Supplementary Material

Effects of In Vitro Digestion on the Content and Biological Activity of Polyphenols from Acacia mearnsii Bark

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Abstract: The stability and bioaccessibility of polyphenol from Acacia mearnsii bark were measured at various stages during in vitro simulated digestion. Subsequently, the changes in the total polyphenol content (TPC) and biological activity were studied. The results showed that the phenolic compounds from A. mearnsii remained stable, and TPC underwent few changes during gastric digestion. Nonetheless, intestinal digestion led to the degradation of proanthocyanidins (PAs) and a significant decrease in TPC (26%). Degradation was determined by normal-phase HPLC and gel permeation chromatography. Only monomers, dimers, and trimers of flavan-3-ols were identified in the serum-accessible fraction for characterization of their bioaccessibility. The results also indicated the obvious antioxidant capacity of PAs from A. mearnsii bark, and ~53% of the α-glucosidase–inhibitory effect was preserved. All these findings show that PAs from A. mearnsii bark as a native plant source may be particularly beneficial for human health as a natural nutritional supplement.

Keywords: Acacia mearnsii; proanthocyanidins; in vitro digestion; antioxidant; antidiabetic; HPLC/MS

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Table S1. Total polyphenol contents (TPC) of samples from A. mearnsii bark at different times during the simulated gastric-intestinal digestion without enzymes.

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Figure S2. Product ion scan (MS and MS²) of the m/z 305 in the IN sample.

Figure S3. Product ion scan (MS and MS²) of the m/z 561 in the IN sample.

Figure S4. Product ion scan (MS and MS²) of the m/z 577 in the IN sample.

Figure S5. Product ion scan (MS and MS²) of the m/z 593 in the IN sample.

Figure S6. Product ion scan (MS and MS²) of the m/z 833 in the IN sample.
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**Figure S8.** Product ion scan (MS and MS²) of the m/z 865 in the IN sample.

**Figure S9.** Product ion scan (MS and MS²) of the m/z 881 in the IN sample.

**Table S1.** Total polyphenol contents (TPC) of samples from *A. mearnsii* bark at different times during the simulated gastric-intestinal digestion without enzymes.

<table>
<thead>
<tr>
<th>Digestion Time (min)</th>
<th>TPC of Digestion Samples (mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Gastric Digestion</td>
</tr>
<tr>
<td>0</td>
<td>5.97 ± 0.05</td>
</tr>
<tr>
<td>30</td>
<td>5.83 ± 0.06</td>
</tr>
<tr>
<td>60</td>
<td>5.88 ± 0.04</td>
</tr>
<tr>
<td>90</td>
<td>5.91 ± 0.03</td>
</tr>
<tr>
<td>120</td>
<td>5.89 ± 0.04</td>
</tr>
</tbody>
</table>

The IN sample represents the solution that diffused into the dialysis tubing, the OUT sample represents the solution outside of the tubing. Data represent the means of three independent determinations ± SD.

**Figure S1.** Product ion scan (MS and MS²) of the m/z 289 in the IN sample.
Figure S2. Product ion scan (MS and MS²) of the m/z 305 fragment in the IN sample.
**Figure S3.** Product ion scan (MS and MS²) of the m/z 561 in the IN sample.

**Figure S4.** Product ion scan (MS and MS²) of the m/z 577 in the IN sample.
Figure S5. Product ion scan (MS and MS²) of the m/z 593 in the IN sample.

Figure S6. Product ion scan (MS and MS²) of the m/z 833 in the IN sample.
**Figure S7.** Product ion scan (MS and MS^2) of the m/z 849 in the IN sample.
Figure S8. Product ion scan (MS and MS²) of the m/z 865 in the IN sample.

Figure S9. Product ion scan (MS and MS²) of the m/z 881 in the IN sample.