

Article

Specific Controlling Essential Oil Composition of Basil (*Ocimum basilicum* L.) Involving Low-Temperature, Low-Pressure Glow Plasma of Low Frequency

Wojciech Ciesielski ^{1,*} , Maciej Gąstoł ² , Damian Kulawik ¹, Zdzisław Oszczyda ³, Elżbieta Pisulewska ⁴ and Piotr Tomasiak ³

¹ Institute of Chemistry, Jan Długosz University, Armii Krajowej Ave. 13-15, 42 201 Częstochowa, Poland; d.kulawik@ujd.edu.pl

² Faculty of Biotechnology and Horticulture, University of Agriculture, 31-120 Krakow, Poland; rogastol@cyfronet.pl

³ Nantes Nanotechnological Systems, Dolnych Młynów Str. 24, 59 700 Bolesławiec, Poland; z.oszczyda@nantes.com (Z.O.); rrtomasi@cyf-kr.edu.pl (P.T.)

⁴ Institute of Health and Economy, State Vocational College, 38 400 Krosno, Poland; elzbieta.pisulewska@gmail.com

* Correspondence: w.ciesielski@interia.pl

Received: 9 November 2020; Accepted: 25 November 2020; Published: 27 November 2020



Abstract: The effect of watering basil (*Ocimum basilicum* L.) with water treated with low-pressure, low-temperature glow plasma of low frequency (LPGP) on growth habits and plant metabolites was tested. Watering with the LPGP treated water was beneficial for sprouting basil seeds. Watering with non-treated water was advantageous solely for the number of leaves per plant and mass of one leaf. Watering with the LPGP treated water in contact with the air (LPGPA), nitrogen (LPGPN), carbon dioxide (LPGPC), and methane (LPGPM) increased the total yield of collected essential oil by 40%, 60%, 20%, and 20%, respectively. Watering with water treated under molecular oxygen (LPGPO) decreased that yield by 12.5%. A diverse effect of particular kinds of the LPGP treated water upon the composition of isolated essential oil was also noted.

Keywords: aromas; fragrances; glow plasma; herbal medicine; spices

1. Introduction

Our recent paper [1] demonstrated how the functional properties of essential oil from lavender (*Lavandula angustifolia*, L.) could be adjusted to its use as either a spice, fragrant for cosmetics, or in aroma therapy. It could be afforded by watering that plant with water treated with low-pressure, low-temperature glow plasma of low frequency (LPGP) [2,3]. Effects of watering depended on whether the treatment of water with LPGP was performed in contact with the air [4], nitrogen [5], carbon dioxide [6], molecular oxygen [7], or methane [8]

Thus, dihydrosabinene (thujone) dominated in essential oil from lavender watered with water treated under molecular oxygen (LPGPO). Essential oil collected from the plant watered with water treated under carbon dioxide (LPGPC) contained high level of pinenes. A high content of β -ocimene in essential oil from lavender watered with water treated in the air (LPGPA) made that oil a suitable as antifungal, antiviral, and anti-inflammatory product. For its high content of artemisole, essential oil from the plant watered with the water treated under molecular oxygen (LPGPO) could be recommended for curing various skin diseases. Whenever juniper aroma and the biological properties of herniarin

used in dyspepsia and inadequate bile secretion were required, essential oil from lavender watered with water treated under nitrogen (LPGPN) could be recommended. Essential oil from lavender watered with water treated under methane (LPGPM) contained a high level of camphor and *endo*-borneol dup-1. Such results rationalize assumption that such kinds of LPGP treated water could be useful in controlled modifications of functional properties of other plants and their preparations.

In the present studies results of such approach to basil *Ocimum basilicum* L. is demonstrated. Plantations of that herb were watered with the water treated with LPGP in the air, under nitrogen, carbon dioxide, molecular oxygen, and methane. The effects of watering are given in terms of the quantitative characteristics of the basil crops and composition of the essential oils extracted from particular basil samples.

Basil, a herb, belongs to the Lamiaceae (mints) family. Although it originates from the region of central Africa to Southeast Asia [9], its plantations can be found worldwide. There are several species and cultivars of that plant which differ from one another with their specific tender, taste, flavour and aroma. Basil (*Ocimum basilicum* L.) belongs to the most common variety. It grows best outdoors as well as indoor including basements provided it is exposed to sun and fluorescent light, respectively. That herb is commonly used as a spice adding flavor and aroma [10–14] but these functional properties may depend on the breeding regime [10]. The plant and its essential oil contain β -carotene and a number of biologically active terpenes, aldehydes, alcohols, esters, phenols, ethers, and ketones as well as such bioelements as magnesium, iron, calcium, and zinc [15–20]. The most recent characteristics of basil and its essential oil were presented by Stanojevic et al. [21].

Biological functions of aqueous solutions of *O. basilicum* L are related to their hypoglycemic and hypolipidaemic activity [16]. The latter is based on the inhibition of the α -glucosidase, which prevents the degradation of starch and sucrose, and consequently, control the absorption of level of blood sugar [22].

For its specific composition basil leaves and extracted essential oil are used for curing and inhibiting several diseases and health disorders [16,23]. The essential oil from basil showed antifungal and insect-repelling properties, [24] including potential toxicity to mosquitos [25]. In folk medicine, such as Ayurveda or traditional Chinese medicine, basil is thought to have therapeutic properties [26,27].

