit. Fluids* **2014**, *93*, 27–37. [[CrossRef](#)]
94. Park, S.H.; Kim, J.-H.; Min, S.-G.; Jo, Y.-J.; Chun, J.-Y. Effects of ethanol addition on the efficiency of subcritical water extraction of proteins and amino acids from porcine placenta. *Korean J. Food Sci. Anim. Resour.* **2015**, *35*, 265–271. [[CrossRef](#)] [[PubMed](#)]
95. Takada, D.; Ehara, K.; Saka, S. Gas chromatographic and mass spectrometric (GC-MS) analysis of lignin-derived products from *Cryptomeria japonica* treated in supercritical water. *J. Wood Sci.* **2004**, *50*, 253–259. [[CrossRef](#)]
96. Minami, E.; Saka, S. Decomposition behavior of woody biomass in water-added supercritical methanol. *J. Wood Sci.* **2005**, *51*, 395–400. [[CrossRef](#)]
97. Varman, M.; Saka, S. A comparative study of oil palm and Japanese beech on their fractionation and characterization as treated by supercritical water. *Waste Biomass Valor.* **2011**, *2*, 309–315. [[CrossRef](#)]
98. Akalin, M.K.; Karagöz, S.; Akyüz, M. Supercritical ethanol extraction of bio-oils from German beech wood: Design of experiments. *Ind. Crops Prod.* **2013**, *49*, 720–729. [[CrossRef](#)]
99. Piqueras, C.M.; Cabeza, A.; Gallina, G.; Cantero, D.A.; Garcia-Serna, J.; Cocero, M.J. Online integrated fractionation-hydrolysis of lignocellulosic biomass using sub- and supercritical water. *Chem. Eng. J.* **2017**, *308*, 110–125. [[CrossRef](#)]
100. Chumpoo, J.; Prasassarakich, P. Bio-oil from hydr-liquefaction of bagasse in supercritical ethanol. *Energy Fuels* **2014**, *24*, 2071–2077. [[CrossRef](#)]
101. Demirbaş, A. Hydrogen production from biomass via supercritical water extraction. *Energy Sources* **2005**, *27*, 1409–1417. [[CrossRef](#)]
102. Knez, Ž.; Hrnčič, M.K.; Čolnik, M.; Škerget, M. Chemicals and value added compounds from biomass using sub- and supercritical water. *J. Supercrit. Fluids* **2017**. [[CrossRef](#)]
103. Cocero, M.J.; Cabeza, A.; Abad, N.; Adamovic, T.; Vaquerizo, L.; Martinez, C.M.; Pazo-Cepeda, M.V. Understanding biomass fractionation in subcritical and supercritical water. *J. Supercrit. Fluids* **2017**. [[CrossRef](#)]
104. Warner, G.; Hansen, T.S.; Riisager, A.; Beach, E.S.; Barta, K.; Anastas, P.T. Depolymerization of organosolv lignin using doped porous metal oxides in supercritical methanol. *Bioresour. Technol.* **2014**, *161*, 78–83. [[CrossRef](#)] [[PubMed](#)]
105. Patil, P.D.; Gude, V.G.; Mannarswamy, A.; Deng, S.; Cooke, P.; Munson-McGee, S.; Rhodes, I.; Lammers, P.; Nirmalakhandan, N. Optimization of direct conversion of wet algae to biodiesel under supercritical methanol conditions. *Bioresour. Technol.* **2011**, *102*, 118–122. [[CrossRef](#)] [[PubMed](#)]

106. Patil, P.D.; Gude, V.G.; Mannarswamy, A.; Cooke, P.; Nirmalakhandan, N.; Lammers, P.; Deng, S. Comparison of direct transesterification of algal biomass under supercritical methanol and microwave irradiation conditions. *Fuel* **2012**, *97*, 822–831. [[CrossRef](#)]
107. Patil, P.D.; Reddy, H.; Muppaneni, T.; Schaub, T.; Holguin, F.O.; Cooke, P.I.; Lammers, P.; Nirmalakhandan, N.; Li, Y.; Lu, X.; et al. In situ ethyl ester production from wet algae biomass under microwave-mediated supercritical ethanol conditions. *Bioresour. Technol.* **2013**, *139*, 308–315. [[CrossRef](#)] [[PubMed](#)]
108. Caputo, G.; Dispenza, M.; Rubio, P.; Scargiali, F.; Marotta, G.; Brucato, A. Supercritical water gasification of microalgae and their constituents in a continuous reactor. *J. Supercrit. Fluids* **2016**, *118*, 163–170. [[CrossRef](#)]