2. Materials and Methods

2.1. Materials

2.1.1. Basil

Seed material of basil ‘Marian’ was delivered by Enza Zaden Enterpraise (Enkhuizen, The Netherlands). As reported by the manufacturer, plants from those seeds provide dark green leaves of medium size and plants are resistant to diseases and tip burn and well tolerate low temperature and transport.

2.1.2. Substrate

Substrate was composed of medium size turf fraction Florabalt®Pot Medium-Coarse (Floragard, Oldenburg, Federal Republic of Germany). The medium of pH 5.6, contained 1.2 g/L total salts including 210 mg N/L, 120 mg P₂O₅/L, 260 mg K₂O/L. It was supplemented with multicomponent PG-Mix 18-10-20 fertilizer (1.20 kg/m³) (Yara, Oslo, Norway)

2.1.3. Water

Tap water from Bolesławiec of total hardness 129 mg/dm³ CaCO₃, pH 7.1, conductivity 334 μ S/cm, Fe < 50 μ g/dm³, Mn < 5 μ g/dm³ and 6.93 mg/dm³ dissolved oxygen was used as the standard. That water was LPGP treated for 30 min in contact with the air following to Białopiotrowicz et al. [4]

providing LPGPA and, alternatively, treated for the same time with LGPG under nitrogen as described by Chwastowski et al. [5], providing LPGPN.

LPGPC, LPGPO and LPGPM were prepared following methods described by Chwastowski et al. [6–8]. LPGP of 38 °C was generated at 5×10^{-3} mbar, 800 V, 50 mA and 10 KHz frequency in a plasmothrone patented by Oszczyda et al. [2,3]. The produced water was stored at ambient temperature in 1L closed teflon containers.

2.1.4. Trays

QP 15RW multiplates QP 15RW trays (Herkuplast Kubern GmbH, Ering/Inn, Federal Republic of Germany) were used. Each multiplate consisted of 3×5 trays. Each tray had 280 cm³ capacity. Further, 1 m² of greenhouse hosted 880 plants.

2.2. Methods

2.2.1. Basil plantation

The monofactorial experiment was carried out from Feb 24th (sowing) till May 18th (harvesting) 2019 in a greenhouse at the University of Agriculture in Cracow. Temperature in the greenhouse was set for 22 and 18 °C during the day and night, respectively. The daytime took 16 h from dawn. The passing from the day into the night regime was controlled with computer. The automatic additional 16h illumination with sodium lamps was used when natural light intensity decreased below 100 W/m². All plants were exposed to identical temperature, illumination, and humidity. The experiment involved three sets of trays with 24 pots each. Ten seeds of basil were sown into every pot. In one series of experiments 2 multiplates hosted 300 plants. In order to eliminate parietal effect 60 plants on the edge of trays were left apart and, therefore, only 240 plants were harvested. Since the experiments were run in triplicates maximum 720 plants were collected for a given series.

The watering was adjusted according to tensiometer readings (Irrometer model SR 150 mm) when soil water tension was <-40 kPa. The plants were watered by hand to avoid the accidental contact of water with leaves. Initially, plants consumed totally 3 L water, that is 1 L per each replication in the 5 day period until March 24th. In the subsequent 1-month period, the watering was intensified and the same amount of water was administered to the plants in 3-day periods. In the final period of breeding plants were watered daily consuming the same amount of water. In such manner the watering consumed totally 40 mL each kind water daily. The experiment terminated on May 18th when the plants were collected and separated into leaves and stems. The plants were then dried at 105 °C for 4 hours to determine dry mass of the crops.

2.2.2. Preparation of Extracts

Extracts were prepared on 30 min. grounding of the plant material in a mortar (20 g) with 96% ethanol (100 cm³) added.

2.2.3. Separation of Essential Oils

Samples of the plant dried at 35 °C to constant weight (1 g) were steam distilled in a Deryng apparatus with a closed water circulation. The collected oils were transferred to a closed vial and stored in dry ice until analyzed. Analysis was performed within three days.

2.2.4. Gas Chromatographic Analyses

Sample (5 µL) was transferred to closed chromatographic vial and evaporated on a heating plate. Using gas-tight syringe gaseous sample (10 µL) was analyzed using a Bruker 436-GC gas chromatograph coupled with Bruker SCION SQ (single quadruple, electron ionization) mass spectrometer (Durham, UK). The estimations were duplicated.

The instrument was equipped with BR-5ms; 0.25 nm × 30 m, $df = 0.25 \mu\text{m}$. The column operated at the following temperature programme: 50 °C (2 min) at the temperature rate increase 10 °C/min up to 170 °C (0 min), then at 25 °C to 280 °C (5 min).

Dispenser, transfer line, and the source temperature was 300, 280, and 200 °C, respectively. Sample separation was set for 1:20, helium was used as the carrier gas. The flow of the mobile phase was 1.0 mL/min, and ionization energy was 70 keV. Scanning was performed in the 50–500 m/z range.

Chromatographic signals were identified by comparison with mass spectra available in the NIST 11 library. The areas under particular chromatographic peaks were calculated using a computer programme installed in the chromatograph.

2.3. Statistics

The results were subjected to statistical interpretation, mean values and standard errors were calculated, and the significance of the variables was determined. Statistically significant differences between means ($p < 0.05$) were evaluated using one-way analysis of variance (ANOVA) with a post hoc multiply Duncan's range test [28]. Moreover, the Pearson product-moment correlation coefficients between analyzed variables were calculated. The significance level for correlation coefficient was $p = 0.05$, and the number of pairs for the calculations was $N = 216$. All statistical analyses were calculated using Statistica 13.3 software (Tibco Software Inc., Palo Alto, CA, USA).

3. Results and Discussion

The effect of watering depends, first of all, on the LPGP treated water macrostructure. On the treatment with LPGP that macrostructure was ruined forming smaller clusters. For that sake, water could more readily penetrate cell membranes. Simultaneously, declustreized water better solubilized several compounds. Thus, one could assume that water treated with LPGP could be better vector for dissolved components transporting them more efficiently to the flora and fauna cells.

The effect of the treatment of water with LPGP depended on the atmosphere in which the treatment was performed. Thus, solely LPGPA [4] and LPGPN [5] contained small structural units of aqueous clathrates. They hosted excited molecules of oxygen and nitrogen, respectively. Size of clathrates facilitated their permeation across cell membranes. LPGPA enriched interior of the cells in singlet molecular oxygen. Excited, singlet oxygen molecules liberated from their clathrates inside the cells released energy on returning to the normal triplet state. That energy could affect the course of cellular bioprocesses not necessarily resulting from the oxidation. LPGPN directed into the cells excited forms of molecular nitrogen.

All three LPGPC [6], LPGPO [7] and LPGPM [8] did not contain clathrates. Their macrostructures were stable to the extent dependent on proportion of involved hydrogen bonding configurations and the content of niches constituting these macrostructures. The macrostructure of LPGPM was relatively stable. It hosted methane molecules in its niches. In contrast to LPGPM, LPGPO carried in its niches molecular oxygen in the triplet state. The oxygen molecules participated in building the macrostructure. LPGPO clearly distinguished from the remaining kinds of water in its potential oxidative properties. LPGPC was built chiefly of surface tetrahedral and deformed tetrahedral structural units. The niches of its macrostructure incorporated O-free radicals of triplet carbon dioxide. Hence, one could anticipate different effects of those kinds of water upon the growth of *O. basilicum* L.

An insight into Table 1 provides an evidence that watering with LPGPA, LPGPN, LPGPC and LPGPO was beneficial for sprouting basil seeds. Solely LPGPM provided worse sprouting, although the effect was better than that noted for the plant watered with controlled non-treated water. The use of non-treated water was advantageous for the number of leaves per plant and mass of one leaf (Table 1). The benefit of watering with LPGP treated water was beneficial for the height of plants, total mass of crops, the total number of leaves, mass of stems, and total mass of foliage.

Table 1. Quantitative characteristics of the basil crops ^a.

Estimation	Plazmed Water					
	Non-Plazmed	LPGPA	LPGPN	LPGPC	LPGPM	LPGPO
Number of plants	3.04 ± 0.15	5.02 ± 0.16	4.79 ± 0.33	5.02 ± 0.12	4.96 ± 0.13	3.65 ± 0.31
Height of plants/pot (cm)	41.1 ± 4.0	56.4 ± 2.1	50.6 ± 2.7	57.6 ± 1.7	51.4 ± 2.1	44.6 ± 2.1
Total mass of plant (g)	11.52 ± 0.51	16.27 ± 0.77	15.14 ± 0.55	16.14 ± 0.23	16.11 ± 0.35	12.14 ± 0.53
Total number of leaves	24.9 ± 1.1	33.2 ± 1.2	32.4 ± 1.8	33.4 ± 1.4	32.4 ± 1.3	27.3 ± 1.1
Number of leaves/plant	8.39 ± 0.23	6.72 ± 0.26	7.14 ± 0.34	7.32 ± 0.24	6.95 ± 0.24	6.23 ± 0.23
Mass of stems (g)	2.69 ± 0.13	5.29 ± 0.19	4.06 ± 0.16	4.26 ± 0.21	4.84 ± 0.18	4.78 ± 0.13
Total mass of foliage (g)	8.33 ± 0.39	10.99 ± 0.64	11.08 ± 0.48	12.13 ± 0.38	11.98 ± 0.23	10.51 ± 0.32
Mass of one leaf (g)	0.359 ± 0.010	0.337 ± 0.019	0.351 ± 0.012	0.337 ± 0.018	0.348 ± 0.018	0.334 ± 0.017

^a Average number of plants collected from triplicated experiments involving 3 × 24 trays. Each tray contained 10 seeds. Presented data were recalculated for the number of plants in one tray.

The plants watered with the LPGP treated water were better shaped. The mass of the foliage was approximately 20% higher. Numbers of plants per one pot were about 20% higher. Similar effects of such watering were noted for the total mass of plants. The total number of leaves increased by approximately 40%. Although LPGP treated water favored formation of the leaves, the latter were slightly smaller. Watering basil with LPGPA, LPGPM, and LPGPO increased the mass of stems by approximately 60–70%. LPGPN and LPGPC provided hardly a 30–40% increase in the case of watering with LPGPN and LPGPC (Table 1).

Watering basil with LPGPA, LPGPN, LPGPC, and LPGPO considerably influenced the yield and composition of essential oils (Table 2). LPGPA, LPGPN, LPGPC, and LPGPM increased the total yield of collected essential oil by 40, 60, 20 and 20%, respectively, whereas LPGPO decreased that yield by 12.5%.

Stanojevic et al. [21] recognized and characterized 65 components of essential oil from *O. basilicum* L. In this paper only components of 0.01% and higher yield were taken under consideration. Thus, essential oil extracted from the plants watered with control, non-treated water consisted of 33 characterized components (Table 2). Although watering those plants with LPGPA, LPGPN, LPGPC, and LPGPM increased the yield of collected oil its composition was impoverished in the number of components to 19, 15, 25, and 25, respectively. Watering with LPGPO provided essential oil with 22 characterized components.