109. Xu, D.; Savage, P.E. Supercritical water upgrading of water-insoluble and water-soluble biocrudes from hydrothermal liquefaction of *Nannochloropsis* microalgae. *J. Supercrit. Fluids* **2017**. [[CrossRef](#)]
110. Demirbas, A. Hydrogen-rich gas from fruit shells via supercritical water extraction. *Int. J. Hydrog. Technol.* **2004**, *29*, 1237–1243. [[CrossRef](#)]
111. Aysu, T.; Turhan, M.; Kicuk, M.M. Liquefaction of *Typha latifolia* by supercritical fluid extraction. *Bioresour. Technol.* **2012**, *107*, 464–470. [[CrossRef](#)] [[PubMed](#)]
112. Akalin, M.K.; Karagöz, S.; Akyüz, M. Application of response surface methodology to extract yields from stinging nettle under supercritical ethanol conditions. *J. Supercrit. Fluids* **2013**, *84*, 164–172. [[CrossRef](#)]
113. Durak, H.; Aysu, Y. Effects of catalysts and solvents on liquefaction of *Onopordum heteracanthum* for production of bio-oils. *Bioresour. Technol.* **2014**, *166*, 309–317. [[CrossRef](#)] [[PubMed](#)]
114. Wenzel, J.; Samaniego, C.S.; Wang, L.; Nelson, L.; Ketchum, K.; Ammermann, M.; Zand, A. Superheated liquid and supercritical denatured ethanol extraction of antioxidants from crimson red grape stems. *Food Sci. Nutr.* **2015**, *3*, 569–576. [[CrossRef](#)] [[PubMed](#)]
115. Caputo, G.; Rubio, P.; Scargiali, F.; Marotta, G.; Brucato, A. Experimental and fluid dynamic study of continuous supercritical water gasification of glucose. *J. Supercrit. Fluids* **2016**, *107*, 450–461. [[CrossRef](#)]
116. Tekin, K.; Akalin, M.K.; Karagöz, S. Experimental design for extraction of bio-oils from flax seeds under supercritical ethanol conditions. *Clean Technol. Environ. Policy* **2016**, *18*, 461–471. [[CrossRef](#)]
117. Ang, G.T.; Ooi, S.N.; Tan, K.T.; Lee, K.T.; Mohamed, A.R. Optimization and kinetic studies of sea mango (*Cerbera odollam*) oil for biodiesel production via supercritical reaction. *Energy Convers. Manag.* **2015**, *99*, 242–251. [[CrossRef](#)]
118. Oniszczyk, A.; Podgorski, R.; Oniszczyk, T.; Zukiewicz-Sobczak, W.; Nowak, R.; Waksmundzka-Hajnos, M. Extraction methods for the determination of phenolic compounds from *Equisetum arvense* L. herb. *Ind. Crops Prod.* **2014**, *61*, 377–381. [[CrossRef](#)]
119. Machado, A.P.D.F.; Pereira, A.L.D.; Barbero, G.F.; Martinez, J. Recovery of anthocyanins from residues of *Rubus fruticosus*, *Vaccinium myrtillus*, and *Eugenia brasiliensis* by ultrasound assisted extraction, pressurized liquid extraction, and their combination. *Food Chem.* **2017**, *231*, 1–10. [[CrossRef](#)] [[PubMed](#)]
120. Klejdus, B.; Plaza, M.; Snoblova, M.; Lojkova, L. Development of new efficient method for isolation of phenolics from sea algae prior to their rapid resolution liquid chromatographic-tandem mass spectrometric determination. *J. Pharm. Biomed. Anal.* **2017**, *135*, 87–95. [[CrossRef](#)] [[PubMed](#)]
121. Kraujalis, P.; Venskutonis, P.R.; Ibanez, E.; Herrero, M. Optimization of rutin isolation from *Amaranthus paniculatus* leaves by high pressure extraction and fractionation techniques. *J. Supercrit. Fluids* **2015**, *104*, 234–242. [[CrossRef](#)]
122. Syahariza, Z.A.; Torkamani, A.E.; Norziah, H.M.; Mahmood, W.A.K.; Juliano, P. Optimization of pressurized liquid extraction for antioxidative polyphenolic compound from *Momordica charantia* using response surface methodology. *Food Sci. Technol.* **2017**, *52*, 480–493.