In the essential oil from basil watered with non-treated water dominated methyl eugenol (36.51% of the total), linalool (14.25%), eugenol (13.65%), 1,8-cineole (7.15%), and germacrene D (5.60%).

Methyl eugenol, a phenolic compound, usually plays a role of attracting pollinator and a component of floral fragrance. It has some antifungal activity. It also repels many insects [29]. In 2018, Federal Drug Administration withdrawn authorization for the use of methyl eugenol as a synthetic flavoring substance in food. It was found that methyl eugenol induced cancer in laboratory animals [30]. From 2021 any product containing over 0.01% methyl eugenol has to be stated as per the CPL regulations [31]. LPGPA, LPGPN and LPGPC considerably increased the content of that compound in essential oil whereas LPGPO and, particularly, LPGPM decreased it.

Linalool, unsaturated alcohol, is widely used in perfumery and as insecticide [32], however, it can evoke some allergic responses [33]. It is considered as a potential drug for curing in some cancer diseases [34–36]. LPGPA, LPGPO, and LPGPM significantly increased the content of linalool in the essential oil but LPGPC and, particularly LPGPN, decreased it drastically.

Eugenol, a phenol, a typical fragrant compound which disposes also with antiseptic and anaesthetic properties. It is utilized, among others, in stomatology [37] and as an anticoagulant for blood cells [38]. LPGPA, LPGPN, and LPGPO decreased content of eugenol in essential oil whereas LPGPC considerably increased it. LPGPM had no effect upon the level of eugenol in that oil.

Table 2. Composition of essential oil [% of the total = 100%] collected from basil watered with non-treated water, and water LPGP treated for 30 min either under air (LPGPA), nitrogen (LPGPN), carbon dioxide (LPGPC), molecular oxygen (LPGPO) and methane (LPGPM).

Peak Position in Chromatogram	Retention Time [min]	Component	Non Treated	LPGPA	LPGPN	LPGPC	LPGPO	LPGPM
1	7.08	β -Thujone	0.03	-	0.02	0.04	0.49	0.04
2	7.27	α -Pinene	0.11	0.03	-	0.03	0.16	0.09
3	7.68	Camphene	0.04	-	0.01	0.03	0.02	-
5	8.22	Sabinene	0.51	0.27	0.28	0.14	8.25	2.68
9	8.60	β -Pinene	2.31	2.44	0.21	0.48	2.09	1.92
10	8.79	α -Caryophyllene	-	-	0.03	1.28	-	-
15	9.37	p-Cymene	-	-	-	0.97	-	-
16	9.51	o-Cymene	0.17	0.14	2.55	2.31	0.05	0.06
17	9.63	D-Limonene	2.82	2.23	0.06	0.10	1.60	3.84
19	9.71	Eucalyptol	2.36	-	-	-	3.68	4.52
27	11.40	Linalool	14.25	18.54	1.85	8.15	19.16	18.42
29	11.98	Fenchone	0.14	-	-	0.25	-	-
32	12.50	α -Bulnesene	0.14	-	-	-	0.04	0.04
35	12.89	Fenchyl acetate	0.38	0.33	-	-	0.24	-
39	13.19	β -Cubebene	-	-	0.05	0.06	-	-
47	13.84	Isocaryophyllene	0.29	-	-	-	-	0.12
54	14.91	4-Carvomenthol	-	-	-	0.65	-	-
62	15.81	Carveol	0.18	-	-	-	0.68	-
63	15.82	Lavandulol acetate	-	-	-	1.48	-	1.25
64	15.92	Bornyl acetate	0.05	-	-	-	0.06	0.05
73	17.45	Eugenol	13.65	8.56	9.41	16.84	6.51	13.69
75	17.62	Methyl eugenol	36.51	40.58	58.62	41.85	35.41	23.77
77	17.96	Geranyl acetate	0.23	0.06	-	-	0.17	0.16
78	18.02	β -Elemene	0.34	-	-	3.25	2.19	-
79	18.03	Copaene	0.35	-	-	1.25	-	0.95
80	18.79	α -Bergamotene	0.95	3.24	-	4.27	-	1.68
82	18.97	Caryophyllene	1.25	0.54	-	0.82	1.30	1.30
85	19.06	β -Eudesmol	1.52	-	-	-	-	-
87	19.59	Geranyl propionate	0.57	2.38	-	-	3.28	4.62
90	20.02	α -Farnesene	2.36	1.89	5.23	-	1.20	0.18
92	20.16	γ -Cadinene	1.28	5.28	-	2.56	-	3.99
93	20.20	σ -Cadinene	0.30	-	-	-	-	-
94	20.74	1,8-Cineole	7.15	3.41	8.14	4.28	0.25	6.58
95	20.80	Germacrene D	5.60	1.57	0.02	0.02	-	0.06
96	21.03	1,10-di-epi-Cubenol	0.25	0.03	-	-	-	-
97	21.18	1-epi-Cubenol	0.15	-	-	0.62	0.38	0.45
98	21.21	τ -Cadinol	0.24	-	-	-	-	-
99	21.68	Estragole	3.52	8.48	4.52	8.27	12.79	9.54
Total number of components			33	19	15	25	22	25
Yield of essential oil (mL/100 g dry mass) ^a			0.5	0.7	0.8	0.6	0.4	0.6

^a In every case accuracy of the estimation was ± 0.02 . The dash in particular position denotes an absence of given compound in the corresponding essential oil.

The terpene 1,8-Cineole (eucalyptol) is used as an insecticide and insect repellent [39,40] and insect pheromone [41,42]. In higher-than-normal doses, eucalyptol is hazardous via either ingestion, skin contact or inhalation. It is classified as a reproductive toxin for females and a suspected reproductive toxin for males [43]. Anti-inflammatory properties of eucalyptol are also reported [44].

LPGPN increased the eucalyptol content in the essential oil, whereas the remaining types of treated water, particularly LPGPO, decreased it.

Germacrene D, a sesquiterpene, has antimicrobial and insecticidal properties [45]. It constituted 5.6% of the essential oil from basil watered with non-treated water. The watering with all LPGP treated kinds of water significantly reduced its level.

Watering basil with LPGPO enriched content of sabinene in the essential oil from 0.51% to 8.25%. It could be beneficial for the bactericidal properties of that monoterpene. Sabinene exhibits also anti-fungal activity against pathogenic fungi [46]. LPGPM elevated the sabinene content to 2.68% and the other kinds of LPGP treated water had no effect on it.

It is worth to mention that LGPGN increased by over twice the content of α -farnesene. In essential oil from basil watered with non-treated water, the content of α -farnesene reached hardly 2.36%. That terpene acts as alarm pheromone in termites [47] and food attractant for codling moth, the apple tree pest [48].

Essential oil from basil watered with non-treated water contained 3.52% estragole. LPGPN increased the content of that compound to 4.52% but LPGPC, LPGPA, LPGPM and LPGPO rose that content to 8.27%, 8.48%, 9.54%, and 12.79%, respectively. These results were alarming because, as indicated by the European Union Committee on Herbal Medicinal Products [49,50], estragole is carcinogenic and genotoxic.

Insights shown in Table 2 revealed that the application of particular kinds of LPGP treated water also influenced the content of components residing in essential oils in below 5% concentration. For instance, every kind of LPGP treated water completely eliminated τ -cadinol, β -eudesmol and α -cadinene. On the other hand, in essential oils isolated from basil watered with LPGP treated water, some components absent in original oil could be found. They were α -capryllene, p-cymene, β -cubebene, 4-carvomenthol and lavandulol acetate.

An explanation of the mechanisms of biosynthesis of particular components of the essential oil would require separate studies. However, one may speculate in advance on the role of particular kinds of the LPGP-treated water applied in the biosynthesis of components of essential oil.

The terpenes formed initially are subjected to further enzymatic modifications, including various modes of oxidation, reduction, isomerization and conjugation. These reactions produce several terpenoids found in that plant. The enzymes responsible for relevant transformations are not restricted to terpenoid biosynthesis. A specific hydroxylation catalyzed by the cytochrome P₄₅₀-dependent oxygenases can also be taken into account [51].

The presence of molecular oxygen in LPGPO can rationalize the elevated level of sabinene whose biosynthesis takes place mainly in the plastids. The biosynthesis of sesquiterpenes α -caryophyllene and germacrene D is restricted to the cytosol [52]. The inhibition of the α -caryophyllene and germacrene biosynthesis, and at the same time stimulation of biosynthesis of sabinene, can be rationalized involving the oxidative potential of LPGPO in plastid where the compound is formed involving monoterpene synthases [1]. On the other hand, the biosynthesis of sesquiterpenes is restricted to the cytosol. There are different mechanisms of the synthetases regulation which could be responsible for observed differences [53]. Several transformations of terpenoids (either hydroxylation or epoxidations) involved insertion of the oxygen atoms into their skeletons. They were provided by cytochrome P-450 mixed-function oxidase, e.g., oxidation of acyclic monoterpene alcohols [54] and glucosylation of diterpene alcohols by glucotransferases [55]. Supposedly, the cytochrome P450 enzymes involved in the terpenoid secondary metabolism in plants are substrate specific [56,57]. The striking difference of biosynthesis level of germacrene and β -elemene on watering basil with LPGPO can result from the stereo-specificity of appropriate hydroxylases. There is a strong similarity in three-dimensional structure of germacrene and β -elemene.

4. Conclusions

Watering basil with water treated with properly adjusted low-pressure, low-temperature glow plasma of low frequency provides a manipulation with the yield of herb crops and their functional properties. Generally, watering with water treated with glow plasma increased number of plants, height of plants, total mass of plants, total number of leaves, total mass of foliage. These parameters depended on the kind of plazmed water. Solely mass of one leave and number of leaves per plant were slightly higher in plants watered with non-treated water.

Application of particular kinds of water treated with low-temperature, low-pressure glow plasma of low frequency influenced the content of components and yield of the essential oils. Essential oil from the plant watered with non-treated water was the richest in the number of components. At the same time, watering with water treated with glow plasma under nitrogen provided the highest yield of the oil, but the poorest in terms of the number of components.

Author Contributions: W.C. run determination of separation of essential oils, D.K. gas chromatographic analyses, M.G. run basil cultivation, E.P. run basil cultivation, Z.O. equipped a research team in nanowater, P.T. invented the project, coordinated study and designed the text of this report. All authors jointly participated in interpretation of all data and in writing report. All authors have read and agreed to the published version of the manuscript.

Funding: This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflicts of Interest: No conflict of interest is known to the authors.

References

1. Ciesielska, K.; Ciesielski, W.; Girek, T.; Kołoczek, H.; Oszczyda, Z.; Tomasik, P. Reaction of *Lavandula angustifolia* Mill. to water treated with low-temperature, low-pressure glow plasma of low frequency. *Water* **2020**. accepted. [[CrossRef](#)]
2. Oszczyda, Z.; Elkin, I.; Stręk, W. Equipment for Treatment of Water with Plasma. Polish Patent PL 216025 B1, 28 February 2014.
3. Reszke, E.; Yelkin, I.; Oszczyda, Z. Plasming Lamp with Power Supply. Polish Patent PL 227530 B1, 26 October 2017.
4. Białopiotrowicz, T.; Ciesielski, W.; Domański, J.; Duskocz, M.; Fiedorowicz, M.; Graż, K.; Khachatryan, K.; Kołoczek, H.; Kozak, A.; Oszczyda, Z.; et al. Structure and physicochemical properties of water treated with low-temperature low- frequency plasma. *Curr. Phys. Chem.* **2016**, *6*, 312–320. [[CrossRef](#)]
5. Chwastowski, J.; Ciesielska, K.; Ciesielski, W.; Khachatryan, K.; Koloczek, H.; Kulawik, D.; Oszczyda, Z.; Tomasik, P.; Witczak, M. Structure and Physicochemical Properties of Water Treated under Nitrogen with Low-Temperature Glow Plasma. *Water* **2020**, *12*, 1314. [[CrossRef](#)]
6. Ciesielska, A.; Ciesielski, W.; Khachatryan, K.; Koloczek, H.; Kulawik, D.; Oszczyda, Z.; Soroka, J.A.; Tomasik, P. Structure and Physicochemical Properties of Water Treated under Carbon Dioxide with Low-Temperature Low-Pressure Glow Plasma of Low Frequency. *Water* **2020**, *12*, 1920. [[CrossRef](#)]
7. Chwastowski, J.; Ciesielski, W.; Khachatryan, K.; Kulawik, D.; Kulawik, D.; Oszczyda, Z.; Soroka, J.A.; Tomasik, P.; Witczak, M. Water of Increased Content of Molecular Oxygen. *Water* **2020**, *12*, 2488. [[CrossRef](#)]
8. Ciesielska, A.; Ciesielski, W.; Khachatryan, K.; Koloczek, H.; Kulawik, D.; Oszczyda, Z.; Soroka, J.A.; Tomasik, P. Structure and Physicochemical Properties of Water Treated under Methane with Low-Temperature Glow Plasma of Low Frequency. *Water* **2020**, *12*, 1638. [[CrossRef](#)]
9. Simon, J.E. *Basil*; Center for New Crops & Plant Products, Department of Horticulture, Purdue University: West Lafayette, IN, USA, 1995.
10. Sajjadi, S.E. Analysis of the essential oils of two cultivated basil (*Ocimum basilicum* L.) from Iran. *DARU J. Pharm. Sci.* **2006**, *14*, 128–130.
11. Telci, I.; Bayram, E.; Yılmaz, G.; Avci, B. Variability in essential oil composition of Turkish basil (*Ocimum basilicum* L.). *Biochem. Syst. Ecol.* **2006**, *34*, 489–497. [[CrossRef](#)]
12. Nowak, J. *Basil Rośliny Ozdobne*; Instytut Sadownictwa w Skierniewicach: Skierniewice, Poland, 2014; p. 5. (In Polish)

13. Andrzejewska, J.; Pisulewska, E. *Breeding of Herbal Plants*; Editorial Board of the Technical and Life Science University: Bydgoszcz, Poland, 2019. (In Polish)
14. Carroll, A.; De Persiis Vona, E.; De Persiis Vona, G. *The Dictionary of Foods: A Passionate A-to-Z Guide to the Earth's Healthy Offerings with More than 140 Delicious Nutrient Recipes*; Da Capo Press: Boston, MA, USA, 2020; ISBN 978-1-56924-395-4.
15. Grayer, R.J.; Kite, G.C.; Goldstone, E.J.; Bryan, S.E.; Paton, A.; Putievsky, E. Intraspecific taxonomy and essential oil chemotypes in Sweet basil, *Ocimum basilicum*. *Phytochemistry* **1996**, *43*, 1033–1039. [[CrossRef](#)]
16. Volpato, G.T.; Damasceno, D.C.; Calderon, I.M.P.; Rudge, M.V.C. Review of Brazilian plants with proven hypoglycemic effect in the control of diabetes mellitus. *Rev. Bras. Plant. Med.* **2002**, *4*, 35–45.
17. Jayasinghe, C.; Gotoh, N.; Aoki, A.T.; Wada, S. Phenolics Composition and Antioxidant Activity of Sweet Basil (*Ocimum basilicum* L.). *J. Agric. Food Chem.* **2003**, *51*, 4442–4449. [[CrossRef](#)] [[PubMed](#)]
18. Burt, S. Essential oils: Their antibacterial properties and potential applications in foods—A review. *Int. J. Food Microbiol.* **2004**, *94*, 223–253. [[CrossRef](#)] [[PubMed](#)]
19. Lee, S.J.; Umamo, K.; Shibamoto, T.; Lee, K.G. Identification of volatile components in basil (*Ocimum basilicum* L.) and thyme leaves (*Thymus vulgaris* L.) and their antioxidant properties. *Food Chem.* **2005**, *91*, 131–137. [[CrossRef](#)]
20. Jelacic, S.; Beatovic, D.; Prodanović, S.; Tasic, S.; Moravcevic, D.; Vujosevic, A.; Vuckovic, S. Chemical composition of the essential oil of basil (*Ocimum basilicum* L. *Lamiaceae*). *Chem. Ind.* **2011**, *65*, 465–471. [[CrossRef](#)]
21. Stanojevic, L.P.; Marjanovic-Balaban, Z.R.; Kalaba, V.D.; Stanojevic, J.S.; Cvetkovic, D.J.; Cacic, M.D. Chemical composition, antioxidant and antimicrobial activity of basil (*Ocimum basilicum* L.) essential oil. *TEOP* **2017**, *20*, 1537–1569.
22. Wongsu, P.; Chaiwarit, J.; Zamaludien, A. In vitro screening of phenolic compounds, potential inhibition against α -amylase and α -glucosidase of culinary herbs in Thailand. *Food Chem.* **2012**, *131*, 964–971. [[CrossRef](#)]
23. Gupta, S.K.; Prakash, J.; Srivastava, S. Validation of traditional claim of Tulsi *Ocimum sanctum* Linn. as a medicinal plant. *Indian J. Exp. Biol.* **2002**, *40*, 765–773.
24. Dube, S.; Upadhyay, P.D.; Tripath, S.C. Antifungal, physicochemical, and insect-repelling activity of the essential oil of *Ocimum basilicum*. *Can. J. Botany* **1989**, *67*, 2085–2087. [[CrossRef](#)]
25. Maurya, P.; Sharma, P.; Mohan, L.; Batabyal, L.; Srivastava, C. Evaluation of the toxicity of different phytoextracts of *Ocimum basilicum* against *Anopheles stephensi* and *Culex quinquefasciatus*. *J. Asia-Pac. Entomol.* **2009**, *12*, 113–115. [[CrossRef](#)]
26. Grayer, R.J. Basil. The genus *Ocimum*. *Phytochemistry* **2001**, *58*, 533. [[CrossRef](#)]
27. Lupton, D.; Khan, M.M.; Al-Yahai, R.A.; Hanif, M.A. *Basil (in) Leafy Medicinal Herbs, Chemistry, Technology and Uses*; Chapter 3; Ambrose, D.C.P., Manickavasagan, A., Naik, R., Eds.; CABI: Boston, MA, USA, 2016; ISBN 9781780645599.
28. Duncan, D.B. Multiple Range and Multiple F Tests. *Biometrics* **1955**, *11*, 1–42. [[CrossRef](#)]
29. Nishida, R.; Tan, K.H. Methyl eugenol: Its occurrence, distribution and role in nature, especially in relation to insect behaviour and pollination. *J. Insect Sci.* **2012**, *12*, 1–60.
30. Federal Register, Regulations: Synthetic Flavouring Agents and Adjuvants. 2018. Available online: <https://www.govinfo.gov/app/details/FR-2018-10-09/2018-21807> (accessed on 9 October 2020).
31. European Community. Regulation No. 1272/2008. Available online: <https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A32008R1272> (accessed on 5 September 2020).
32. Govindarajan, M.; Sivakumar, R.; Rajeswary, M.; Yagalakshmi, K. Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae). *Exp. Parasitol.* **2013**, *134*, 7–11. [[CrossRef](#)] [[PubMed](#)]
33. Ung, C.Y.; White, J.M.L.; White, I.; Banerjee, P.; McFadden, J.P. Patch testing with the European baseline series fragrance markers: A 2016 update. *Br. J. Dermatol.* **2018**, *178*, 776–780. [[CrossRef](#)]
34. Gu, Y.; Ting, Z.; Qiu, X.; Zhang, X.; Gan, X.; Fang, Y.; Xu, X.; Xu, R. Linalool preferentially induces robust apoptosis of a variety of leukemia cells via upregulating p53 and cyclin-dependent kinase inhibitors. *Toxicology* **2010**, *268*, 19–24. [[CrossRef](#)]
35. Chang, M.-Y.; Shieh, D.-E.; Chen, C.-C.; Yeh, C.-S.; Dong, H.-P. Linalool Induces Cell Cycle Arrest and Apoptosis in Leukemia Cells and Cervical Cancer Cells through CDKIs. *Int. J. Mol. Sci.* **2015**, *16*, 28169–28179. [[CrossRef](#)] [[PubMed](#)]

36. Iwasaki, K.; Zheng, Y.-W.; Murata, S.; Ito, H.; Nakayama, K.; Kurokawa, T.; Sano, N.; Nowatari, T.; Villareal, M.O.; Nagano, Y.N.; et al. Anticancer effect of linalool via cancer-specific hydroxyl radical generation in human colon cancer. *World J. Gastroenterol.* **2016**, *22*, 9765. [[CrossRef](#)] [[PubMed](#)]
37. Nikonorow, M.; UrbaneK-Karłowska, B. *Food Toxicology*; PZWI: Warsaw, Poland, 1987. (In Polish)
38. Tognolini, M.; Barocelli, E.; Ballabeni, V.; Bruni, R.; Bianchi, A.; Chiavarini, M.; Impicciatore, M. Comparative screening of plant essential oils: Phenylpropanoid moiety as basic core for antiplatelet activity. *Life Sci.* **2006**, *78*, 1419–1432. [[CrossRef](#)] [[PubMed](#)]
39. Klocke, J.A.; Darlington, M.V.; Balandrin, M.F. 1,8-Cineole (Eucalyptol), a mosquito feeding and ovipositional repellent from volatile oil of *Hemizonia fitchii* (Asteraceae). *J. Chem. Ecol.* **1987**, *13*, 2131–2141. [[CrossRef](#)]
40. Sfara, V.; Zerba, E.N.; Alzogaray, R.A. Fumigant insecticidal activity and repellent effect of five essential oils and seven monoterpenes on first-instar nymphs of *Rhodnius prolixus*. *J. Med. Entomol.* **2009**, *46*, 511–515. [[CrossRef](#)]
41. Schiestl, F.P.; Roubik, D.W. Odor compound detection in male euglossine bees. *J. Chem. Ecol.* **2003**, *29*, 253–257. [[CrossRef](#)]
42. Schemske, U.W.; Lande, R. Fragrance collection and territorial display by male orchid bees. *Anim. Behav.* **1984**, *32*, 935–937. [[CrossRef](#)]
43. Material Safety Data Sheet—Cineole. *ScienceLab*, 2012. Available online: <https://web.archive.org/web/20120829230700/http://www.sciencelab.com/msds.php?msdsId=9924005> (accessed on 9 July 2020).
44. Caceres, A.I.; Liu, B.; Jabba, S.V.; Achanta, S.; Morris, J.B.; Jordt, S.-E. Transient Receptor Potential Cation Channel Subfamily M Member 8 channels mediate the anti-inflammatory effects of eucalyptol. *Br. J. Pharmacol.* **2017**, *174*, 867–879. [[CrossRef](#)]
45. Ali, P.; Chen, Y.-F.; Sargsyan, E. Bioactive Molecules of Herbal Extracts with Anti-Infective and Wound Healing Properties. In *Microbiology for Surgical Infections*; Academic Press: Cambridge, MA, USA, 2014; pp. 205–220.
46. Arunkumar, R.; Nair, S.A.; Rameshkumar, K.B.; Subramoniam, A. The essential oil constituents of *Zornia diphylla* (L.) Pers, and anti-inflammatory and antimicrobial activities of the oil. *Rec. Nat. Prod.* **2014**, *8*, 385–393.
47. Šobotník, J.; Hanus, R.; Kalinová, B.; Piskorski, R.; Cvačka, J.; Bourguignon, T.; Roisin, Y. (E,E)- α -farnesene, an alarm pheromone of the termite *Prorhinotermes canalifrons*. *J. Chem. Ecol.* **2008**, *34*, 478–486. [[CrossRef](#)]
48. Hern, A.; Dorn, S. Sexual dimorphism in the olfactory orientation of adult *Cydia pomonella* in response to α -farnesene. *Entomol. Exp. Appl.* **1999**, *92*, 63–72. [[CrossRef](#)]
49. Committee on Herbal Medicinal Products. Final Public Statement on the Use of Herbal Medicinal Products Containing Estragole, EMEA/HMPC/137212/2005. Available online: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2010/04/WC500089960.pdf (accessed on 19 September 2020).
50. SCF/CS/FLAV/FLAVOUR/6. Opinion of the Scientific Committee on Food on Estragole (1-Allyl-4-methoxybenzene); Scientific Committee on Food: Brussels, Belgium, 26 September 2001. Available online: https://web.archive.org/web/20070302102822/http://ec.europa.eu/food/fs/sc/scf/out104_en.pdf (accessed on 19 September 2020).
51. Hallahan, D.L.; West, J.M.; Smiley, D.W.M.; Picket, J.A. *Nepetalactol oxireductase* in trichomes of the catmint *Nepeta racemosa*. *Phytochemistry* **1998**, *48*, 421–427. [[CrossRef](#)]
52. Gleizes, M.; Pauly, G.; Carde, J.-P.; Marpeau, A.; Bernard-Dagan, C. Monoterpene hydrocarbon biosynthesis by isolated leucoplasts of *Citrofortunella mitis*. *Planta* **1983**, *159*, 373–381. [[CrossRef](#)]
53. Back, K.; Chappeli, J. Cloning and bacterial expression of a sesquiterpene cyclases from *Hyoscyamus muticus* and its molecular comparison to related terpene cyclases. *J. Biol. Chem.* **1995**, *270*, 7375–7381. [[CrossRef](#)]
54. Keller, Y.; Bouvier, F.; Dharlingue, A.; Camara, B. Metabolic compartmentation of plastid prenyl lipid biosynthesis: Evidence for the involvement of a multifunctional geranylgeranyl reductase. *Eur. J. Biochem.* **1998**, *251*, 413–417. [[CrossRef](#)]
55. Shibata, H.; Sawa, Y.; Oka, T.; Sonoke, S.; Kim, K.K. Steviol and steviol—Glycoside: Glucotransferase activities in *Stevia rebaudiana* Bertoni—Purification and partial characterization. *Arch. Biochem. Biophys.* **1995**, *321*, 390–396. [[CrossRef](#)] [[PubMed](#)]
56. West, C.A. Biosynthesis of Diterpenes. In *Biosynthesis of Isoprenoid Compounds*; Porter, J.W., Spurgeon, S.L., Eds.; John Wiley and Sons: New York, NY, USA, 1981; pp. 376–411.

57. Mihaliak, C.A.; Karp, F.; Croteau, R. Cytochrome P450 Terpene Hydroxylases. In *Enzymes of Secondary Metabolism*; Lea, P.J., Ed.; Academic Press: London, UK, 1993; pp. 261–279.

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).